

101

Tuesday 7/15

Cueber

518 - make 4x2 R

811 - make autotrophs

1655 - look for gal

1485 - natural assay

750 - gal ducter on film ch

518EK-12 - stability check (checked in E11B gal from very hot cult.)

811EK-12, stability check (checked aut. by hand)

lysate

streaked again 1-2

On the cross 811EK-12 X 1436 page 100

all colonies on plates picked. - 83 grew on first streaking

judging by streaking EMB gal checked

10 gal - 14
73 gal + 73

of the gal + streaking 20 show evidence of mosaic colonies - some streaking with several mosaic colonies / streaking. (-27%)

Not heated

of the cross 518EK-12 X 1436

shortage of gal plates

20 gal - picked and streaked } - 20 grew } all from one cross plate
20 gal + picked " " } - 20 grew } other plate saved.

- of the gal + streaking 5 showed mosaic colonies - (25%)

(rechecked)

1692 X 1402

Purification - not heated 3rd time

EMB gal - 2 representative of 3 originally received.

20/2

1/2

Wednesday July 16 - 52

- 518EK-12 X 1436 } See previous page
 20 gal - } primary streakings - restreaked - ✓
 20 gal + } unstable, with mosaics, picked first in order - then apparent stable.

- 811EK-12 X 1436
 10 gal - picked from 1st streakings and restreaked.
 Out of EMB gal - other gal + until 7/17 -

- 518EK-12 } lysate - cleared after 4 hours. ✓
 - 811EK-12 }

Calculation: $\frac{1.3 \times 10^3}{1.4 \times 10^5} = 1/10^7 \lambda$ Galley -

Thursday 7/17/52 - 1924 from two days previous Monday 7/14
 not streaked, 10% λ restreaked 10-12 λ no top & deep

EMB gal	1924 cell	restreaked	10-12 λ	no top & deep
1	0.1	0.1	-	29
2	0.1	-	0.1	129

check for lysogenicity -

Thurs. - 518EK X 1436 - ABOVE - 2nd streaking.
 20 gal - streaked - 17 grew (1 det) all pure gal - ✓
 20 gal + - streaked - 20 grew all mixed \pm col. less mosaic (3)
 Then in 1st streaking
 1st 5th streaks showing mosaic - ~~one~~ streak mosaic

518EK-12 X 1436 ABOVE.
 12 gal - picked + streaked pure - 2
 of 12, 11 pure gal - , - 1 mixed -

518/ λ - λ - 2 resistant col apparent.

518EK-12-12 } - from stock \rightarrow overnight cult. streaked out on EMB gal
 811EK-12 } both show mixed \pm col - \leftarrow 10 gal +
 1 gal -

Thursday 7/17/52

- 518 EK-12 x 1436
 - 19 gal - from Purification 2 - restreaked. (6/plate) ✓
 - 20 gal + " " " " " (no particular order) ↓
- 811 EK-12 x 1436
 - 17 gal - from pump 2 - restreaked ↓
- 518 EK-12
 - 811 EK-12 from stock cultures
 - 1. 5 streaking ~~from~~ from first bottle culture during mosaic
 - 2. single colony picked, restreaked. ↓
- 1924 - galvanized penny page.
 - all control plate papillae picked streaked EMIB gal = 29 pop.
 - equivalent no of " in galvanized picked. = 29 pop. ↓
- 811 EK-12
 - gal + passed up yesterday
 - ? diacardas, remain der streak (P-2) as EMIB gal. ↓

Friday 7/18/52

Report on above

- 518 EK-12 x 1436
 - 17 gal - all gal - ✓
 - 20 gal + 19 appear mixed, no gal + pure.
- 811 EK-12 x 1436
 - 17 gal - all gal -
 - 64 gal + (1 plate with lost from original 72) 62 unstable, mixed,
- 1924 EK-12
 - 29 untraced plate pop. streaked - 28 grew - all range of +, from dark to light
 - 15 appear mixed, no mosaic colonies noted.
- 811 EK-12
 - 27 from K-12 plate streaked - 21 grew - all range of +
 - 10 appear mixed, maybe a mosaic colony.
- 518 EK-12
 - 811 EK-12 } streaks streaked as EMIB gal
 - still mixed ✓

Friday - 7/18

- 1924 - heated and unheated papillae restudied - P2
- 28 heated papillae streaked against 1485
- 24
- 811EK-12 X 1436 -
- papillae picked from P2, stored overnight in refrigerator.
- streaked P3 Saturday 7/19

Saturday

7/19

1924 above - of streaking P2

Spontaneous 28 gel +, all pure gel +, varying degrees of gel-t-ness

Kingdual 12 +, 3/12 mixed, also varying degrees of gel-t-ness

plate with 8. dropped (contained few if any unstable)

suggestion of genetic stability?

streaking against 1485

28 gel + from K-12 ductin - none showed evidence of phase action in 1485

1924 above restudied.

Sunday - Monday 7/20 - 7/21

isolate 1	EMB gel	protein DCO	ductin heated 1485	K-12 λ	no prop 2 day
+	-	-	0.1	-	9
+	-	-	-	0.1	271
-	+	-	0.1	-	
-	+	+	-	0.1	
isolate 2					
+	-	-	0.1	-	2
+	-	-	-	0.1	180
-	+	-	0.1	-	no growth
-	+	+	-	0.1	solid growth

Gal 4 -

$$\frac{2.7 \times 10^3}{1.9 \times 10^{11}} = 2/10^8$$

solid grow - protrophic?

7/21 1924 EK-12

- 29 P-2 papilla on EM13 gel - only one appears mixed - discard and repeat liquid for better background.
- 28 P-2 spontaneous papillae - all pure gel + - discard -

Monday
7/21 ~~Friday~~ (cont)

- lysate streaked out on NSA

- 750 check on stable salt from transduction
EMB for $\frac{1}{2}$ motility $\frac{0.1}{-}$ $\frac{K-12}{0.1}$ λ
no prep. 2 days
c. 254

$\frac{2.54 \times 10^3}{1.9 \times 10^{10}} = \frac{2}{1.9} \times 10^{-7}$
 $\frac{1}{5.5 \times 10^6}$

- 2062-1, 2062-2

-1 $\frac{EMSAe}{2}$ heated $\frac{0.1}{-}$ $\frac{K-12}{0.1}$ λ

no growth again see pg 106

-2 $\frac{1}{2}$ $\frac{0.1}{-}$ $\frac{0.1}{0.1}$

} again shows prototrophy - discarded

7/22 Tuesday

- Repeat crosses -

81CK-12 X 1436

518CK-12 X 1436

- Repeat 1924

- Repeat 2050

See next page

Thursday July 24 -

1924 - Gal duckin repeat -

EMB gal	Addition	no. of colonies after 7 days
1	-	40
2	0.1 ml K12	161
3	-	19

2062

Attempted moline observation -

EMB gal	Addition	Observation after 7 days
1	-	no growth
2	0.1 ml K12	no growth

2050 gal 3 -

EMB gal	Addition	no. colonies after 7 days
1	-	11
2	0.05 ml K12	1/4 = 172
3	0.1 ml K12	1/4 = 223

Gal 1 -

$$\frac{1.2 \times 10^3}{2.3 \times 10^3} = \frac{1}{1.9}$$

Gal 3 -

$$\frac{8.8 \times 10^8}{2.3 \times 10^{12}} = \frac{1}{2.6 \times 10^6}$$

$$\frac{1.3 \times 10^4}{2.3 \times 10^{11}} = \frac{1}{1.7 \times 10^7}$$

518 TK-12 X 1436 repeat on cross

- unincubated cell suspension used = 10% used 0.1 ml plated -
 518 TK-12 suspension from streaking on EMB gal. approx. mixed (10 gal + / 1 gal - ?)

EMB gal	Addition	no. colonies	no. gal +	no. gal -
1	0.1 ml 518 TK-12 cell	0	0	0
2	0.1 ml 518 TK-12 + 0.1 ml 1436 cell	35	0	35
3	" " " "	30	0	30
4	0.1 ml 1436 cells	0	0	0

approximate results obtained previously -

811 TK-12 X 1436 repeat cross

- unincubated cell suspension as in above cross -
 811 TK-12 suspension from streaking on EMB gal. approx. mixed (20 gal + / 1 gal - ?)

EMB gal	Addition	no. colonies	no. gal +	no. gal -
1	0.1 ml 811 TK-12 cell	0	0	0
2	0.1 ml 811 TK-12 + 0.1 ml 1436	12	11	1
3	" " " "	17	16	1
4	0.1 ml 1436 cells	0	0	0

Phage lysates examined for sterility - streaked on ~~EMB~~ TSA

Source	Result
1821	no growth
heated K-12	no growth
892	no growth
750/1821	1 small colony =
1437	no growth
811 (gal +)	no growth
K-12	no growth
811-2	no growth
811	no growth
750	no growth
902	no growth
1736 gal +	no growth

Invad. K-12 lysate

Time	Result
60	no growth
120	no growth
180	no growth
240	no growth

Mol. tested

Order of stability 750 → 2050 → 1929 → 811, 513
~~750 → 2050 → 1929 → 811, 513~~

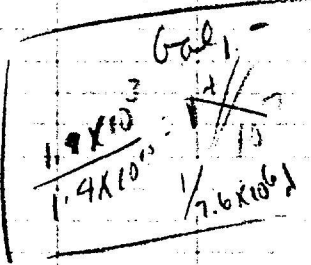
Saturday 2/26

- 750 from pg 105 - Check on stability of goldbees from treated plate 40 poplar plates
 1st streaking - 12 appeared unmixed - ~~14/20~~ 14/20
 28 appeared mixed - ~~28/29~~ 28/29 mixed
 7/27 2nd streak box mixed 28/29 mixed

- 2050 gold - pg 106 - stability check in goldbees
 5th spent picked - all mixed 1st streaking
 - all pure 2nd streaking

10 K-12 goldbees plates - all mixed 1st streaking
 - 9/10 appear pure 2nd streaking
 10/10 appear pure 3rd streaking

- 1929 + K-12 29 pag picked from treated plate for stability check
 1st streaking 15 appeared mixed - 1st streaked -
 5/29 appeared mixed - 2nd streaking
 8/29 " " " 3rd streaking



- 750 - Stability and nature of med. goldbees - cells from over night unselected culture.

1	2	3	4	5	6	7
	K-12	186	186			
	60 K-12	107	886			
	124 K-12	208	1664			
	180 K-12	161	2576			
	140 K-12	183	2928			
	λ-2	0				

— Pag picked for here for stability check.

29 pag picked from un 120 and streaked.
 22 grew - 20/22 appear mixed 2nd streaking

discarded - attempt with 93 to see if possible to pick up gold h⁵

- 811 + K-12 X 1436 -

carried through 8 single colonies isolation after purification - still signifying gold -
 discarded -

Sunday 7/27

gult

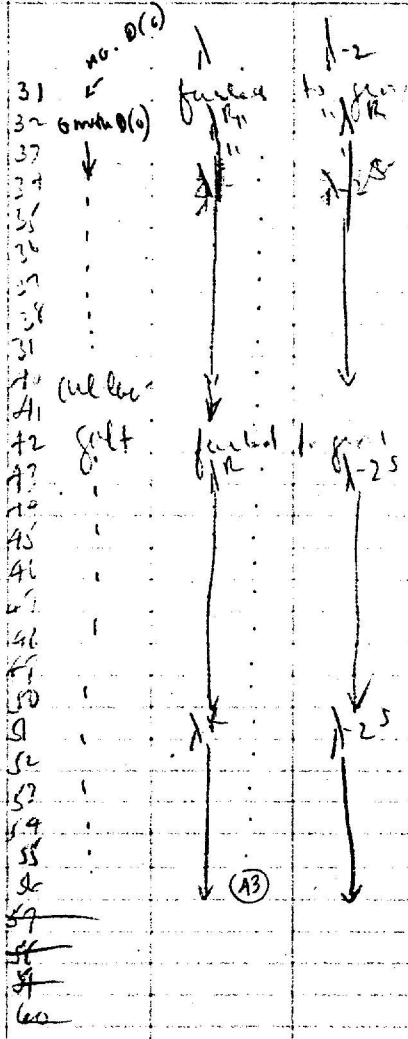
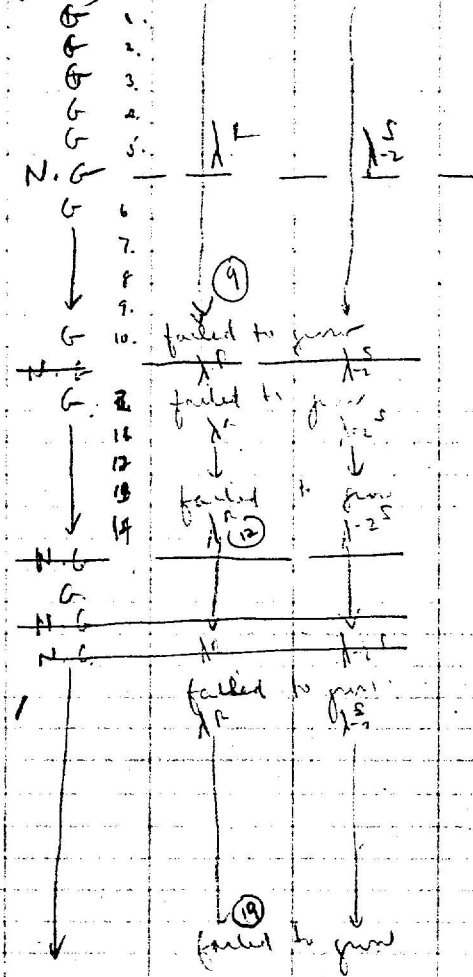
811EK-12 X 1436 - check on purified proto top he

test next week

- 1.
- 2.
- 3.
- 4.
- 5.
- 6.
- 7.
- 8.
- 9.
- 10.
- 11.
- 12.
- 13.
- 14.
- 15.
- 16.
- 17.
- 18.
- 19.
- 20.
- 21.
- 22.
- 23.
- 24.
- 25.
- 26.
- 27.
- 28.
- 29.
- 30.

all done -

new gult in notebook



- Tuesday 7/29 ^{AMP}

518EK-12 X 1436 - I

purified "prototrophs"
20 gal + examined, 19 prototrophic
all lac-
all lysogenic when replicated on 1485 ✓

20 gal - examined, 18 prototrophic
all lac-
all non lysogenic when replicated on 1485

811EK-12 X 1436

purified "prototrophs"
12 gal - examined 9 prototrophic, all lac- all λ-2 sensitive
5 lysogenic when streaked against 1485 } one lysogenic found λ^R from before
4 sensitive when " " " } one found sensitive, found non lysogenic, above

from page 105 { 20 gal + picked, all lac-, 6 failed to grow on D(1)
of those growing - D(1)
43 tested successfully against λ - all λ-2
50 gal replicated against 1485 - all lysogenic

750 X 1503 on EMS gal

1177 X 1655 on EMS med

750TK-11

gal + 6 stable } from pg 107 - picked and restreaked - further motility and progress unstable → stable
6 unstable } EMS gal
6 unstable - unstable after 3 transfers - mixed gal+, gal-
6 appear stable, with possibility of being mixture.

1924TK-11

of 24 cultures - pg 107.
8 unstable on EMS gal - streaked against 1485 - no evidence of lysis - all presumed non lysogenic
8 apparently remain unstable
↳ continued streaking of 8 = streaking (#1) (gal+, gal-)
streaking not onward 8 unstable

Wednesday July 30 -

- 1655 X 902 Lp2^s for v on stability of gal⁺ - begun 7/28

EM18 gal addition no. pop. 2 days

1 0
2 0.1 ml K₂Cr₂O₇
↑
amounts
1.4 x 10¹⁰

- 8 streaked out for stabl.
- 29 streaked out

15
14 = 49.3 x 10⁶ l

- Thursday 7/31

1655 X 902 Lp2^s above

8 simultaneous streaked - all mixed

see pg 111

24 x 12 - all mixed

Cross - mirror page

750 X 1503 to get TLB - gal⁺ - EM18 gal⁺

central plates p.d.uch
mixed plates 1. 2 gal⁺, 13 gal⁻
2. 2 gal⁺, 9 gal⁻

pick gal⁺ + streak on EM18 gal⁻

1177 X 1655 to get mal⁻ - - - for gal⁺ detection run -

central plates barren
mixed plate 1. c. 1000 colonies, (0.9 gal⁺ w/ 1 gal⁻)
2. c. 1000 " " "

- 98 with mod. lambda to get gal⁺ detection without lysogenicity

EM18 gal plate add. ki no. pop. 2 days

1 0
2 0.1 ml 1/1000

19
61

pick 29 - examine for

stability

see pg 111

1. pick up gal⁺ colonies to see if resistant, stable - if no spontaneous mutation
2. if lysogenic, stable, probably spontaneous
3. if lysogenic, unstable, probably

NOT INDEXED
BEYOND HERE

Thursday 7/31
Friday 8/1

- 518 t w 240 K-12 λ - 29 papulae picked in attempt to find 1st gold. ^{unstable?} ✓
all 29 appeared mixed
1st streaking 7 streakings showed mixed colonies - these streaks against ~~1st~~, ^(see 1st)
all restreaked. _{discarded}

- 902 X 1655 Lp₂⁵

✓ on stability of gal duces
of 30 papulae picked

2nd streaking - 5 unstable still on gal duces - 4 unstable 3rd
restreaked -

1 unstable in 6 spontaneous - not unstable 3rd streaking } discard

- 750 X 1500 - isolation of diploid for purpose of securing TB, Gal⁻
only suspicious colony from 1st cross found. stable EMS gal - discard

- 1177 X 1655

1st 3rd ~~gal~~ mal - prototrophs picked, streaked on EMS mal - picked
and streaked against 1-2 - all 1st discard

24 additional picked and streaked - tested against 1-2 -
- these 14 also 1-2 resistant - pick more than -

8th

- 518 K-12 w 240 above ↑

8 unstables noted 2nd streaking - streak against λ ✓
1 proved to be sensitive and probably unstable - picked, restreaked
and tested against λ - probably sensitive, definitely unstable - recheck.

- 902 X 1655 Lp₂⁵ - Gal ductin done in EMS gal

EMS gal	Addition	no. pap 2 days
2	0.1 K-12 λ	295

1/55 X 10⁶

9/14

found 1st but stable - repeat entire exp -

from the crosses of gal duces X 1436 - check gal-segregants from the prototroph
obtains of the cross for 1st

811 K-12 X 1436

7 gal-segregants picked and tested - all 1st

518 K-12 X 1436

5 gal-seg. picked and tested - all 1st ✓

- Monday 8/4

- In the crosses of the unstable, non-lysogenic gal⁻ duces, of 1974 (tp^R) 8 unstable forms obtained.

Cross of 1924tk-12 (1) x 1436 as EMS gal

EMS gal plate	Addition
1	0.1 ml 10X 1924tk-12 (1) cells
2	0.1 ml of 1436, "1924tk-12"
3	" " " " " " " "
4	0.1 ml 10X 1436

Reading after 2 days

barren -
 11 gal +, c. 384 gal -
 5 gal +, (c. 142 gal -) = 568
 barren

16 gal + / c. 952

Cross of 1924tk-12 (-2) x 1436 as above:

1	
2	as above
3	
4	

7 gal -
 7 gal +, (c. 115 gal -) = 460
 4 gal +, (c. 133 gal -) = 532
 barren

poor medium may actually be larger number of +

11/992

- Tuesday 8/5

From the cross 1177X1655 to get Mal⁻ tp^S

30 additional gal- prototrophs picked and checked out (see pg 111) - tested against 1-2 all 12^R - total to date picked 80 all tp^R

From the cross 1924tk-12 x 1436 (#1)

26/30 gal- prototrophs successfully tested against 578 for lys - all showed no evidence of lysis / possible 3? plaque

15/30 gal+ prototrophs successfully tested against 578 for lys - all showed no evidence of lysis

30 prototrophs (#2) picked at random - checked / 578 - all showed no evidence of lysis

Thursday - sick

Friday 8/8

14 36 Gal ductin - Test 1 1954 and 2046 for *Penicillium ductin* -

EMB gal	Additi	no populae 2 day
1	- 0 -	5
2	- 0.1 K12A	$\frac{1}{2} = 329 = 1296$
3	0.1 $\frac{1}{50}$ 290A	77
4	0.1 1954A	$\frac{1}{4} = 277 = 1116$
5	0.1 2046A*	38

1.3×10^4
 0.5×10^4
 0.6×10^4

(has about 1 pop.)
(some pop & halos, some without - no apparent halos)
(no halos)
(some with and some without halos)
(no halos about pop.)

8 plates checked
2nd streaking all
streak + 2 sep
8 plates
2nd streaking
all - 11/10/51

D(O)HBT	no. col.
1 - 0 -	1
2 0.2 296A*	1
3 1954A	2

Re-examine

New lysate - from 8/7

K-12 mal	30 ml	$> 46 \times 10^8 = 4.6 \times 10^9$
"	35 ml	$> 11 \times 10^9 = 1.1 \times 10^{10}$
"	40 ml	$> 3 \times 10^9 = 3.0 \times 10^9$
1954		$> 140 \times 10^8 = 1.4 \times 10^{10}$
2046		$205 \left. \begin{array}{l} \\ \end{array} \right\} \times 10^8 = 2.1 \times 10^{10}$ $206 \left. \begin{array}{l} \\ \end{array} \right\}$

8/11 23 picked, streaked and single colonies tested for λ^S - all were resistant to λ and presumed lyogenic

13 picked and streaked, after 2nd streaking 3/13 mixed -

2 spontaneous picked and streaked, 1/2 pure galT 2nd streaking

From the cross 1177 X 1655 to obtain Mal- λ^S

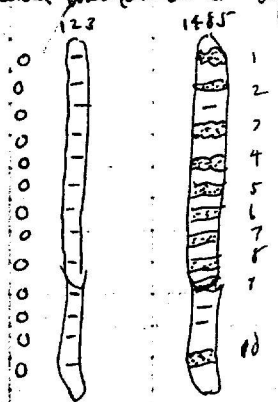
40 odd mal- picked, streaked, picked and streaked against λ^S all λ^S total to date

50
40
120

Wednesday 8/13

- 1924EK12
 - 6 unstable gal-decuss from pg 112 and before (these 6 are the 6 not yet) crossed with 1436
 all cross streaked against 1455 in EMB base -
 reaction slightly interfered with but all showed no evidence of lysogenicity

- 1436 + 2046 of pg 113
 the 13 gal decuss picked and checked - against NCC 123 and 1455



more lysogenic in 123
 10/13 lysogenic in 1455

~~2070~~

- 1673 gal - made by cloning e. 10⁸ cells for 5 sec exposure to ^(kanamycin) U.V lamp.
 picked and purified 3 times.

NO	GAAVCTW	EMB gal	addition	no pep. 2 days
3		+	Δ K12 λ=0.1	0
4		+	811 λ=0.1	0
		+	K-12 λ=0.1	1 (small)

sensitivity of this culture -
 + λ not fixed

probably
 in error
 not gal - but gal strain

From the cross 1177K1655
 24 additional Mal- picked, checked, picked and tested against λ-2 all lysogenic
 total to date 144 - Recross and try again!

Wednesday - 9/3

- 1655 mal - continue purification to obtain low growth background and reexamine for mal direction.
- 1177X 1655, second cross to obtain a mal- type for the purpose of attempting mal-direction.

Thursday 9/4

- Adsorption characteristics of 811

- washed cells, resuspended in saline - conc. of 16 hr cult in br.
- phage titer = dil $10^7 \rightarrow 0.1 \text{ ml} + 0.1 \text{ ml}$ 1485 cells \rightarrow
- cell titer = dil $10^6 \rightarrow 0.1 \text{ ml}$ plated on EMBAe \rightarrow
- 1 ml cells mixed 1 ml phage -

factorial

at time 1 min - 0.1 ml mix added to 10 ml broth - centrifuged 15 min - (all sample at once)

2 min - 0.1 ml as 1 min

5 min - 0.1 ml as above

10 min - 0.1 ml as above

Time Sample	dilution
1	10^7
2	
5	
10	

Monday 9/22

EMLS count of 518 X 1956 - for mal-4pr^s in EMS mal

plate	mal+	mal-	total
1	9	43	52
2	6	29	35
3	14	68	82
4	8	64	72
5	7	28	35
	<u>44</u>		<u>276</u>

$$\frac{276}{44.0} \times 0.158 = 15.7\% \text{ Malt}$$

$$\begin{array}{r} 276 \\ 44.0 \\ \hline 1640 \\ 1380 \\ \hline 2600 \\ 2208 \end{array}$$

EMLS count of 518 X 1817 - for Mal-4pr^s in EMS mal

		F+	
1.	24	5	29
2.	35	0	35
3.	38	2	40
		<u>7</u>	<u>104</u>

$$\frac{104}{17.00} \times 0.067 = 6.7\% \text{ Mal-}$$

$$\begin{array}{r} 104 \\ 17.00 \\ \hline 624 \\ 760 \\ \hline 728 \end{array}$$

1655 mal- obtained from J.L. - incidental sto. in EMB mal -
 purified through 3 single colony isolations
 Entered in stock book - no. 2071

2071

Wednesday 2/24/52

- Characterization of Ag 16 mutants saved up from p. 75-68

Cultures revitalized by fresh growth single auxotrophic colonies of each picked and attached on EM13(-) - Replicated to

- O(-) + A₁ - 1 grew } same
- A₂ - 1 grew }
- A₃ - none

A₄ - 10 of 11 remaining grew for control (thin plate) - rechecked.

- ~~A₄~~ - Can't find must have forgotten or lost - recheck if A₄ give unidentified
- Vit. - all failed to grow
- HC - all grew

- results

811 galductin by 811 gal⁺ λ -

811 gal⁺ lambda presumably contaminated - checked below
EM13 gal⁺ addition on peptone 2 days

1	none - cells 811	39
2	0.1 ml ^{811 gal⁺} λ (no cells added)	64 (colonies)
3	0.1 ml ^{811 gal⁺} λ + 0.1 ml 811 gal ⁺	78

apparently no activity of gal⁺ + 811 λ or 811 negating that this gal⁺ is not a true reversion but a suppressor mutation. Cross 811 gal⁺ x 1436 gal⁻ if gal⁻ occur further information of suppression.

- Summary of Mat- h₂^s from 1177 to date.

since p. 114 about another 60 wal- picked from 165TX1177 cross

total	p. 114	144
since		c. 60
total		204

From the cross plates EM1 p. 119

picked and -	56
	260

- this checked on EM1 and replica'd against h₂

in this lot one possible h₂-2 sensitive.

Thursday 9/15/52

- 811 - Effect of heat on lambda - a goldschmidt effect.

- K-12 λ lot labelled #7 titer approximately 2.3×10^{10}
- immersed in water bath at 36C for 30 minutes

no post-tensigaku likely

EMB gel Addition

no pop. after 2 days

$\frac{2 \times 10^2}{2 \times 10^5} \lambda$ dil



- 1 - none
- 2 - 0.1 ml K-12 λ
- 3 - 0.1 ml K-12 λ heated as above

37
 56
 256
 increase
 19
 219

Apparently the activity in lambda lysate is destroyed by this heat treatment. It remains to be seen what has happened to plaque forming lambda.

titer = $2 \cdot 10^9$ dil = 1 = 2×10^9 λ /ml
 $2 \cdot 10^8$ dil = 12 = 2.4×10^9 λ /ml - reduction in phage titer

- 811 Spontaneous gal+ above - picked, streaked on EMB gel to isolate a few gal+ of both suppressor and true back mutation type - make lysate of both types for goldschmidt (by successive?)

- Wg 16 mutants collected - see 118

1. mutant growing on both A₁ + A₂

On breakdown of these two groups - the mutant grew in both methionine } better in methionine judging by leucine } turbidity

2. Remaining 11 mutants judging by check on D(i) + A₁ replicates are all A₁

Examination in liquid med (A)₁+pr, has 147 grows indicating protose dependence - discarded immediately - these two appear different on A₁ plate - more opaque

remaining 9 are list - (from remainder) preserved to stock.

(methionine - leucine - see list)

- 1655X(17)?

meal - 4⁵ serial continues.

3² addition meal - picked - was tested against 1-2

Friday 9/26

- 811 gult sp papillae from 121
 gult applied into water - checked in box -
16 in all

discarded

- 2070 = 1673 gult -

EMC suspicious that in her 2070 λ^+ is not gal - but gal^s
 or something - λ of 1673 gal - on gal -
 picked from slant with that checked in gal - not gal - but show

- W916 pr⁻ X₁ -
 W916 pr⁻ X₂ - } picked from slant and streaked out in EMBlac.

Sat 9/27

- Cross

811 gult X Y-10 - washed cells - (out cult. swab in 2 ml saline)

811 gult #	EMBlac plate	Addition	73 gal ⁺	73 gal ⁻
1		Y-10	0	
2		811 gult	0	
3		both	18	0
4		"	13	0
5		"	21	0
6		"	21	0
			<u>73</u>	<u>0</u>

sort of gal sl?

- W916 pr⁻ X₁ - replicated to D(6) + pr } X₁ failed to give in replication to D(6) + pr
 W916 pr⁻ X₂ - " " " " } X₂ not discarded

Sunday 9/28

- 1177x 1655 - to obtain Mal- hp_2^S
- 24 additional tested - all hp_2^R -

total to date

260
32
24
<hr/>
316

- W916 pr-x₁ in Embloc - a single lac^s or lac- tested. picked and streaked in lac- odd appearance - some colonies with no dark centers most lac^s but with sectoring ⊙ ⊙ ⊙ opaque sectors etc. discarded

- W916 meth - penicillin run -
culture grew -
plate out EM13 lac^s und., 1-10, 1-100

- Comparison of K-12 lysates.

λ titer (p. 124)	EM13 gal ⁻	addition	no. of <i>Prophages</i>
2.8×10^9	1	none	42
1.1×10^{10}	2	0.1 ml X λ	125
2.8×10^9	3	0.1 ml 30 λ	186
2.9×10^9	4	0.1 ml 35 λ	164
	5	0.1 ml 40 λ	131 ← contaminated.

$\frac{1.44 \times 10^3}{2.8 \times 10^9} = \frac{1}{2 \times 10^6}$

These cells retested 2 days later for ability to be gal⁻ ahead

33
72

- 2nd Cross

EM13 gal⁻ X Y-10⁺ EM13 gal⁻ control plate banner -

washed cells reprod. = saline 3ml, 0.1 ml / plate

EM13 gal ⁻	gal ⁻	gal ⁺
1	215	0
2	161	0
3	142	0
4	112	0
	<hr/>	
	630	0

Cross 1	+	73	0
Cross 2	+	630	0
		<hr/>	
		703	0

Sat. 10/4

- Comparison of λ on 811 and 518
use m180 d this time

EMB gel	811	518	m180 λ	m. pag. 2 days
1	0.1	-	-	5 ²
2	0.1	-	0.1	2008
3	-	0.1	-	56
4	-	0.1	0.1	932

not much clarity added by this ept. see previous epts

- Wednesday 10/8

Wg16 melkimm - three - previously isolated - assigned #2097 - culture to D. Shaw

- Phage titer

lysate from 1-12

u.v. dose	X	10 ⁷	33, 33 = 2.8 x 10 ⁹ λ /ml
	Y (2 probd)		43, 67 = 5.5 x 10 ⁹
do	30-1		116, 98 = 1.1 x 10 ¹⁰ λ /ml
do	35-1		15, 40 = 2.8 x 10 ⁹ λ /ml
	40-1		(0.1)26, (0.3)94 = 2.9 x 10 ⁹ λ /ml
0.5 no tested in Pen-form stable	15		0, 6 = 3 x 10 ⁸ λ /ml
	25		176, 141 = 1.6 x 10 ¹⁰ λ /ml
	35		189, 78 = 1.3 x 10 ¹⁰ λ /ml
6250 811EK-12	811EK-12	10 ⁷	5, 13, 9 x 10 ⁸ λ /ml

Two streak from 811

Thursday 10/9

- Friday prep -
Sambrook

- ① - isolated from lysate used - broth culture for X Y-10
- ② - stock labels #1 - previously X's X Y-10
- ③ - new isolate - purified 3X & broth culture for X Y-10
- ④
- ⑤
- ⑥
- ⑦
- ⑧
- ⑨
- ⑩

John
Stall

- Wg 16 meth - A₂ -

1 is apparently viable - (forms pellicle - curtain?)
1 is sterile -

stocks made

broth added to remaining 3 in the hope of
resurrection since original plate lost.

- Wg 16 meth - Three - X Wg 16 pr - X₁ -

EMIS loc Addition

- 1
- 2
- 3
- 4
- 5

no //

- 811 gal ductins with addition

0.1 ml of various lamella prep
no pipet as 2 days

Count 129

netio gold/1

- 1
- 2 15' K-12 A
- 3 25' K-12 A
- 4 35' K-12 A
- 5 750E 1821 A

53
58
145
146
247

$$\frac{9.7 \times 10^2}{1.6 \times 10^{10}} = X.7 \times 10^7$$

$$\frac{9.3 \times 10^2}{1.3 \times 10^{10}} = X.4 \times 10^7$$

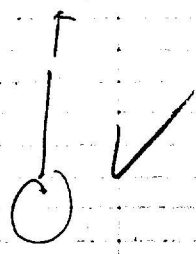
3X10⁸ d/w. ltr
1.6X10¹⁰
1.3X10¹⁰ } d/w
c. 6.5X10⁹ d/w according to prev. assay

0.9X10⁻⁷

- 51802 811 alone

- 1
- 2 15' K-12 A
- 3 25' K-12 A
- 4 35' K-12 A
- 5 750E 1821 A

- 20
- 78
- 691+
- 587+
- 963+



$$\frac{6.7 \times 10^2}{1.6 \times 10^{10}} = \frac{4}{10} = \frac{1}{2.5 \times 10^6}$$

- 750E 1821 lysate - 0.1 ml plated EMP gel - no col.