

Table 12
Transduction assay

76

Used typed antibodies

Significance

Rx	cells	lysis	cell type	idiotypic	allotypic	polyclonal	total
S	1		+	33	0	0	33
+	1		+	16	0	0	16
S	2		+	20	0	0	20
+	2		+	15	0	0	15
				46	0	0	46
S	4		+	20	0	0	20
+	4		+	6	1	0	7
S	1		x 3 (1) 1210 lysate	1	0	0	1
S	1		x 4 (2) 902 lysate	1	0	0	1
+	1		x 5 (1) 902 lysate	36	6	0	42
			x 6 (2) (1210) lysate	18	3	0	21
S	2		1	20	0	0	20
			4	21	1	1	23
(1), (8), (10) = cult. 2175							
(2), (7), (9) = cult. 1210							
+	2		x 7 (2) 1210 (cult)	19	2	0	21
			x 8 (2) 2175 (cult)	14	3	2	19
(3), (6), (11) = lysate 1210							
(4), (5), (12), (13), (14) = lys. 902							
+	2		x 9 (1) 1210 (cult)	22	1	0	23
W			x 10 (2) 2175 (cult)	9	7	0	16
S	4		x 11 (4) 1210 lysate	17	2	0	19
			x 12 (2) 902 (lysate)	35	5	1	41
+	4		x 13 (2) (902) (lysate)	16	3	0	19
R	4		x 14 (902) (lysate)	15	3	0	18
							450

Segregant Analysis via hyaline actin
in known cultures.

(22)

Segregant	bp. Genotype	Transd. Hyaline	Segregant			Total		
			Ident. Hyaline	Acetogenic	Ident. Acetogenic			
						5		
Gal ₁ ⁻	+	+	5	0		4		
Gal ₂ ⁻	+ √(1) 2175	+	4	0		4		
		(2) 1210	4	0		4		
Gal ₄ ⁻	s	+	4	0		4		
		+	4	0		4		
Gal ₁ ⁻	+	Gal ₂ ⁻ √(3) 902	4	5		9		
			(4) 1210	0	3	3		
Gal ₂ ⁻	s	Gal ₄ ⁻	0	1		1		
			0	2		2		
			+ √(5) 2175	Gal ₁ ⁻	0	0		4
			+ √(6) 2175	Gal ₄ ⁻	4	1		5
		(7) 1210	0					
Gal ₄ ⁻	s	Gal ₂ ⁻ √(8) (w 902)	16	3		19		
			(9) 1210	0	1	1		
			+ √(10) 902	15	3		18	
						79		

Segregants from table 12 were classified as ~~pure~~ ~~hyaline~~ ~~acetogenic~~ ~~by means of their action on~~ ~~known cultures.~~
 confirmed by the ^{hyaline} ~~action~~ of ~~these~~ ~~segregants~~, classified by means of their action on known cultures.

Table 13 ~~with some corrections~~ ~~confirmatory~~ ~~Segregants from table 12~~ ~~classified by means~~ ~~of their action on~~ ~~known cultures.~~

Table 14

78

Galactose negative cultures giving
high frequency of raw ductus
cystae

Culture	Galactose isotaxis	Ramp Cells	Trand lyate	Nature of Gal(+) mutation	NFT sequant	Nature of Gal(+) revertin NFT seq.
230-4	(+) Gal ₁ ⁻	Gal ₁ ⁻	Gal ₂ ⁻	unstable	Gal ₁ ⁻	stable
246A-15	(+) Gal ₁ ⁻	Gal ₁ ⁻ Gal ₂ ⁻	Gal ₁ ⁻ (+) Gal ₂ ⁻	"	Gal ₁ ⁻ Gal ₂ ⁻ Gal ₁ ⁻ Gal ₂ ⁻	"
241-14	(+) Gal ₂ ⁻	Gal ₁ ⁻	Gal ₂ ⁻	"	Gal ₂ ⁻	stable
241-19	(+) Gal ₂ ⁻	Gal ₁ ⁻	Gal ₂ ⁻	"	Gal ₁ ⁻ Gal ₂ ⁻	none observed
1928-16	(+) Gal ₂ ⁻	Gal ₁ ⁻	Gal ₂ ⁻	"	Gal ₁ ⁻ Gal ₂ ⁻	unobserved (2346)
257-2	(+) Gal ₂ ⁻	Gal ₁ ⁻	Gal ₂ ⁻	unstable	Gal ₂ ⁻	stable
257-4	(+) Gal ₂ ⁻	Gal ₁ ⁻	Gal ₂ ⁻	unstable	Gal ₂ ⁻	stable
153-1	(-) Gal ₂ ⁻	Gal ₄ ⁻	Gal ₂ ⁻	unstable	Gal ₂ ⁻	—
153-4	(-) Gal ₂ ⁻	Gal ₄ ⁻	Gal ₂ ⁻	"	Gal ₂ ⁻	stable
202-10	(+) Gal ₄ ⁻	Gal ₄ ⁻	Gal ₂ ⁻	"	Gal ₂ ⁻	stable
202-18	(+) Gal ₄ ⁻	Gal ₄ ⁻	Gal ₂ ⁻	unstable	Gal ₁ ⁻ Gal ₂ ⁻	stable
2478-1	(+) Gal ₄ ⁻	Gal ₄ ⁻	Gal ₂ ⁻	"	Gal ₁ ⁻ Gal ₂ ⁻	"

GALACTOSE NEGATIVE CULTURES GIVING HIGH FREQUENCY OF RAW DUCTUS CYSTAE
 GALACTOSE NEGATIVE CULTURES GIVING HIGH FREQUENCY OF RAW DUCTUS CYSTAE

FREQUENCY OF ALTERNATION MUTATED
 GALACTOSE NEGATIVE CULTURES GIVING HIGH FREQUENCY OF RAW DUCTUS CYSTAE

267

270

Phage

Table 16

Gal₁⁻ Gal₄⁻ Interaction

79

282

I The transductions

Gal₄⁻ Lp^d cells exposed to HFT
gal₁⁻

~~gal₁⁻~~
(+) colonies

(-) colonies

Colonies
seen after
2 days

lysate treated
control

0

408

2

0

440

0

II The propagating colonies gave mixed (+), (-) and propagating colonies

(A) streaked out 24 pure (+) colonies picked and streaked out

(B) of the 24 colonies - 6 were found stable gal(+)

(C) of the 18 apparently (-) colonies (picked at 24 hours) derived from pure (+) tested against HFT 1⁻ and HFT 4⁻

Gal ₄ ⁻	Gal ₁ ⁻	Gal ₁ ⁻ Gal ₄ ⁻	Parental (-) propagating	Doubtful results
6	5	2	4	+

all colonies lambda resistant

leave
out

285 I the transductions

Gal₁⁻ Lp^d cells

treated with HFT gal₄⁻

(+)

(-)

prop. colonies

partially
lysed

(A) Control ~~plating~~

0

465

0

0

(B) lysate ~~plating~~

0

316

2

38

II (A) the propagating colonies streaked out - Each gave (+), (-), prop (-)

(B) Colony 1 - 24 gal(+) not streaked out - 11 were stable gal(+)

(-) tested against HFT 1⁻ and HFT 4⁻ are lambda prot.

Gal ₄ ⁻	Gal ₄ ⁻	Gal ₁ ⁻ Gal ₄ ⁻	propagating (-) Parental
10	2	0	1

(C) Colony 2 - 48 pure (+) picked and streaked out - 23 (?) were stable (+)

(-) tested against HFT 1⁻ and HFT 4⁻

Gal ₁ ⁻	Gal ₄ ⁻	Gal ₁ ⁻ Gal ₄ ⁻	Propagating (-)
+	*	0	22

Table 16

and - are retained for closer study.

The gal⁻ - gal⁺ interaction

About 650 cultures have been tested in this way, each from a separate

1. The heavy duobios

Requirements: Transduced
 Cells Hyal⁻ Gal (+) Gal (-) Papillating Gal -

Gal⁻ ~~with~~ ~~HFT Gal⁻~~ 0 440 0
 HFT Gal⁻ 0 408 2

Gal⁻ with 0 415 0
 HFT Gal⁻ 0 316 2

2. Classification of galactose negative segregants from galactose positive clones found in papillating galactose negative colonies

Classification of segregants
 Cells Transduced Hyal⁻ Gal⁻ Gal⁺ Gal⁻ Gal⁺ Papillating Gal -

Gal⁻ HFT Gal⁻ 6 5 2
 Gal⁻ HFT Gal⁻ 10 0

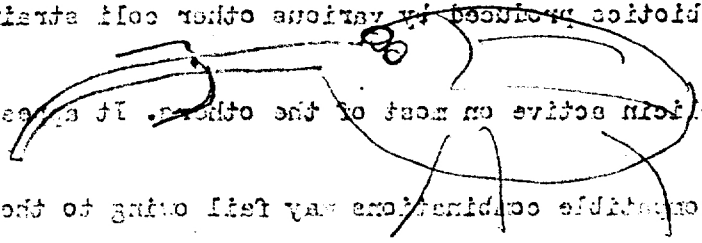
and of course, and particularly in patterns of sensitivity to phages, including

and to antibiotics produced by various other coli strains, colonies. W-3

produces a colicin active on most of the other. It is very likely that many

potentially compatible conditions may fail owing to the suppression of W-1

by a colicin produced by the other parent.



81

Table 17

~~These authors have not reported the results of the following experiments~~

Parents: SSA and Prototrophic Recombinants
The great majority of the prototrophs were Gal⁻ and Galactose (+)

a very small number of the other combinations have been found also.
In one competition of the Salmonella and E. coli systems, Davis (1970)
Gal⁺ 1 Gal⁻ 2
541** 99
In a competition experiment was duplicated. A U-tube with a sintered glass diaphragm

in the cross-limb, the two compartments were flushed with prot and inoculated with SSA and SSA respectively.
* unstable for galactose, galactose negative, galactose positive
From time to time the prot was flushed back and forth between the compartments
** 25 of 25 examined were unstable for galactose fermentation. Segments, by alternating suction. The prot was saturated, the cells from each compartment from each of the 25, were all gal⁻.
The results were repeated on animal agar. It was repeatedly found that prototrophs appeared from the SSA culture, but not from the SA culture.
109/57
115/13

Control experiments in which only one compartment was inoculated verified

the integrity of the filter.

This experiment appeared to show that a filtrable agent (FA) was produced by SA that reacted with SSA to produce prototrophs. However, filtrates prepared directly from SA were inactive. The paradox was resolved when it was found that the addition of a SSA filtrate, or of a lysate of SA cultivated from a lysogenic phase secreted by SSA, provoked the formation of FA by SA.

FA, then, is not a normal component of SA, but is produced under the stimulus of a lysogenic phase secreted by SSA, provoked the formation of FA by SA.

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destroy FA activity.

<u>page</u>	<u>lyate</u>	<u>Ass of cells</u>	<u>Needs</u>	<u>Time lyate</u>
<u>176b</u>	04E750	811, 2050, 750, 2175,	$(42/43)$	$(420/16^+)$ $(2/2)$ $(12/24)$ —
				repeated

(89)

<u>177</u>	8421	578	4% of cell (+)	—
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<u>177b</u>	lyate sterility			—
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<u>201</u>	217E750	811,	solid swan	—
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Summ. sheet 214 before - page 578+14/2 etc not order

W2070 (= gal₆ - ?) derived from W1673 by uv.
Lp^s

Page

135a - origin (three gal- obtained, one retained) (#1)

136 - found by (+) no add = 7
412(25) = 1560

137 - pop. check - 7/5 pentamer (+) stbl.
7/1 by rate (1412) (+) unstbl.

145 - trsd. test no add 10
750 λ 54 * — 4/5 (+) unstbl
902 λ 1256 — 5/5 (+) unstbl
84 λ 175 — 0/5 (+) unstbl.

295 - trsd. by π8, π9 λ to recover (-) of the T.
lysates not active Tit. 5/1

π9 = 7/2
apparently, segments found
of 16 examined 15 2-
1 1-2-
16

→ Apparent (+) unstbl
segments better found.
5 1-
2 2-
7

226 Stocks of 2070 λ + made

Stability of transducers by revision of sales

Age Observation

134	STAT 8U ₈ + 5	7/8 stable	$\frac{\text{transducer}}{\text{control}} = \frac{3532}{30}$
135a	STAT 8U ₈ + 8	8/8 stable	$\frac{\text{transducer}}{\text{control}} = \frac{291}{25}$

Correlation of Lip Charge & Transduction

236A 2281 + K12

236D 2281 + 750

Pap	LP ^{RK} 2281	Seq	λ	Pap	LP ^{RK} 1445	Seq	λ
1.	+	2-	R	19	+	2-	R
2.	+	"	R	1	S or R	2-	S
3.	+	"	R	2			
4.	+	"	R				
5.	+	"	R				
6.	+	"	R				
7.	+	"	R				
8.	+	"	R				
9.	+	"	R				
10.	+	"	R				
11.	+	"	R				
12.	R or S		S				

The transformed cells
 Examination of the other characteristics of the cells transformed
 was in regard to growth characteristics. It was found that the
 ability to ferment galactose has uniformly shown no changes in any
 of them with the exception of the mutation of transducibility in the mutants
 examined. Changes in the amount of galactose fermented by some of
 the mutants were noted. It was found that the mutants which fermented
 galactose and sporulated for 48 hours.

P Cross and details

100 of seg. 811K12 x 1436 73 (+) 14 (-)
 ↑
 at least 20/F3 showed mosaic colonies on stk:1
 all later shown unstable

518K12 x 1436 86 (+) 1840 (-) ✓
 ↑
 5/20 showed mosaics on stk-1
 all " were mixed on stk 2

106 518K12 x 1436 0 (+) 65 (-) ✓

84K12 x 1436 27 (+) 2 (-)
 ↑
 shown unstable (107)

Total	
518K12	86 (+) 1905 (-)
84K12	100 (+) 16 (-)
1924K12	27 (+) 1944

112 1924K12 x 1436 16 (+) 952 (-)
 ↑
 11 (+)
 27 (+) 992
 1944

shown not to be lysogenic in cmo broth.

may be higher medium not good

Rpt 286(?) 3+/1(-) active with non aerated cult. 518 ← was this ... checked 518 1436?
 26 found non lys on ...
 15 ...
 - 30 prob picked at random found non lys.

279 902 met +⁺ 518 F⁻
 ① 2580K12 x 1321 F⁺ F⁻ → (1+) / c. 6000 (-) → streaked out. - +
 6 seg taken = gal₂⁻

② 2580K12 x 518 F⁻ F⁺ → 541 (+) / 99 (-) → 30 picked 4 → stb⁺
 25 seg all gal₂⁻

Table 17

HFT - histriol - Why + " not found HFT ?

(GA)

Page	lyrate	Array Cells	Reactive	lyrate titer	
125	750E1821	811	2470 pap/ha	6.5 x 10 ⁹	
	"	578	9630+ / ml	"	$\frac{10^4+ / ml}{10^{10} \lambda / ml} = \frac{1+}{10^6 \lambda}$
date 10/9/52					
142	518E892-1	518	no pop. less than control	?	<p>← lyrate found not sterile</p> <p>← lyrate found sterile</p>
11/26/52	811E892	"	no pop. at all	?	
	518E892-1	2050	solid smear	?	
	811E892-1	"	" "	?	
143	518E892-1	2050	solid smear	?	<p>$\approx 1.8 \times 10^9 \lambda / ml$</p> <p>all lyrate titers sterile</p> <p>↳ penicillin (10 mc)</p> <p>control. 200 200 (very high)</p> <p>small papillae - spreader control?</p>
	518E892-2	"	solid "	?	
	811E892-1	"	solid "	?	
	811E892-2	"	188 pap.	?	
	518E892-1	2050	solid smear	?	
157	518E892 → (-) 200 811E892-1	"	" "	?	
	518E892-1	"	" "	?	
	1436E1412-1	"	227/22	?	
161	excluded by contamination				
165, a, b	518E892-1 (and dilutions)	750, 518, 2050, 2175	solid smear.	1.8 x 10 ⁹ (p 166)	
	01 (518E892 → 500, 800)	518, 750, 2050	" "	" "	
		(518 2175)	— not found.		
168	D1 E750 D4 E750	skated as unstable		—	—
169	D4 E750	titer	> 10 ¹⁰	—	—
170 b	D4 E750 (1-10)	811E892	24/67	↑	

HFT
NFT Summary U^+

	Recipient cells	lyate source	Test		
			<u>1</u>	<u>2</u>	<u>4</u>
	gal_2^- (12375)	gal_1^- (12375)			
①	gal_1^- ①	+	+++	++	+++
②	" ②	+	+++	++	+++
③	Gal_1^- (hp ^d)	+	+++	100	+++
④	gal_1^-	gal_2^-	+++	+	-
<u>Others</u>					
⑤	Gal_2^- ①	Gal_1^-	++	200	+++
	②		○	○	2
⑥	Gal_1^-	Gal_2^-	+++	+++	+++

?

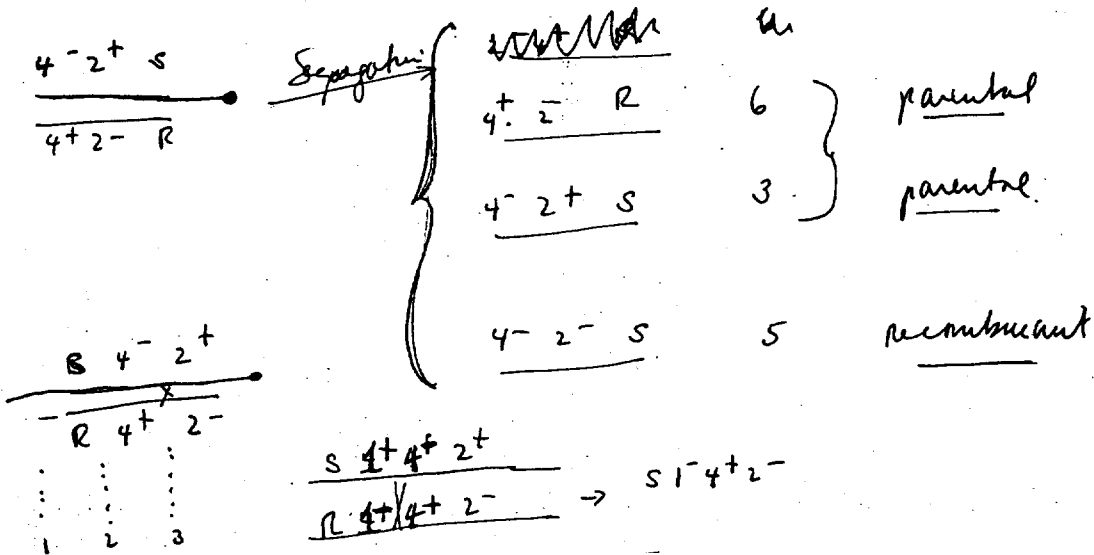
Culture	Recipient cells	lyate	Transd genotype	HFT segregant	mutable des	NFT secondary segregant	Presumed structure of HFT
2342	gal_2^-	gal_1^-	$\frac{2^- 1^+}{2^+ 1^-}$	2^-	$1\frac{1}{2}$	$1^- 2^-$	$\frac{2^- 1^-}{2^+ 1^+}$
2346	gal_1^-	gal_2^-	$\frac{2^+ 1^-}{2^- 1^+}$	1^-	$\frac{4}{5}$	1^- (8% mv. stable)	$\frac{2^+ 1^-}{2^+ 1^-}$
241-14	gal_1^-	gal_2^-	$\frac{2^+ 1^-}{2^- 1^+}$	2	$1\frac{1}{2}$	2 ($1\frac{1}{2}$ stable)	$\frac{2^- 1^+}{2^- 1^+}$
241-19	"	"	"	"	$1\frac{1}{2}$	$1^- 2^-$ (-)	$\frac{2^- 1^-}{2^+ 1^+}$
246A-15	$gal_1^- gal_2^-$	$\left\{ \begin{matrix} gal_1^- \\ gal_2^- \end{matrix} \right\}$	$\frac{1^- 2^-}{1^+ 2^-} \rightarrow \frac{1^- 2^-}{2^- 2^+}$	2^-	-	$1^- 2^-$	$\frac{2^- 1^-}{2^- 1^+}$

Segregation from $4^- 2^+ 4p^s$ transformed cell

SIFT N6

89

P 262



① Relationship of transduction + lysogenization.

→ diploids. h_p^+ / l_p^+ from $+/s$; s/s .

SM

Mated λ ; h_p^+ .

② Hft basis: construct $\frac{s}{s}$ i UV'd phage. defect λ

S.H. (C)

③ Position effect (S)

④ Association of fragment + chromosome. → diploids; crossing behavior

SM

SM

of various λ transduction types. Size of fragment. Behavior from $\frac{+-}{==+}$. Crossover + segregation mechanisms.

⑤ other transducible loci; other phages.

⑥ Cytology of λ , h_p^+ Hft.

⑦ lytic λ ! (Especially when grown on $\frac{s}{s}$ types!)

How?

⑧ How many λ types; mapping (sep. 12)

More 4/10/54

(91)

Table 8: Study absorption with multiplicity < 1 . Heated cells? Hft
of h_2^s / h_2^r .

Table 9: Any h_1^s Clarify headings. Discuss Gal, $-x$ Gal
behavior.

Table 11 Explain Obs column

12 Total: homo/heterotype + test homogeneity.

Hft: inductive behavior! (basis now studied)

Table 18. Again verify Gal types. p.12 P.2: meaning?

Double $-$. Papillae \hat{c} mixed phage $\approx c^2$?

Fig 2. Or UV improves survival. Effect of excess UV'd incamp. Δ
Variance in output of Hft.

$\left. \begin{array}{l} 24 \rightarrow x 1 \\ 12 \rightarrow x 4 \text{ should fail} \\ 14 \rightarrow x 2 \text{ should be OK} \end{array} \right\}$

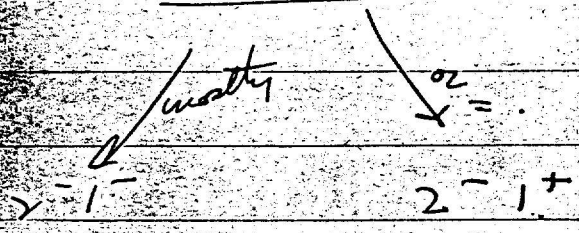
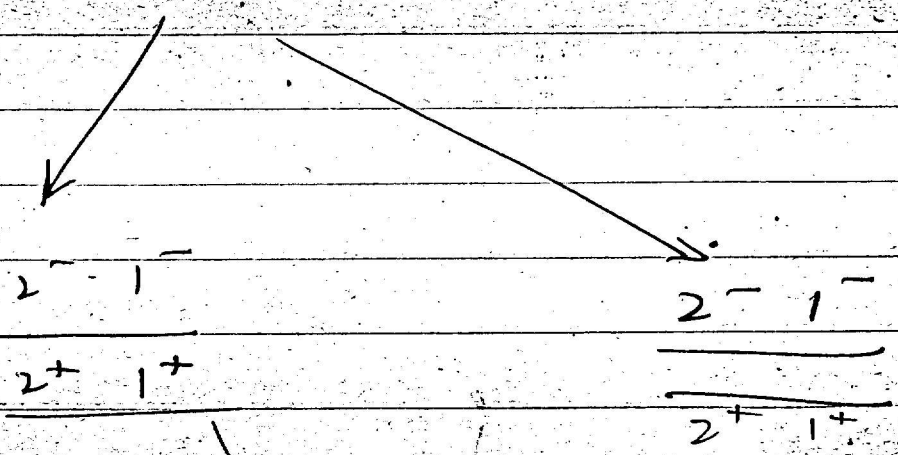
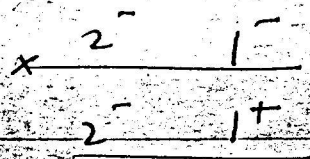
 102
 102

1.2. Behavior of $\dots \rightarrow x \dots$

Detail?

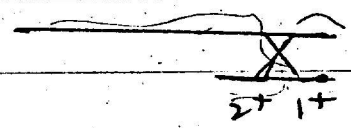
13. ? Are exchange + segregation independent? Many "segregations" may be automatic.

What's up with \dots ?



are the 2^- segments now ~~para~~ hemizygous?

Suppose fragment is terminal.



Only 1 crossover type feasible! I.E. $1^- 2^+$ recombinant would be

a fragment. Why no recessions of type $\frac{2^+ 1^-}{2^- 1^+}$? These

should give mostly the $2^+ 1^-$ type. Were enough tested?

Effect of Ultraviolet
on Gal₂ HFT λ
M 210, 216a

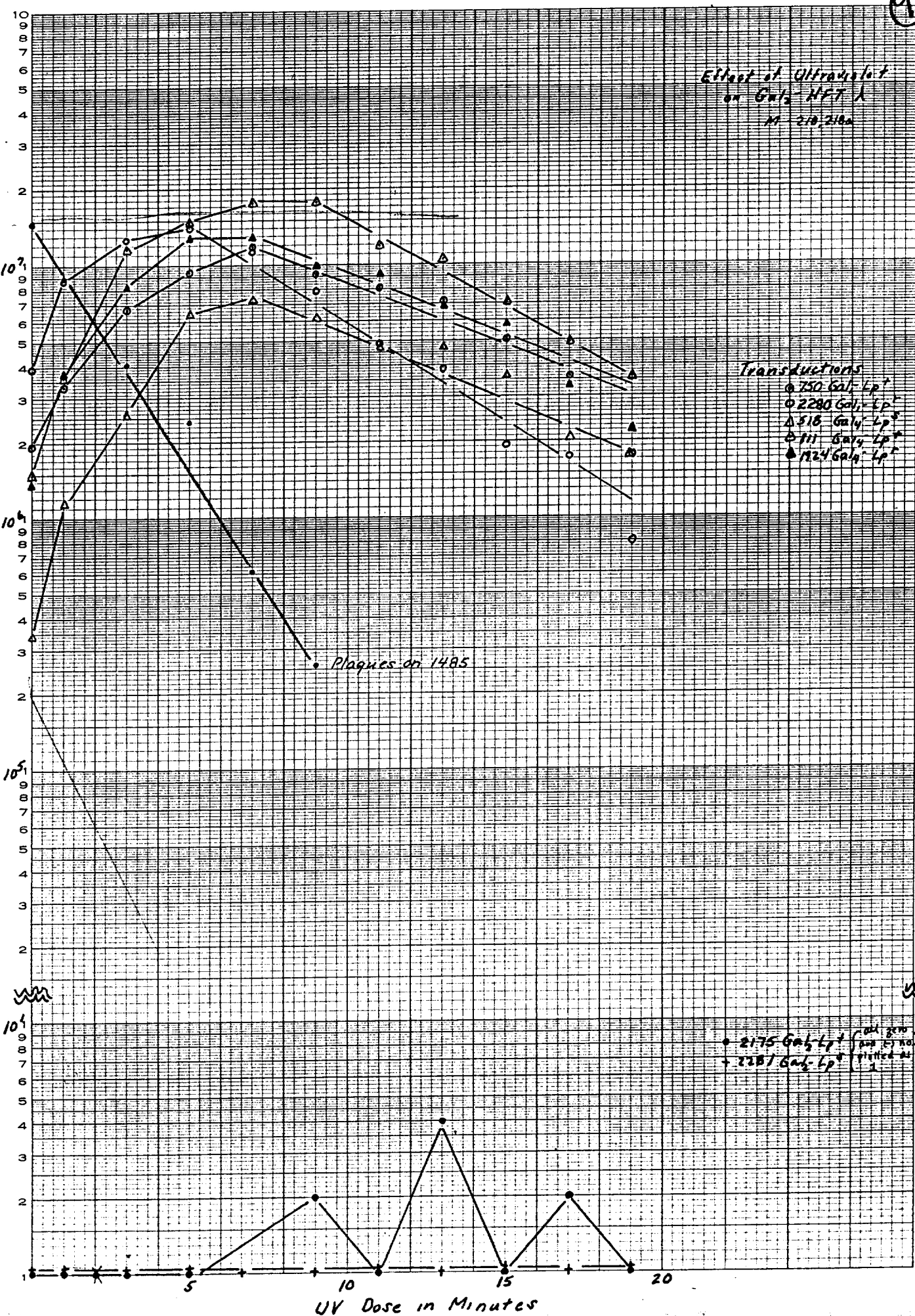
Number Per Ml. (Irradiation Tube)

Transductions
○ 750 Gal₂ Lp⁺
□ 2280 Gal₂ Lp⁺
△ 518 Gal₂ Lp⁺
◇ 711 Gal₂ Lp⁺
▲ 1224 Gal₂ Lp⁺

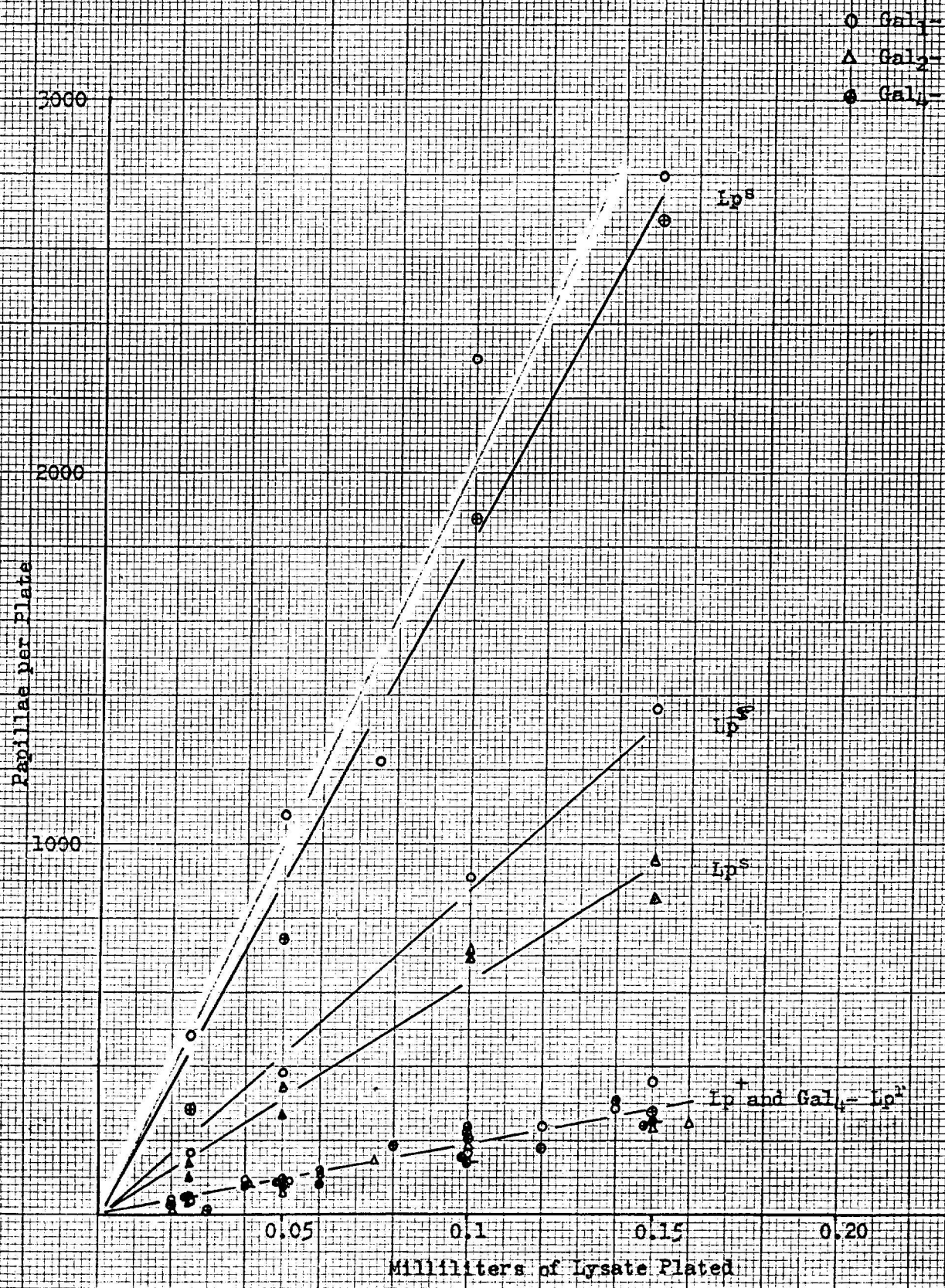
Plaques on 1485

• 2175 Gal₂ Lp⁺ (all zero)
+ 2281 Gal₂ Lp⁺ (all zero)

NO. 340-LS12 DIETZGEN GRAPH PAPER
SEMI-LOGARITHMIC
5 CYCLES X 12 DIVISIONS PER INCH
EUGENE DIETZGEN CO.
MADE IN U. S. A.



UV Dose in Minutes



96

Action of K-rays
on transducing
and plaque activity
in hyaline

EUGENE DIETZGEN CO.
MADE IN U. S. A.

NO. 340-LS12 DIETZGEN GRAPH PAPER
SEMI-LOGARITHMIC
5 CYCLES X 12 DIVISIONS PER INCH

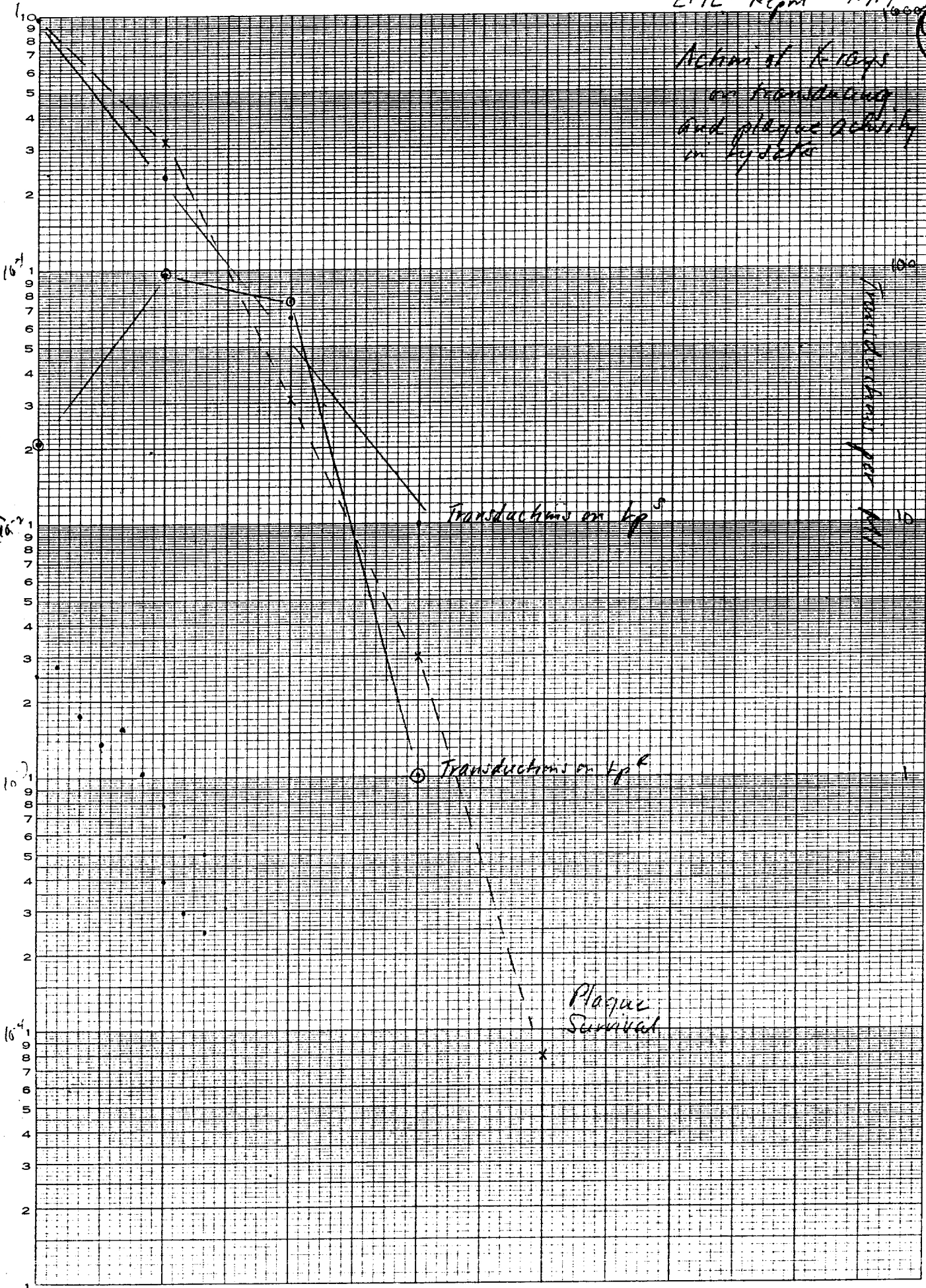
Surviving Fraction

Transductions per cell

Transductions on 10^5

Transductions on 10^6

Plaque Survival



Dose $\times 10^2$

Table 1

Principal cultures

Wisconsin Stock Number

Genotype

W518	F ⁺ M ⁻ Lac ₁ ⁻ Gal ₁ ⁻ Lp ^S
W750	F ⁺ M ⁻ Lac ₁ ⁻ Gal ₁ ⁻ Lp ⁺
W811	F ⁺ M ⁻ Lac ₁ ⁻ Gal ₁ ⁻ Lp ⁺
W902	F ⁻ T ⁻ L ⁻ B ₁ ⁻ Mal ₁ ⁻ Gal ₂ ⁻ Lp ⁺
W1210	F ⁺ M ⁻ Lac ₁ ⁻ Gal ₂ ⁻ Lp ⁺
W1436 W1437	F ⁺ T ⁻ L ⁻ B ₁ ⁻ Lac ₁ ⁻ Gal ₁ ⁻ Lp ^S S ^R
W1924	F ⁺ M ⁻ Lac ₁ ⁻ Gal ₁ ⁻ Lp ^R
W2175	F ⁺ Gal ₂ ⁻ Lp ⁺
W2229	F ⁺ M ⁻ Lac ₁ ⁻ Gal ₁ ⁻ Lp ^S
W2281	F ⁺ M ⁻ Gal ₂ ⁻ Lp ^S

Genotype symbols refer to the following characters

F⁺ is compatibility status, F⁻ is
M, T, L, B₁ nutritional requirements for methionine, tryptophan, leucine, thiamine

- Compatibility status, F⁻;
- Nutritional requirements; M, methionine; T, tryptophan;
- L, leucine; B₁, thiamine;
- Fermentation reaction; Lac₂⁻, lactose negative; Gal⁻, galactose negative; Mal⁻, maltose negative;
- Phage status; Lp^S, lambda sensitive; Lp⁺, lambda lysogenic; Lp^R, lambda resistant, but not overly lysogenic;
- Dry resistance, S^R, streptomycin resistant.

Table 3 (EK panam.)

Factors to transduce

98

Marker

Recip Culture

Donor

Pg

lac

W112 (L_P^R)

K12

71

" (L_P^R)

"

85

" (L_P^R)

"

94

" (L_P^R)

"

94

Serini or flye

W1678

"

76

Leuc.

W1736

"

75

W1736

"

78

W1476

(W928) (W931)
W2046, W1954

113

Methionin

58-161

K12 (uv. irradiated)

82

W811

K12 (uv. irr.) (beta)

83 (200)

(M added also to pet B⁺)

W1821

K12

85

W578

HFT 892 (mix)

180

(in Bgal, replica to D0)

Lysine

W1821

K12 (uv. irr.)

83

W1821

K12

85

W811

85

W1821

K12

130

Streptomycin

W578

W1821

95

Proline

W2062

K12

104 (?)

W2062

K12

105

W2062

K12

106

W1692

K12

96

W1920

K12

96

W2062

(prototroph)
HFT 1⁻ (2)
(prototroph)
HFT 2⁻

220

(M⁻)
HFT 4⁻
(prototroph)
heterotrophic

W2062

lytic λ (from M⁻)

227

Table 3 (Cont)

99

<u>Marker</u>	<u>Recip Cnt</u>	<u>Dmn</u>	<u>Page</u>
Maly-	W2071	K12(?)	119
Maly-	W2347, W2331	HFT2	298, 275
Sra -	W 2307	HFT2	298
F+	1321	HFT2	294

Frequency of Unstable Transduction - lysals

(100)

Parent Cross	(+)	1-	2-	4-
1- L_p^0	$9/22$ 17/24 (41)	-	$0/11$ 0	$0/29$ (0)
$L_p^+(1)$	$23/24$ (96)	-	$23/24$ (96)	$0/27$ (0)
$L_p^+(1)(2)$	$17/24$ (71)	-	$24/24$ (100)	-
2- L_p^0 (2291)	20/48 (58)	63/78 (88)	-	64/78 (89)
$L_p^+(1)$ (275)	$22/24$ (92)	$19/24$ (79)	-	$4/24$ (67)
$L_p^+(2)$ (1210)	$16/24$ (67)	$2/24$ (88)	-	$22/24$ (92)
4- L_p^0	$13/24$ (54)	0/72 (0)	$21/24$ (87)	-
L_p^+	$20/24$ (83)	0/96 (0)	$19/24$ (79)	-
L_p^R	$29/48$ (60)		18/24 (67)	-

Genes L_p^0 6/8
 Cals = 1210, 275
 above

$S = 56/102$ (47%) = 59%
 $+ = 98/120 = 81\%$
 $n = 29/48 = 60\%$

Total
 Total
 4844^4
 609 total

$67 \overline{) 484.0}$
 0.7
 63