

Cistron analysis of *Lac* loci using *Lac*₂-F13.

29/11 1959

REF: W3112F13

	1	2	3	4	5	6	7	8	9	10
Phage										
Method										
1		Method: Spot method								
2		cultural age: overnight culture in <i>Pen. 5ml.</i> at 37°C.								
3										
4										
5										
6										
7										
8										
9										
0										
		Result:								
		x 3112 F13	x 3112	x 3747		blank				
		(<i>Lac</i> ₂) ^{nr}	(<i>lac</i> ₂)	(<i>Lac</i> ₂ ⁺ M-F13)						
<i>Lac</i> ⁻										
DMPG	1	3112	2	---	---	---				
	2									
	3									
unstable	4	3127	4	+++	+	+++				
	5									
	6	2247	7	●	---	---				
	7									
+ M unstable	8	3230	1a	++	+	+++				
	9									
- M	0	3089	1b	++?	---	+++?				
	1									
+ m	2	4112	52	+++	---	+++				
	3									
- f	4	4128	66	+++	---	+++				
	5									
- m	6	4147	87	+++	---	+++				
	7									
- m	8	4148	88	+++	---	+++				
	9									
- m	0	4149	89	++	---	+++				
	1									
- m	2	4150	90	++	---	+++				
	3									
- m	4	4151	91	++	---	+++				
	5									
- m	6	4153	93	++	---	+++				
	7									
	8	2243	<i>Lac</i> ₃	+++	---	+++				
	9									
	0	2245	15	++	---	+++				
	1									
	2	2247	<i>Lac</i> ₇	---	---	---				
	3		also Gal ⁻							
	4									
	5									
	6									
	7									
	8									
	9									
	0									

on H₂O. * various *Lac*⁻ mutants. (F⁺ *Lac*⁻)
 cultural age: overnight culture in *Pen. 5ml.* at 37°C.
 W34) F⁺ *Lac*₂ X⁺ —, cure.

penicillin and β-galactosidase
 2247 7 ● — ? (check) — nontransducible *Lac*.

many unstable recombinations
 Reaction is very slow.

Test segregants.

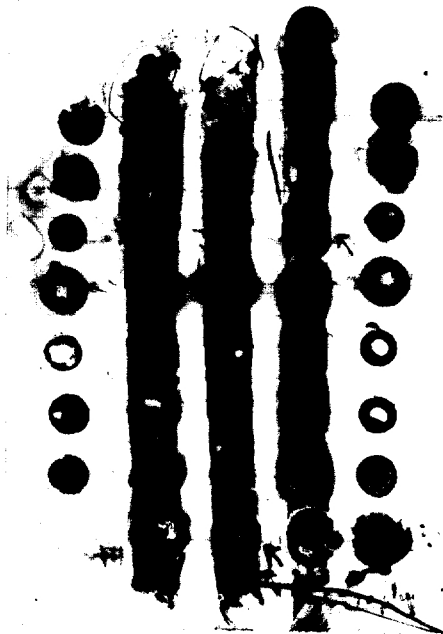
This black spot gives many *Lac*⁻ on BLac after streaking.

synthropy

non-transducible *Lac*
 λ_2^S
 non-transducible Gal

46/43

3112F13
 3127D S
 3127E S
 3127G S
 3112
 3127
 3747
 T6 on BLac



	2a
3112	2
3127	4
2247	7
3230	1a
3089	16
4412	52
4121	61
2243	3

3112 3112 3747

F Lat K O
M

Superimposed between two different lines



4147	
4148	
4149	
4150	
4153	
2295	
2247	7

3112 3112 3747

Stability of Lac⁺ of the treated and untreated heterozygous diploid with A.O.

28/11/59

REF:

1	2	3	4	5	6	7	8	9	10
Lac ⁺ AO treated	Lac ⁺ stability	T ₆	T ₆	Lac ⁺ Untreated T ₆	Lac ⁺ T ₆		Colony		
1	U	m	U m	1	U	m	9f. Lac ⁺ found it is noted as unstable.		
2	U	m	U m	2	S	r	U: Unstable S: Stable.		
3	U	m	U m	3	U	m			
4	U	m	U m	4	U	m	Method: picked Lac ⁺ from streaked plate, inoculated into Prasey (1ml), and incubated then for overnight.		
5	U	m	S S r	5	U	m			
6	F S	S	U m	6	U	m	② streak on Blac, and see whether Lac ⁺ only or Lac ⁺ and Lac ⁻ on it.		
7	U	m	S m	7	U	m			
8	U	m	S S S	8	U	m			
9	U	m	S S r	9	U	m			
10	F S	S	S S	10	U	m			
11	U	m	U m	11	U	m			
12	U	m	U m	12	U	m			
13	U	m	U m	13	U	m			
14	U	m	U m	14	U	m			
15	U	m	U m	15	U	m			
16	U	m	U m	16	U	r			
17	F S	S	U m	17	U	m			
18	U	m	U m	18	U	m			
19	U	m	S S S	19	U	m			
20	U	m	U m	20	U	m			
21	U	m	U m	21	U	m			
22	U	m	Lac ⁺ (stable, unstable)	22	S	r			
23	U	m	S: 24	23	U	m			
24	F S	S	U: 48	24	U	m			
25	F S	S		25	U	m			
26	S S	r		26	U	m			
27	U	m		27	U	m			
28	U	m	T ₆ -resistance	28	U	m			
29	F S	S	U ₆ ^R : 8/69	29	U	m			
30	S S	r	U ₆ ^S : 13/69	30	U	m			
31	U	m	U ₆ ^S : 13/69	31	U	m			
32	U	m	U ₆ ^{mix} : 58/69	32	S	r			
33	F S	S		33	U	m			
34	U	m		34	U	m			
35	F S	S		35	U	m			
36	U	m		36	U	m			
37	U	m		37	U	m			
38	U	m		38	U	m			
39	U	m		39	U	m			
40	U	m		40	U	m			
41	U	m		41	U	m			
42	U	m		42	U	m			
43	U	m		43	U	m			
44	U	m		44	U	m			
45	F S	S		45	U	m			
46	F S	S		46	U	m			
47	F S	S		47	U	m			
48	U	m		48	U	m			
49	S S	r		49	U	m			
50	S S	r		50	U	m			

Method: picked Lac⁺ from streaked plate, inoculated into Prasey (1ml), and incubated then for overnight.

② streak on Blac, and see whether Lac⁺ only or Lac⁺ and Lac⁻ on it.

1. AO-treatment: AO 30g/ml. NSB-PH. 7.6. make up one small loop of ca. 10 cells/ml / 5ml NSB AO. The treated culture was streaked on Blac after 24hr incubation at 37°C.

Lac⁺ (stable, unstable)

S: 24
U: 48

T₆-resistance (Lac⁺ U₆^R)

U₆^R: 5/69
U₆^S: 0/69
U₆^{mix}: 62/69

m: mixture of U₆^R and U₆^S, segregated from U₆^{R/S}.

S: U₆^S
r: U₆^R
U: Unstable for Lac⁺ accounted for Lac diploidy.
S: Stable for Lac⁺

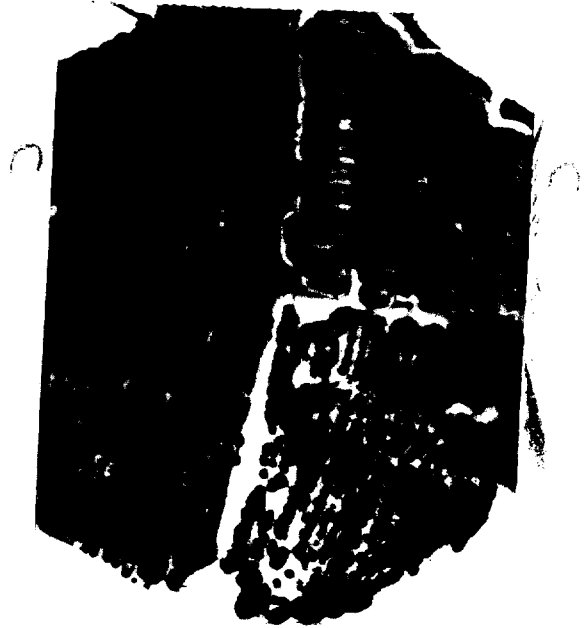
Conclusion treated with AO

1. All of the F₂ are Lac⁺ stable, U₆^R or U₆^S and Lac⁻ stable.
11/12 F- Lac⁺ U₆^R or U₆^S.
#17 1/12 F- Lac⁺ U₆^{mix}.

2. Lac⁺ is in T₆^R or T₆^S, and T₆^{R/S}.

This means, AO remove full segment of FlacU at a time.

U : Unstable
S : stable



S

U

Elimination of Lac-F13 segment with AO-treatment

23/VI; 1959

REF: Cf. P118

Experimental conditions.
 Inoculum size: ca. 10^8 cells/ml. AO: 30x/ml. Med: NSB for AO-T.
 Time: overnight at 37°C. Strain treated w/ 594 F13 (M^s V₁ L₁ V₂ V₃ V₄ V₅ V₆ V₇ V₈ V₉ V₁₀ V₁₁ V₁₂ V₁₃)
 Method: ① Suspend single colony into 1.0 ml of H₂O, and dilute to use the 10^{-6} ml. Add 0.1 ml to 5 ml NSB with AO, of and without AO.
 ② Seed 10^{-6} ml onto B-lac. 5×10^8 of untreated control.

Result I. Elimination of lac⁺ segment from Lac⁺/Lac⁻ heterozygotes.

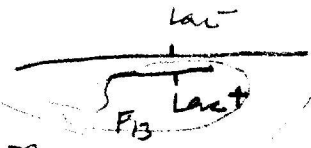
Plate #	I AO-treated (30x/ml)			II AO-treated (30x/ml)			C untreated control		
	Lac ⁻	Lac ⁺ Lac ⁻	Σ (%)	Lac ⁻	Lac ⁺ Lac ⁻	Σ (%)	Lac ⁻	Lac ⁺ Lac ⁻	Σ (%)
1	310	131	4	439	42	2	103	311	2
2	275	100	2	383	34	3	86	246	2
3	284	120	3	398	35	2	109	298	3
4	274	95	2	453	43	2	119	297	3
5	283	138	2	422	33	1	84	322	2
Σ									
%									

Results II. Elimination of F13 from Lac⁺/Lac⁻ heterozygotes.

E %	AO-treated (from #II)				untreated control			
	Lac ⁺		Lac ⁻		Lac ⁺		Lac ⁻	
	F ₁₃ H ₁ L ₀	F ⁻	F ₁₃	F ⁻	F ₁₃ H ₁	F ⁻	F ₁₃ H ₁ L ₀	F ⁻
	59	0	13	0	64	0	22	50
	(see back page)							

(Tested by bal-transfer. x 4573)

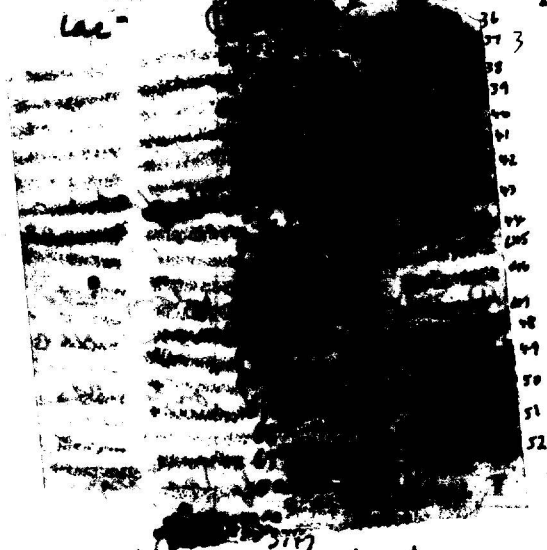
x4573 on M6al



Untreated control
Lac+ 3747 FD 2594 lac+ F13

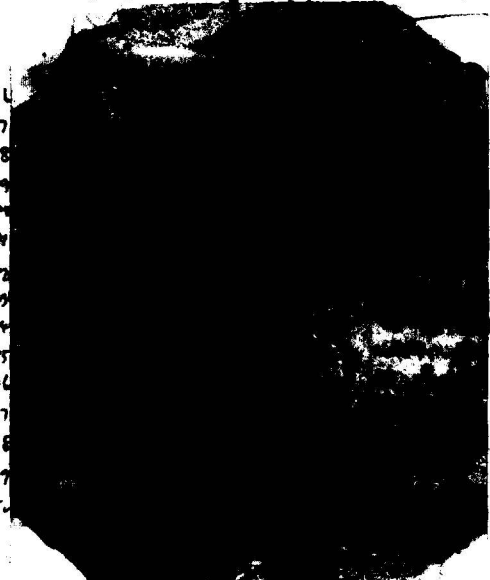


Treated with AD

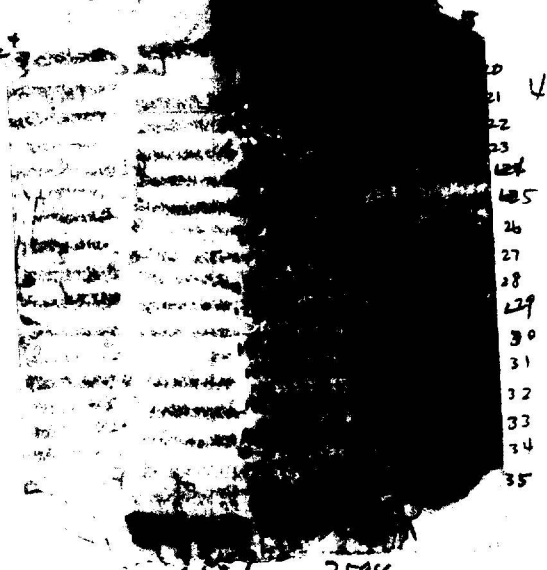


2594 lac+ F13

Lac+ 2594 Lac- x4573 on M6al



Lac- Lac+ 2594 lac+ F13



3747 2594

4121 (Lac61 F-)

3112 F13

3112 F-

on M6al

24 hrs.

Multiplication of Gal-F₈ segment.

w4520
W6F8 x W4573.

REF: p. 123

30/11 ~ 1959

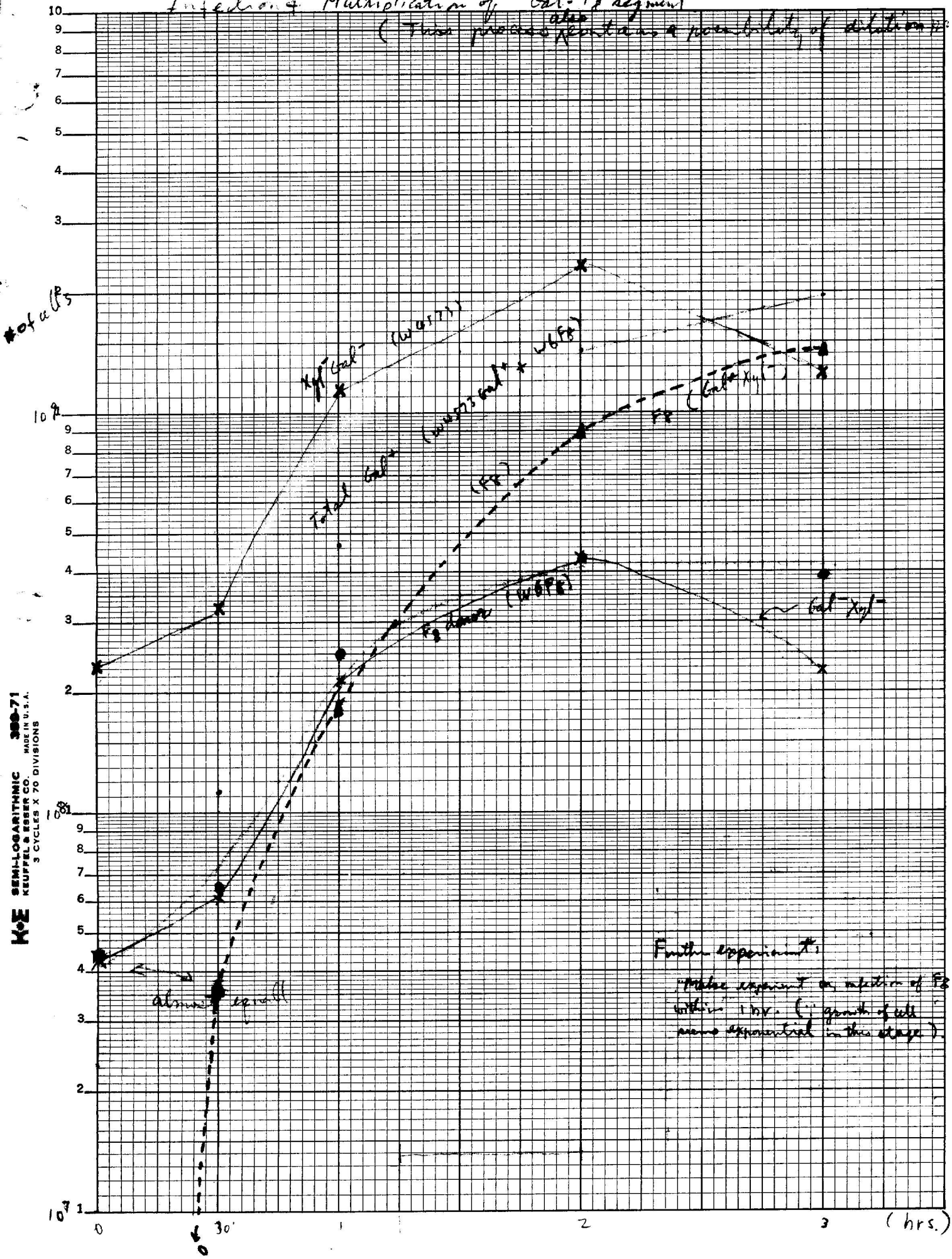
	1	2	3	4	5	6	7	8	9	10	
1	overnight culture										
2	0.2 ml / 10 ml pln.										
3						Inoculate it at 37°C					
4						on rotator.					
5						for 2 hrs.					
6						Mix { 1 ml W6F8 + 10 ml W4573.					
7	AM				PM						
8	12:00		12:30		1:00		2:00		3:00		
9	10 ⁻⁶ ml		10 ⁻⁶ ml		10 ⁻⁶ ml		2 x 10 ⁻⁷		3 x 10 ⁻⁷		
10									10 ⁻⁷		
11									10 ⁻⁷		
12	Time		+ 30'		+ 1 hr		+ 2 hr		+ 3 hr		
13	ml		+ -		+ -		+ -		+ -		
14	on Petal		0 0		0 0		0 0		0 0		
15	1	8	54	27	92(1)	84	224(7)	101	167(10)	61	62(6)
16	2	6	36	27	88(1)	90	238(6)	108	157(9)	69	88(5)
17	3	10	49	19	101(6)	78	211(12)	113	224(10)	68	76(6)
18	4	4	48	29	72(2)	84	258(4)	99	196(11)	103	74(8)
19	5	16	46	12	59(4)	104	229(3)	105	181(4)	60	59(3)
20	6										
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93	79										
94	80										
95	81										
96	82										
97	83										
98	84										
99	85										
100	86										

0 : actually counted number

10
11
12

Infection & Multiplication of Gal-F₂ segment

(This process ^{also} presents a possibility of dilution ~~inaccuracy~~)



Further experiments:

Make experiment on infection of F₂ within 1 hr. (growth of cells more exponential in this stage).

K&E SEMI-LOGARITHMIC 300-71
 KEUFFEL & ESSER CO. MADE IN U.S.A.
 3 CYCLES X 70 DIVISIONS

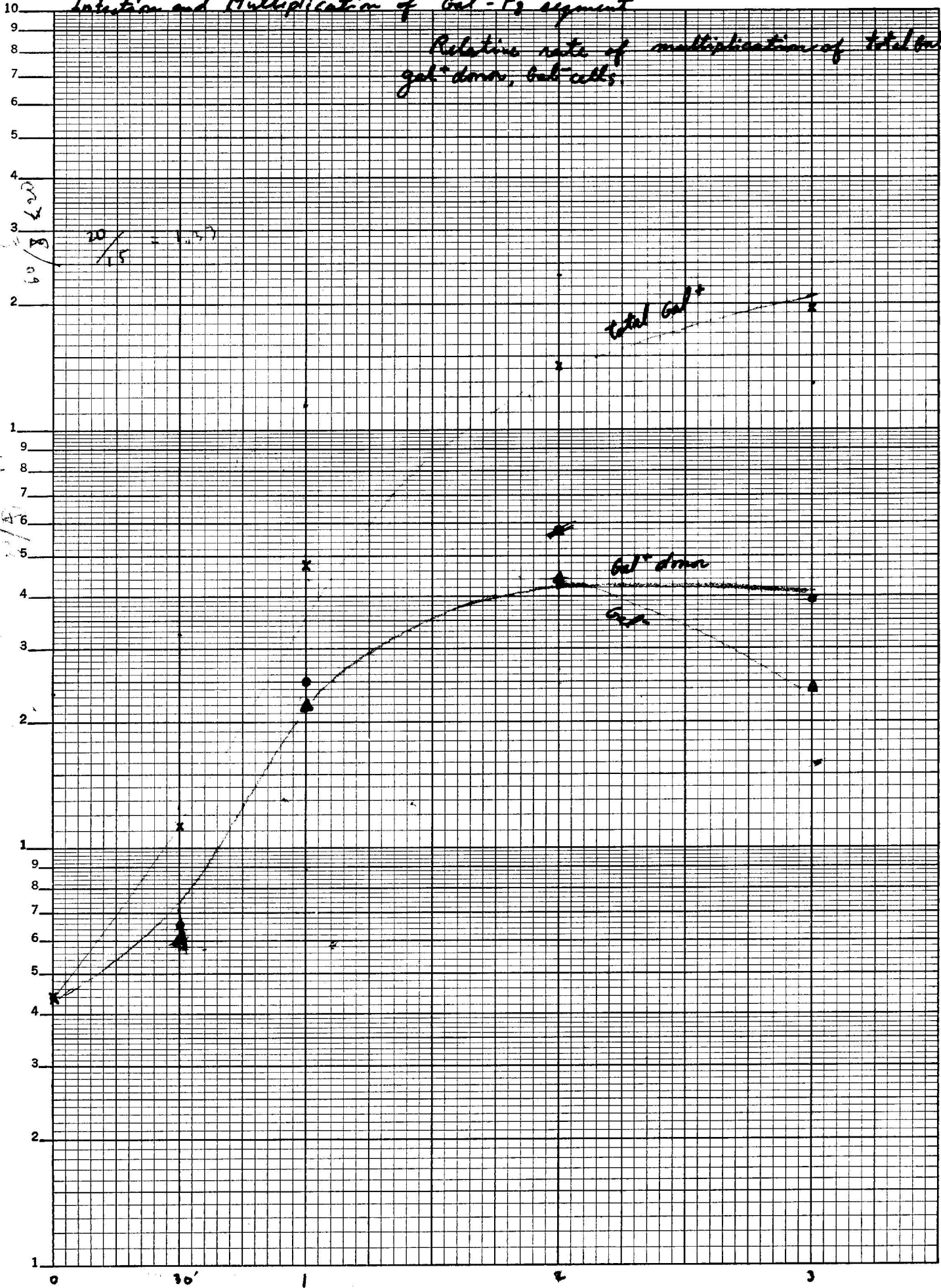
Intention and Multiplication of Gal⁻F₂ segment

Relative rate of multiplication of total Gal⁺, gal⁺ donor, Gal⁻ cells.

4.27
60/30

$$\frac{20}{15} = 1.33$$

KE SEMI-LOGARITHMIC 359-71
KEUFFEL & ESSER CO. MADE IN U.S.A.
3 CYCLES X 70 DIVISIONS

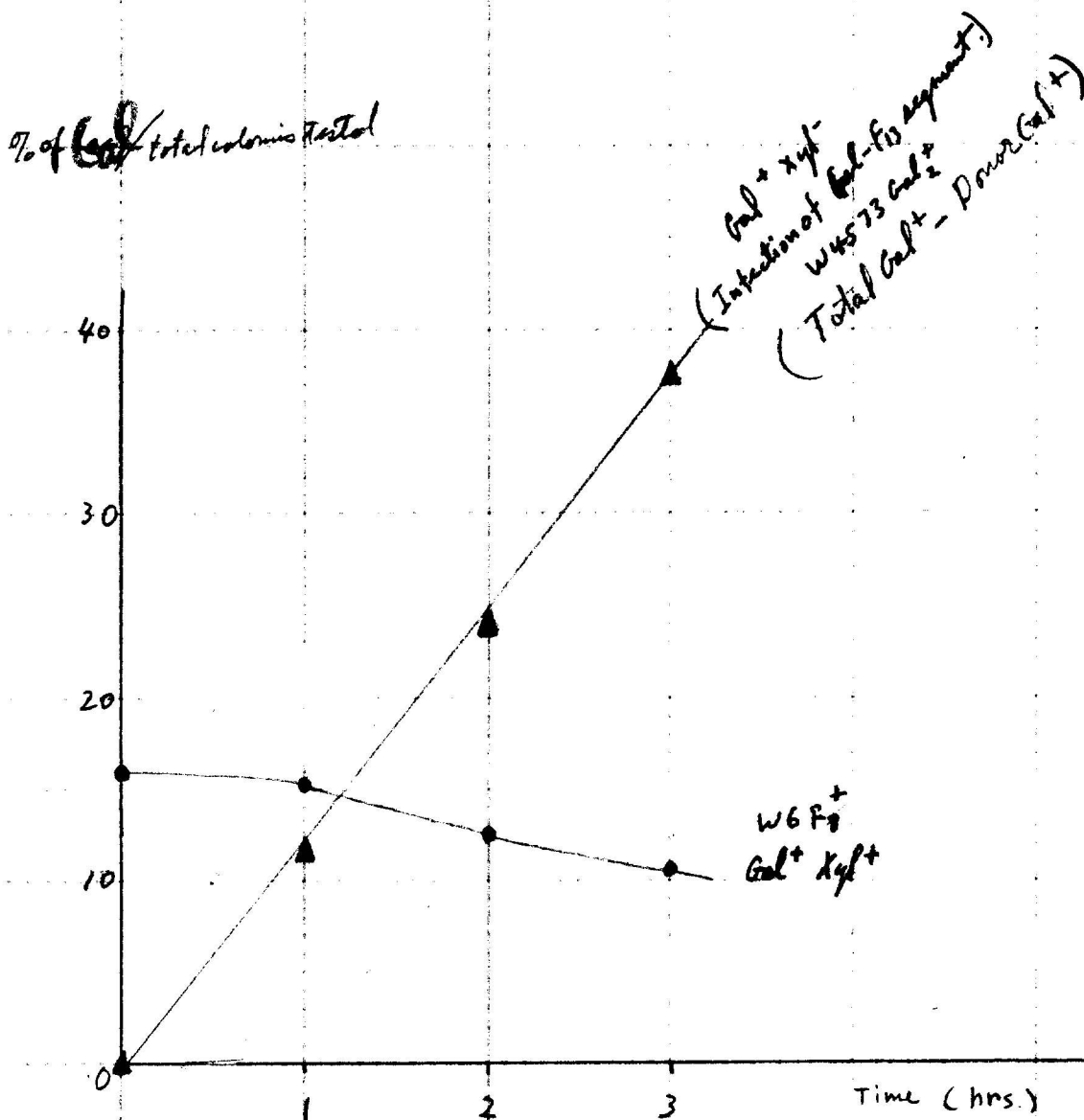


Rate of multiplication of Gal-F₈ segment.

1281

W6 F₈ → x W4573

37°C, on rotator,
in primary broth,
exponentially growing culture.



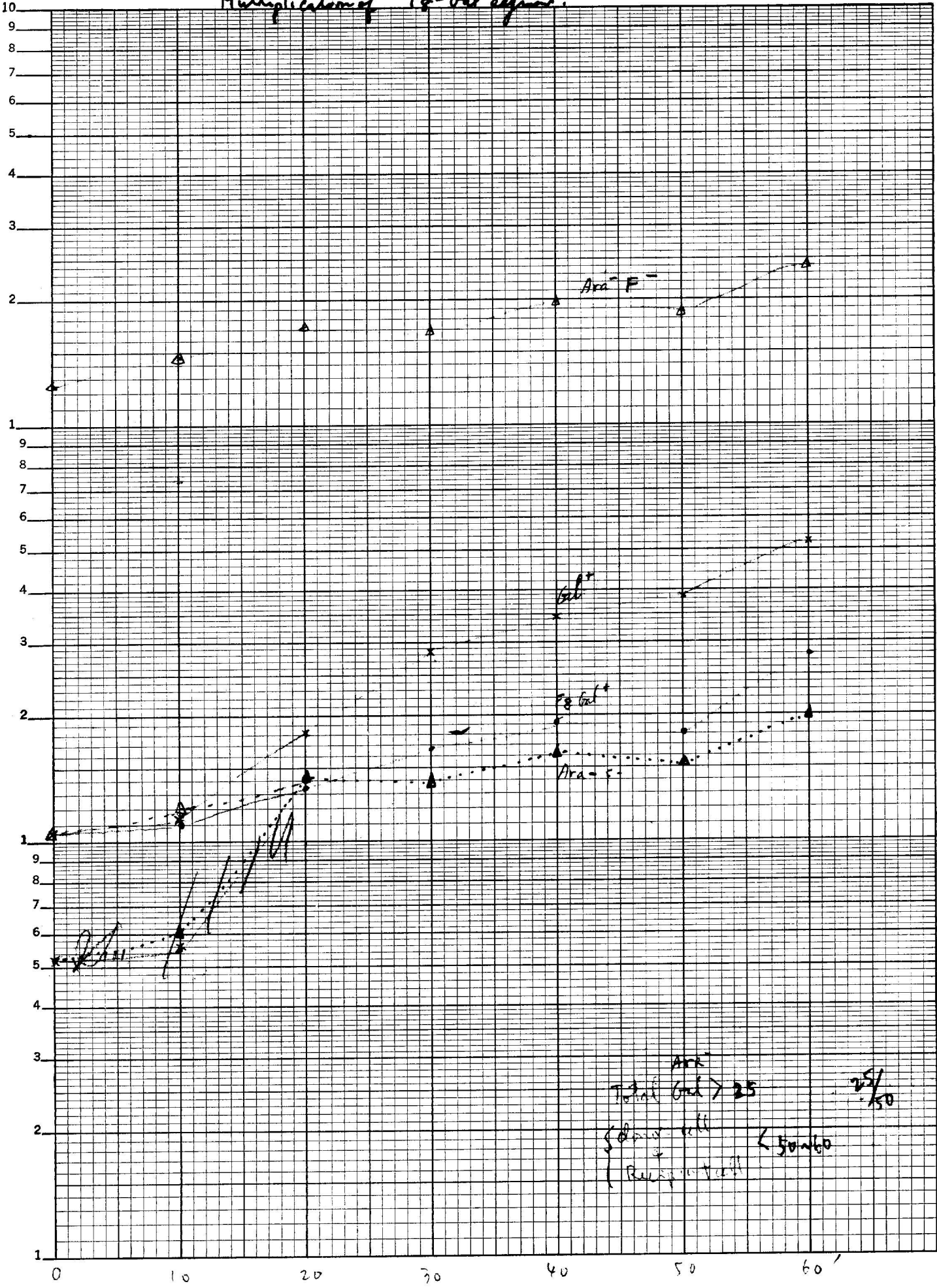
$\frac{F}{\text{und}}$

$\frac{F}{\text{und}}$

F₀

Multiplication of Fe-Gal segment.

SEMI-LOGARITHMIC 359-71
KEUFFEL & ESSER CO. MADE IN U.S.A.
3 CYCLES X 70 DIVISIONS



Segregation of *Lac* from Heterozygote for *Lac* segment.

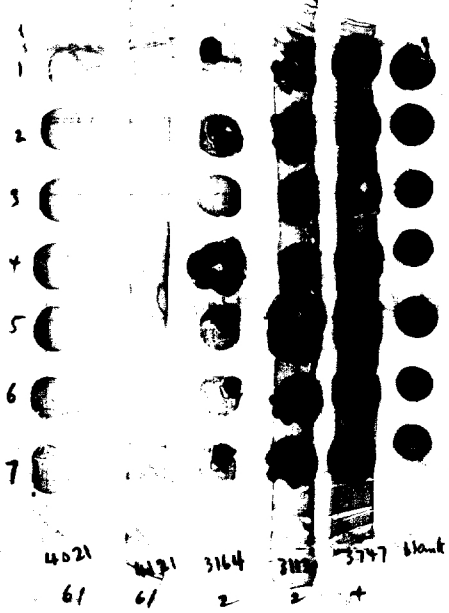
30/VII 1959

REF:

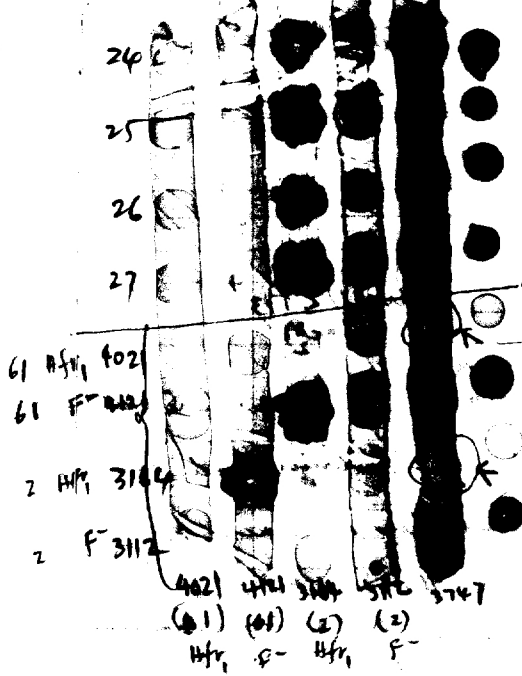
	1	2	3	4	5	6	7	8	9	10
1	(Lac 61)									
2	W4121 x W3112 F13 ⁺ (Took from spot test (see P.126))									
3	Lac ₆₁ (Lac ₂)									
4	Method: Streak									
5	1. Purify it on B _{lac} . (5 plates)									
6	2. Pick Lac ⁺ colonies and suspend them into 1ml H ₂ O. tubes.									
7	3. Restreak them on B _{lac} , and pick Lac segregant (one Lac from each ^{pick} streak).									
8	4. streak the isolated Lac on B _{lac} . (ca. 48 colonies are isolated) Some of them maybe Lac ⁺ .									
9	5. Test Lac x.A by cross streaking on M _{lac} . (27 were Lac ⁺)									
10										
11	V ₆	Lac mutation	F	x 3112 (Lac ₂) F ⁻	x 3164 3164 (Lac ₂) Hfr	x W4121 (Lac ₆₁) Hfr F ⁻	x W4021 (Lac ₆₁) Hfr		x 3747 (Lac ⁺)	
12		end. ex								
13	1	S	61	+	+	-	-	-	-	-
14	2	S	61	+2	+3	-	-	-	-	-
15	3	S	61	+4	+4	-	-	-	-	-
16	4	S	61	-	+12	-	-	-	-	-
17	5	S	61	#	+2	-	-	-	-	-
18	6	S	61	+2	+1	-	-	-	-	-
19	7	S	61	+4	+1	-	-	-	-	-
20	8	S	61	#6	-	-	-	-	-	-
21	9	S	61	#8	+1	-	-	-	-	-
22	10	S	61	#	+5	-	-	-	-	-
23	11	r/s	61	#	+4	-	-	-	-	-
24	12	S	61	#4	+3	-	-	-	-	-
25	13	S	61	-	+26	-	-	-	-	-
26	14	S	61	#6	-	-	-	-	-	-
27	15	S	61	-	+24	-	-	-	-	-
28	16	S	61	-	+8	-	-	-	-	-
29	17	S	61	#2	-	-	-	-	-	-
30	18	S	61	+3	+1	-	-	-	-	-
31	19	S	61	+3	+3	-	-	-	-	-
32	20	S	61	#	+2	-	-	-	-	-
33	21	S	61	+4	-	-	-	-	-	-
34	22	S	61	+10	+1	-	-	-	-	-
35	23	S	61	-	+29	-	-	-	-	-
36	24	S	61	-	#23	-	-	-	-	-
37	25	S	61	-	#3	-	-	-	-	-
38	26	S	61	-	#23	-	-	-	-	-
39	27	S	61	-	#34	-	-	-	-	-
40	Hfr 4021	Lac ₆₁	61	-	-	-	-	-	-	-
41	Hfr 4121	Lac ₆₁	61	-	-	-	-	-	-	-
42	Hfr 3164	Lac ₂	2	-	-	-	-	-	-	-
43	Hfr 3112	Lac ₂	2	-	-	-	-	-	-	-
44	Conclusion: 27/27 : Lac ₆₁ (100%)									

non-transmissible to Hfr.
Hfr. transmits to F₂.

Mlac



Mlac



control.

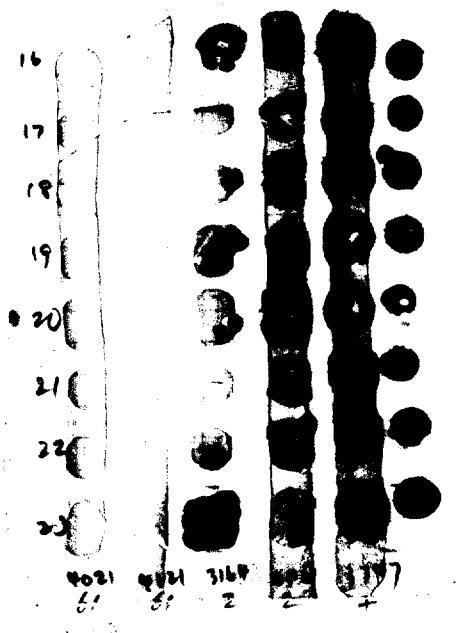
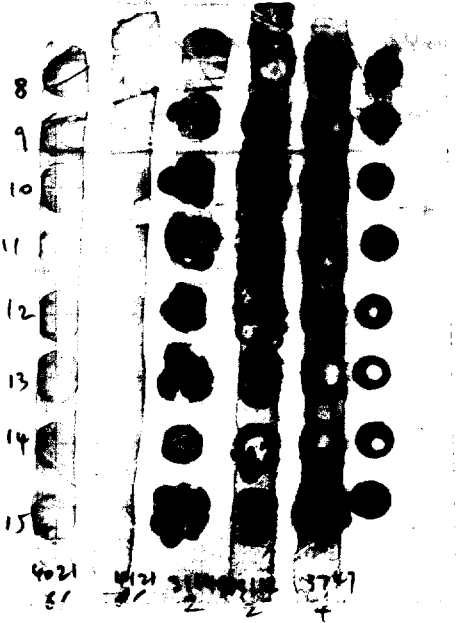
* is Hfr, x F' sterile!

4021 looks wrong! It may be a amp. resist.
(Lac⁺).

W4021 x Lac⁺ F-
is sterile.

Tests for Vi-low.

Retest # 11.



Segregants Analysis of Lac_2^{+}/Lac_61
F₁₃

7/1/59 ; 1959

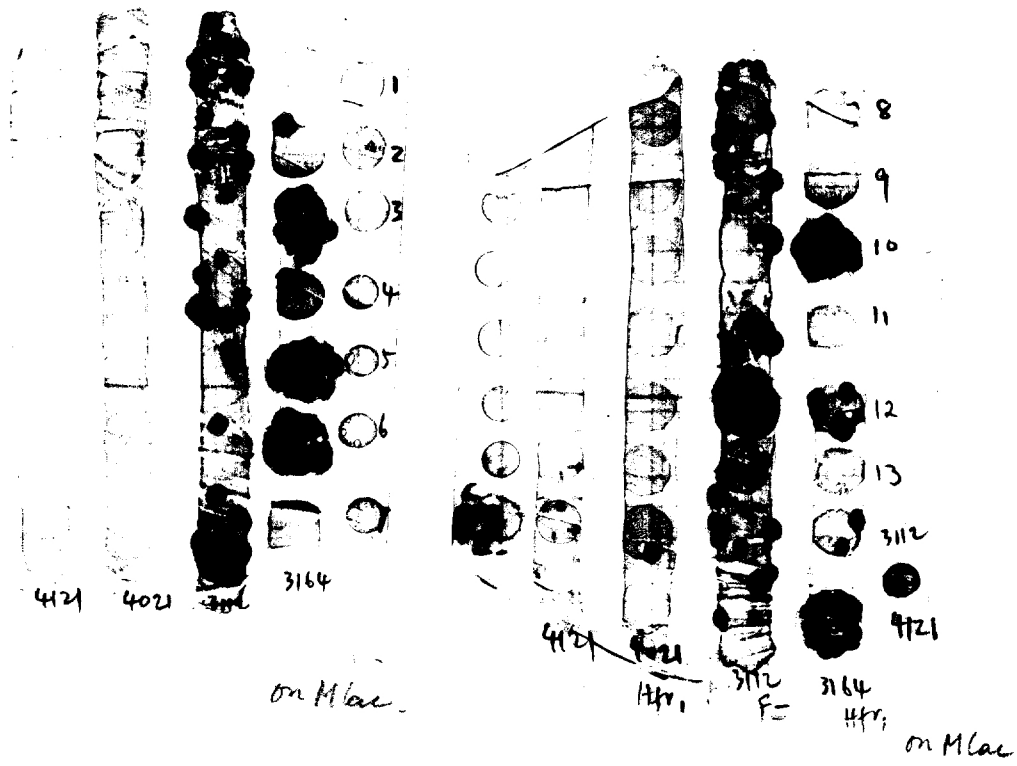
REF:

	1	2	3	4	5	6	7	8	9	10
1			W4121	x W3112	F ₁₃					
2			Lac ₆₁	Lac ₂						
3										
4										
5		Lac	F	x 3112	x 3164	y 4121	x 4021			
6		and ₆₁		(Lac ₂)	(Lac ₂)	Lac ₆₁	Lac ₆₁			
7		ex ₂		F	Hfr ₁	F ⁻	Hfr ₁			
8										
9	1	S	61	5	+20	-	-	-		
0	2	R	61	5	+60	+1	-	-		
	3	S	61	5	+2	+23	-	-		
1	4	S	61	5	+9	+1	-	-		
2	5	S	61	5	+2	+20	-	-		
3	6	S	61	5	+1	+32	-	-		
4	7	S	61	5	+23	-	-	-		
5	8	S	61	5	+11	-	-	-		
6	9	S	61	5	+5	-	-	-		
7	10	S	61	5	+2	+28	-	-		
8	11	S	61	5	+8	-	-	-		
9	12	R/S	61	5	+4	+9	-	-		
0	13	S	61	5	+4	-	-	-		
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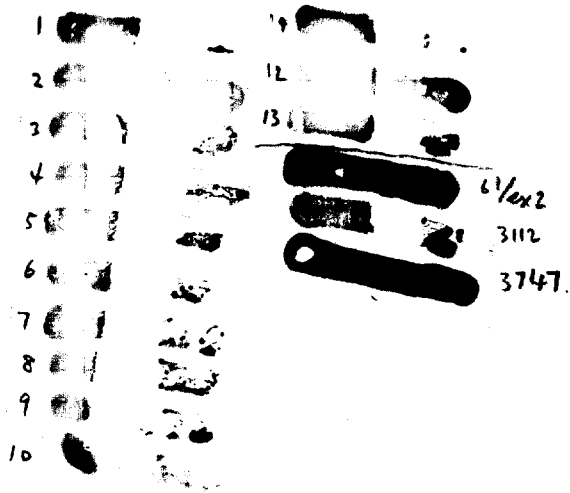
See back page

conclusion : 13/13 : Lac₆₁ (100%)

25
13
27
65



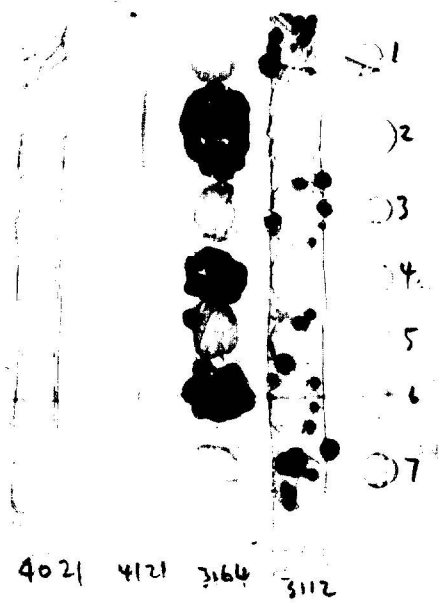
Tut of *V₆ loci*



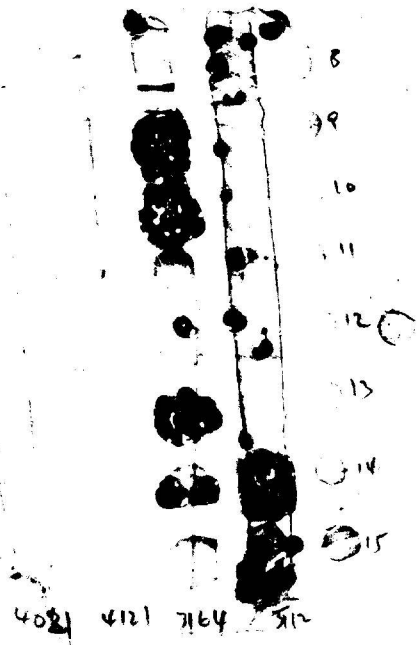
on Blue

Retreat # 12.

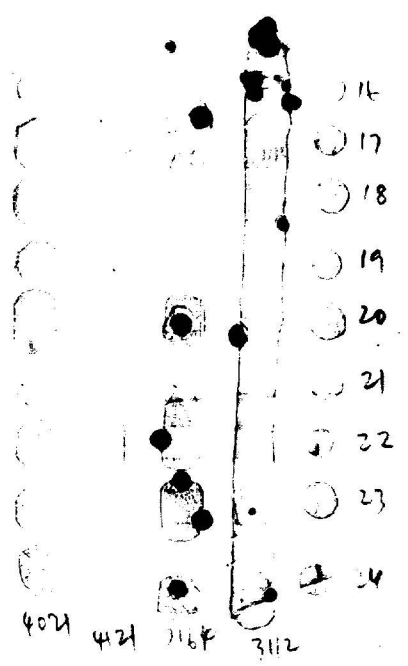
This strain looks F13⁺ Lac₆₁^{V₆S} / Lac₆₁^{V₆R}



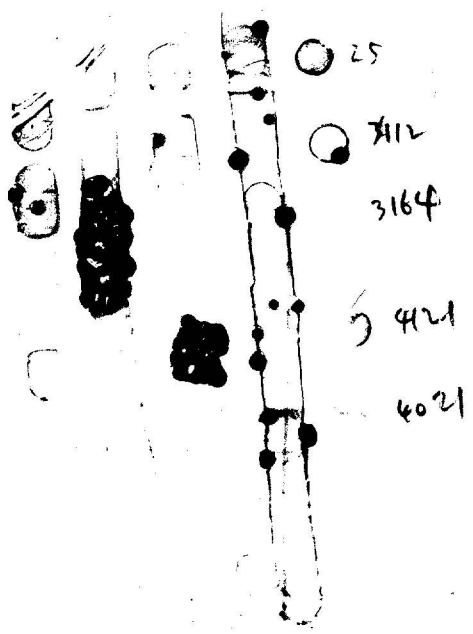
M lac



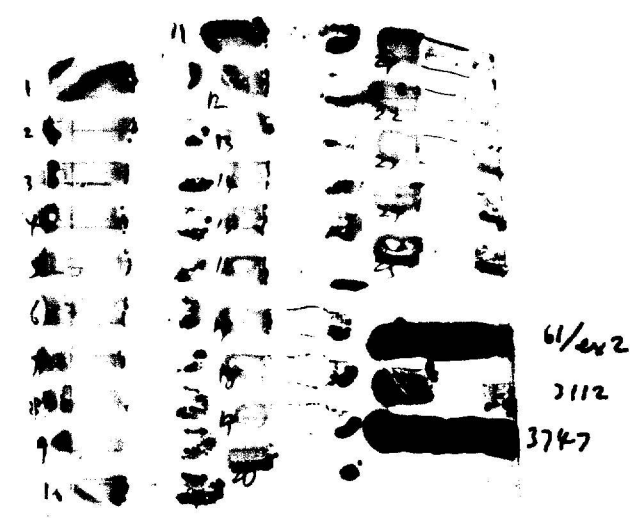
M lac



M lac



Test of V_6 loci



on blue

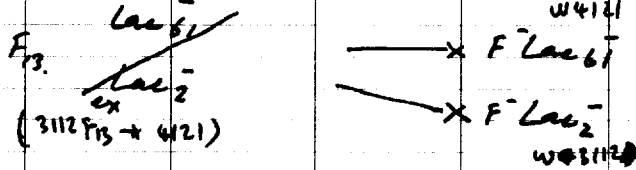
Retest # 14
This maybe V_6^3 lac 61 / V_6^5 lac 61

Unsuccessful: use M Lac Sm in next time, (Lac⁶¹/lac² does grow on M Lac)

Comparison of the ~~trans~~ frequencies of the transduction of exogenote and endogenote from heterozygous diploid (Lac⁶¹/lac²). cf. 105, 146

4/24, 1959

Principle



cultural age: 4 hrs. on rotator.

Strain: all three strains used as source purified on B Lac and Pict single colony and expanded into B (Ca 10⁷) 0.1 ml is inoculated into 10 ml of M Lac and kept on rotator for 4 hrs.

- Method:
- Mix Lac⁶¹/lac² and Lac⁶¹ or Lac² use excess F⁻, Ratio, F₃ 1 : F⁻ 10 : 0.1 ml 10.0 ml
 - Inoculate them for 1/2 hrs. ~~incubation~~ at 37°C. 3:25 pm
 - Seed them on M Lac agar after optimal dilutions. Use: 10⁻³ ml, 10⁻⁴ ml, 10⁻⁵ ml per plate for each crosses.
 - Count cell numbers of each experiment.

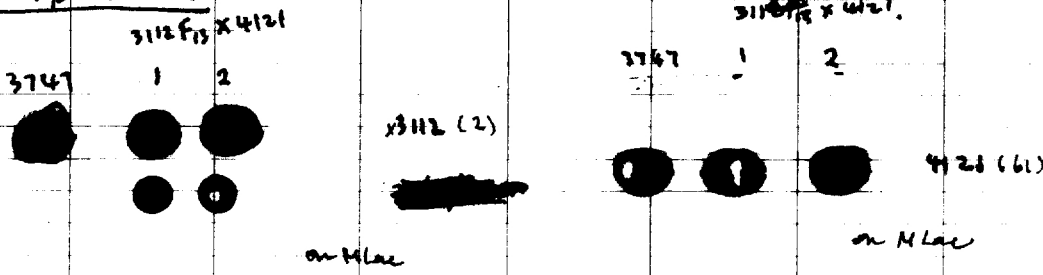
Results:

Survival count	① F ₃ 61/2 x lac ² W3112		② F ₃ 61/2 x lac ⁶¹ W3112		③ F ₃ 61/2 x lac ² W3112		④ F ₃ 61/2 x lac ⁶¹ W3112	
	Lac ⁻	Lac ⁺	Lac ⁻	Lac ⁺	Lac ⁻	Lac ⁺	Lac ⁻	Lac ⁺
10 ⁻⁶ ml/plate	1057	89	1084	81	944	51	868	85

fertility on 10⁻¹ ml/plate too much, it cannot count them. (Use 1/100 of these experiments)

10⁻³ ml/plate 1084 81 944 51 868 85

Quantitative experiment.



11/VIII 1959

REF: cf. 145, 146

Principle:

Lac⁶¹/ex Lac² S^s → Lac⁶¹ F⁻ S^R W4121 S^R (W4629)
 W4121 (Lac⁶¹/ex Lac²) → Lac² F⁻ S^R W3112
 compare the fertility in both crosses.

check S_m-sensitivity on B Lac S_m of these strains.

Result:  on B Lac S_m

Qualitative test. Preliminary test by spotting method

Spot 4121 (61/ex 2) on W4121 S^R and W3112 streaked on M Lac S_m.

Result: see back page.

	(Lac ²)	(Lac ⁶¹)	Blank
4121 61/ex 2	+	++	
3112 2/ex 2	+ reversion	++	+ reversion
3747 +/ex+	+++	++	

Quantitative test.

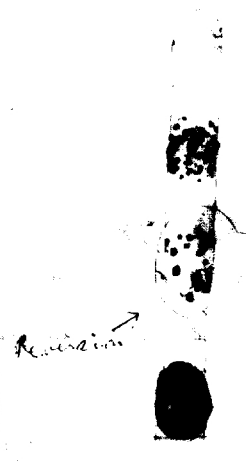
- cultural age: 0.2 ml: overnight culture / 5 ml phagey
- Ratio of mix: 1 ml 3112 + 4121 Lac⁶¹/ex 2
1 ml 4121 S^R (W4629) + 4121 Lac⁶¹/ex 2
- Time of incubation: ~~1 hr~~ 2:30 - 3:30
- survival count: 10^6 m³ / plate
- Inoculum size for counting recombinations: 10^3 ml. / plate on M Lac S_m.

Result:

	# of Recombinants. / Plate	# of Colonies / plate		% of Rec.
		10 ⁻⁸ ml	10 ⁻⁷ ml	
3747 +/+ x 3112	too much	~ 10 ³	~ 10 ³	
3747 x 4629	~ 10 ³	~ 10 ³	~ 10 ³	
4121 61/2 x 3112	0	0	0	~ 10 ⁻³
4121 61/2 x 4629	2	5	5	~ 10 ⁻³

Conclusion: Exogamete is transferred with very high frequency, but not very high for endogamete. This implies all F₁ Lac H₁ clone are diploid in that (Lac) segment. And also speculate that linkage between F₁ and endogamete, and exchange of end-ess fragment are relatively low.

F⁻ Lac⁺ F⁻ Lac⁻

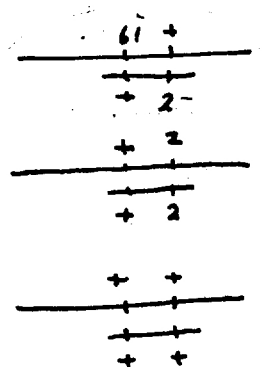


4121 (61/ex2)

3112 (2/ex2)

3747 (+/ex+)

↑ Reversion



W3112
2 F

W4215A
61

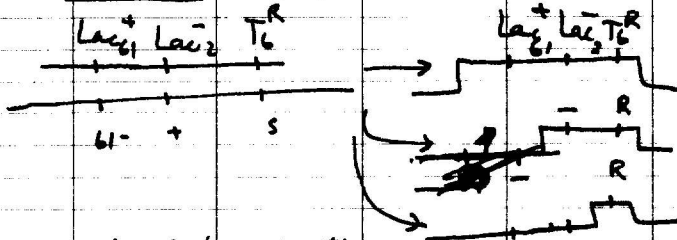
on P/Lac⁺ Sam

Linkage of T_6^R and lac_2^- in exogenote of $(Lac_2 T_6^R \times Lac_1 T_6^S)$

6/11/59 1959

REF:

Principle. Segregant (non-recombined) is lac^-



The rate of recombination must express ratio of length of $V_6 - Lac_2 - Lac_1$.

If Lac is closely linked with T_6 , lac^- can be found at the $Lac_1 T_6^S / Lac_2 T_6^R$ by T_6 .



3112 F13

3112 F13 x 4121

①

②

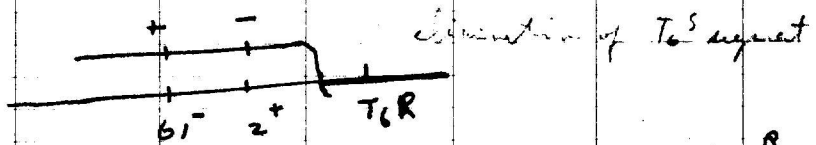
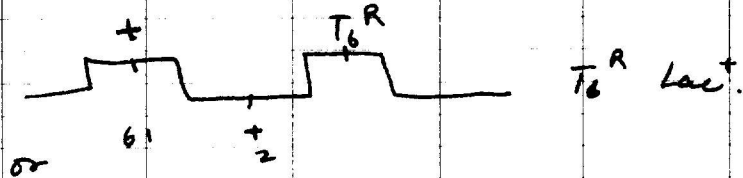
4112 (52)

3747

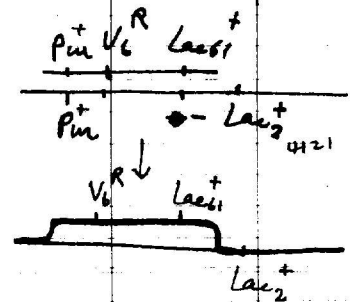
T_6 on Bla^c

Result: actually the lysed part is also lac^- .

Explanation: There are enough number of segregants which arise by recombination between two different lac^- mutations. Their fermentation of lactose may cover the presence of lac^- colonies.



or $Lac_2 F13$ is hemizygote for lac_2 .



Comparisons ^{on} the stability of maleness of F⁺ and F['] to AO treatment.

8/44 ; 1969

REF:

	1	2	3	4	5	6	7	8	9	10
									Make opposite combination	
1	Strain	W3086 F ₈		W3747		W6			W6 F ₈	
2				F ₈		F ₈			3086 F ⁺	
3										
4		cultural age : overnight culture in Phoscopy broth.								
5		Inoculum size : ca. 10 ⁸ cells/ml. 10 ² x 10 ⁻² x 10 ¹ (0.1 ml) / 5 ml								
6		AO-treatment : AO-medium : 30g/ml ; NSB pH. 7.6.								
7										
8										
9										
10										

Result :

A). Survival count on B Mal. 10⁻⁶ ml / plate. (before treatment.)

1	Exp II	A).	Mal ⁺	Mal ⁻	Exp II	compatibility		
2			W6 F ₈	3086 F ⁺		total W6 F ₈		total 3086 F ⁺
3						total		F ⁻
4			234	89		44		93
5			327	152		70		
6			227	107				
7								
8								
9								
10								

B). Survival count After treatment. (After treatment.)
Treated with AO. Replicated on M Gal (x 4573.)

1	Exp I	3086 F ₈ + W6	W3086 F ₈	W6 F ⁺	Conclusion: Looks no difference in availability of F ₈ to acidine method.			
2					Untreated control.			
3			Σ F ₈ F ⁻	Σ F ⁺ F ⁻	3086 F ₈	W6 F ⁺		
4			35 4 31	35 5 30			Σ F ⁺ F ⁻	
5			21 1 2 19	24 1 23			55 30 0	
6					Σ F ₈ F ⁻	Σ F ⁺ F ⁻		
7					55 55 0	55 30 0		
8			Σ 56 6 50	Σ 59 6 53				
9			% 89.3 F ⁻	89.9 % F ⁻	% 100 0	55 100 0		
10								

Exp II W6 F₈ + 3086 F⁺. Untreated control. Treated experiment (with AO: 30g/ml)

1		W6 F ₈	W3086 F ⁺	W6 F ₈	W3086 F ⁺
2		Σ F ₈ F ⁻	Σ F ⁺ F ⁻	Σ F ₈ F ⁻	Σ F ⁺ F ⁻
3		32 32 0	32 0	0 59	0 42
4				% 0 100	0 100

x 4573 on M Gal.

x 4573 on M Gal.

Exp (I).

Knowledge Co. 10² cells/line

AD 308 / unit
6 units per pt

AD 3086F8 + W6

3086F8

W6

①



x 4573
on M6Gal

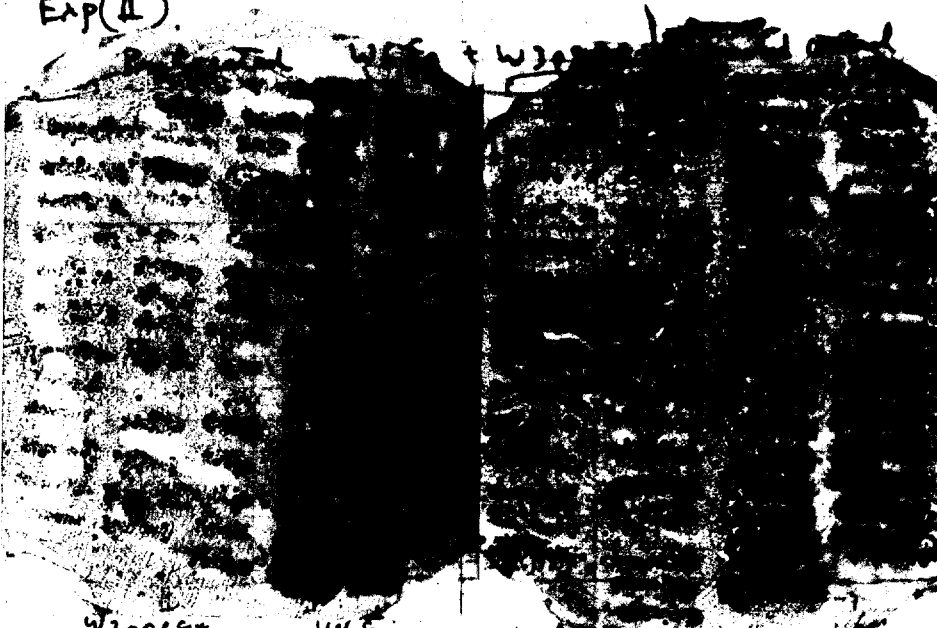
W6

3086F8 ②



x 4573
on M6Gal

Exp (II)



W3086F8

W6F8

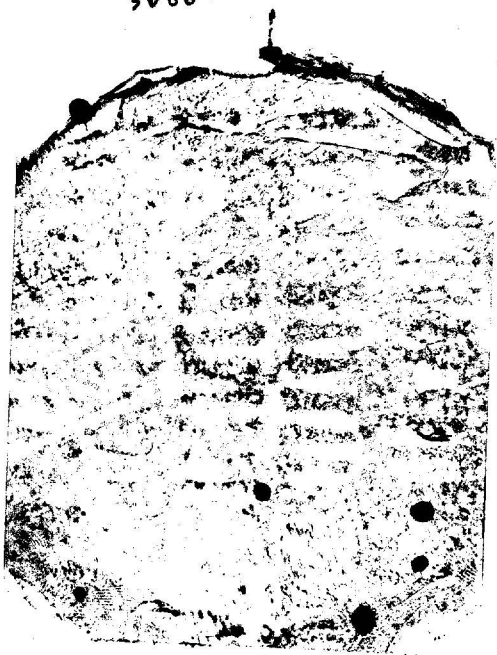
x 4573
on M6Gal

W3086F8

W6F8

on M6Gal
x 4573

40 treated
3086 ft w6FB.



x 4573
on M6ul

40 treated.
w3086ft w6FB



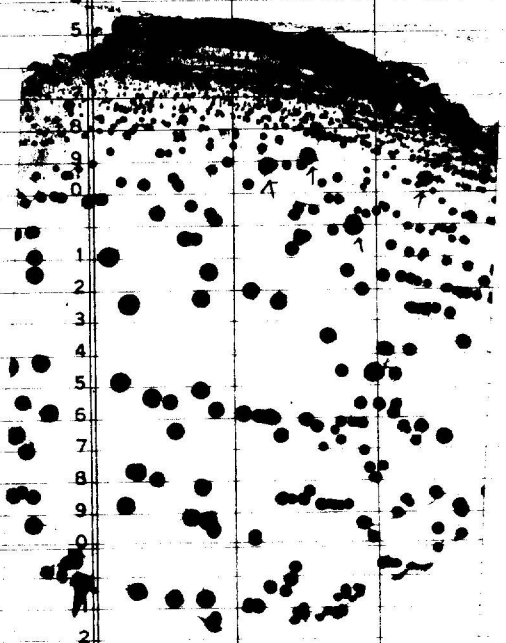
on M6ul
x 4573

Size of Gal-F₈₀ segment.

12/VIII; 1959

REF:

	1	2	3	4	5	6	7	8	9	10
1	Strain used: ♀. Pur ⁻ Try ⁻ Gal ₂ ⁻ Xyl ₂ Met ¹ Mal ₁ Ara ₂ S ^R F ⁻									
2	♂ F ₈ M ⁻ (W6 F ₈)									
3	Cultural age: 0.2 ml: overnight culture / 5 ml Penassay broth.									
4	↓ 2 hr on rotator at 37°C.									
5	10:45 ~ 12:45 AM									
6	Rate of mix: 10: F ₈ 0.1 ml F ⁻ → Incubate it overnight at 22°C									
7	Principle: W6 F ₈ → x F ⁻ Pur ⁻ Try ⁻ Gal ₂ ⁻ S ^R									
8	Select on B Gal Sm. B Gal Sm.									
9	Pick Gal ⁺ and test the other markers.									
10	Result: Very few colonies become Gal ⁺ It may be come from <u>pur⁻ Try⁻</u> of the recipient strain.									



on B Gal Sm.
5 days
Gal⁻ becomes black

Unsuccessful

Infection of F_3 and Remove of F_1 from it.

20/04

1959

Principle : $F_3 - F_1 = Rfd_3$

REF: W4296 shows strong syntrophy [with M₁ see back page 10]

Principle : W4616 ($lac^- F_3$)

W4295 $F^- L^- SR$

W4296 $F^- T^- SR$

↑ Too much syntrophy.

W4616 → x W4295.

F_3	Total Colonies tested
20	350
15	379

o cultural age : 2 hrs. 0.2ml / 5ml.

• excess F_3^+ : 10:1. F^-

• Ratio: (2ml 0.2ml)

• Purified on Mlac Sm.

• Replicated on Mlac Seeded W3086.

Result: The first step.
3133 F_3 → x 4295

Retest of newly isolated F_3^+
by spot test. X 3086 on Mlac.



on Mlac
~~W3086~~
W3086

on Mlac
X3086

Try it again

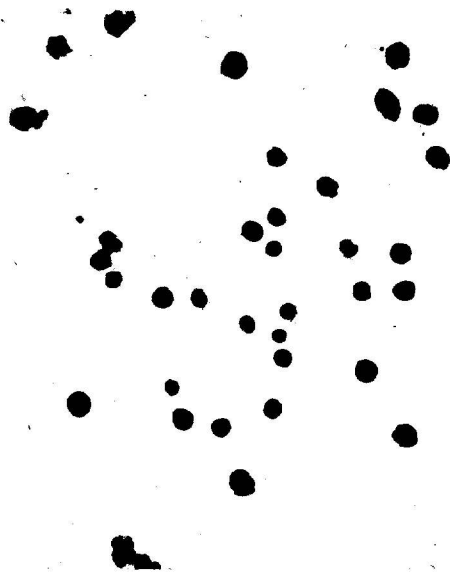
Syntrophy of M and T.

3137f₃ → w4296



~ Mlac
w884407.

Martin plate



m8lac

Isolation of $Lac^- F_{13}$ by ~~simple~~ ^{simple} infection of F_{13}

number 139

21/VIII

1959

3747 ——— X 4121^{S^R}
M V₆ F₁₃ (Lac⁺ F⁻ S^R)

REF: cf P120 & P120a
P. 55

1	2	3	4	5	6	7	8	9	10
---	---	---	---	---	---	---	---	---	----

Purpose: Still it is very hard to isolate $F_{13} Lac^-$ strain from segregants.
This experiment is carried out to find the most more easy method.

Principle: 1. F_{13} may split off from Lac locus and infect to Lac^- . (This possibility is very low)
2. $Lac^+ F_{13}$ consist heterozygous diploid state in first step, then recombination occurs, and endogenous Lac^- becomes exogenous. This $Lac^- F_{13}$ becomes infective, and multiplies rapidly.

In any way, if it does occur, it may be possible to isolate $Lac^- F_{13}$ using this method.

Result:

1. Rate of infection of $Lac^- F_{13}$ into 4121^{S^R}.

Lac^- : 91 17.2 %
 Lac^+ : 438 82.8 %

2. Rate of $F_{13} Lac^-$.

Lac^- isolated	F^-	F_{13}
108	108	0

~~method~~ see below.

Conclusion: F_{13} itself does not infect into F^- in this experiment.

This ratio may be much less than 1 %.

Method: 1. Make 2 hrs culture (shaked on rotator in phassy at 37°C.)

2. Mix 10 : 1 : 10
 F_{13} : F^- : fresh broth.

2 ml, 0.2 ml, 2 ml of fresh broth.

3. Incubate it for overnight.

4. Purify it on Dlac Sm.

5. Pick Lac^- colony on Dlac Sm.

6. Replicate on Mlac, on which $Lac^- F^-$ are seeded.

Isolation of $Lac61/ex.Lac61.F13$.

24/VIII, 1959

REF: Lac_2 must be S^5

Principle:

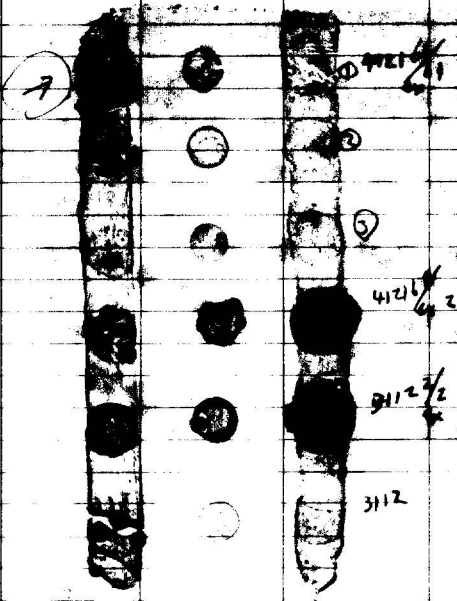
1st step: $Lac_2/ex.Lac_2.F13 \rightarrow Lac61 F^- \rightarrow Lac61/ex.Lac_2.F13$

2nd step: $Lac61/ex.Lac_2.F13 \xrightarrow{\text{exchanging Promos of exogenote and endogenote}} Lac_2/ex.Lac61.F13$
 (Detect it by replica plating method) see back page.

3rd step: $Lac_2/ex.Lac61.F13 \rightarrow Lac61 \rightarrow Lac61/ex.Lac61.F13$

Principle is the indirect selection of a clone which contains many cells of $Lac_2/ex.Lac61$ arising by exchange of Lac locus from $Lac61/ex.Lac_2$.

Retest



Replicated from Blac.

1st selection.



on MLac
x 3112.

4121 $\frac{61}{6x2}$

3112 $\frac{2}{2}$

3747 $\frac{2}{2}$

4121 $\frac{2}{6x51}$



Pick.

Repurify

← Replicated

4629

Lac 61.
SR

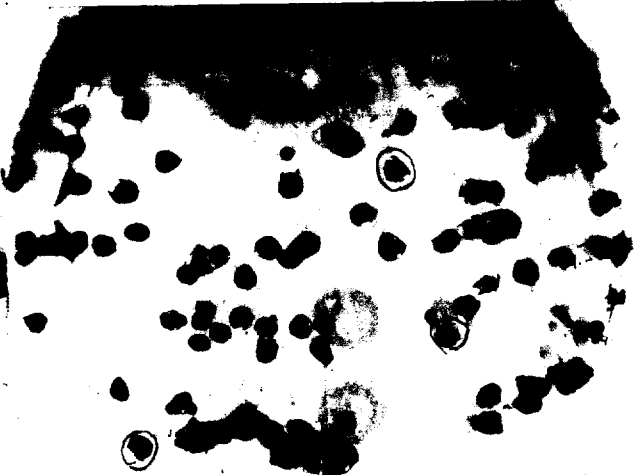
3112

Lac 2 on MLac SR

2nd selection.

on Blac

on MLac



x 3112.

4121

3112



4121

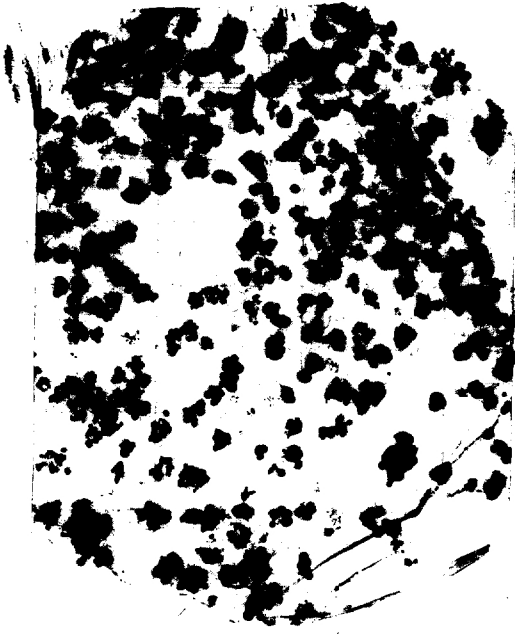
$\frac{61}{6x2}$



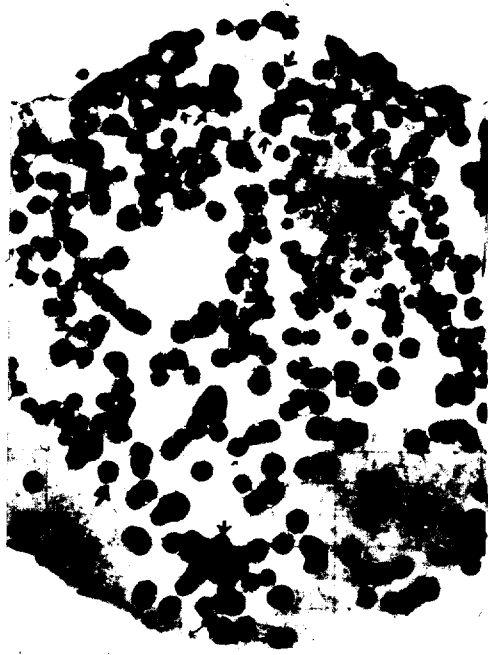
4629



3112



MGal.
x 2979.



on Blac

Comparison of the fertility of W3747, H1Gal and H1Lac.
I II.

1959.

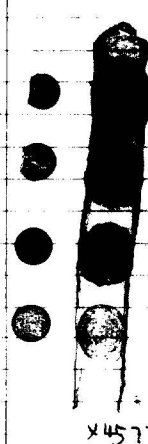
REF:

Purpose: W3747 shows two kinds of colonies, one is high fertility to

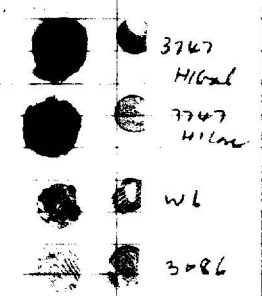
1
2
3
4
5
6
7
8
9
0
1
2
3
4
5
6
7
8
9
0
1
2
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0
1
2
3
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6
7
8
9
0
1
2
3
4
5
6
7
8
9
0



4074 T₄
4293 A₁
2984 P
1394 TLB₁
3949 A₃
4573 Lac
4506 pur
- Syntrophy?
- Recomb.



3747
H1Gal
H1Lac
W6
3086
x4573
on MGal



3747
H1Gal
3747
H1Lac
W6
3086
x4573
on Mxyl

H1Gal H1Gal
3747 3747 W6 W3086-

Conclusion

on Mlac
W1
H1gal TLB₁ A₁ T₄
H1Lac Pur

W6
Lac Pur
gal TLB₁ A₁ T₄

Order of integration of chromosome via ~~cross~~ cross F₂ x F⁻

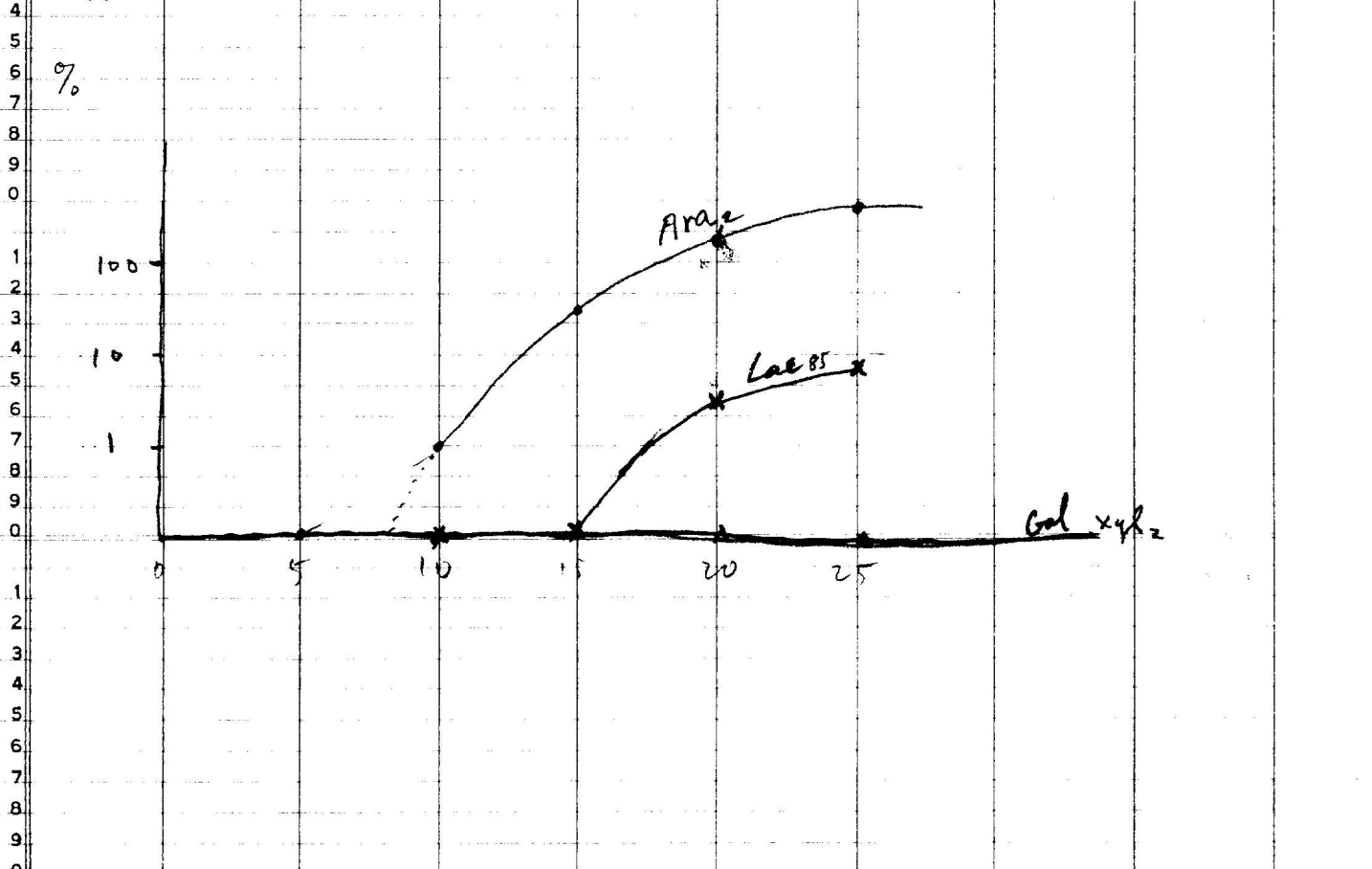
26/VIII 1959

REF:

	1	2	3	4	5	6	7	8	9	10
		W454 ³								
1	Strain: W6 F ₂		W4573							
2	M ⁻ F ₂		Ara ₂ Lac ₈₅ Gal ₂ S ^R Mal, Xyl ₂ Mt1.							
3	Blending: Gage: 60,		1 min.							
4										
5										
6	overnight culture		0.1 ml / 5 ml pen →							
7	Culturing age: 3:30 ~ 4:00		2 1/2 hrs. at 37°C on rotator							
8					W6 F ₂ : 2 ml					
9					W4573: 0.2 ml					
10										
11	# of Recombinants/plate									
12	Time	0	5	10	15	20	25			
13	plate # 1	Ara ₂ Lac Gal Xyl	A, L, G, X	ALGX	ALGX	ALGX	ALGX			
14	0-3 ml	0 0 0 0	0 0 0 0	1 0 0 0	9 0 0 0	45 1 0 0	51 0 0 0			
15	2	0 0 0 0	0 0 0 0	0 0 0 0	12 0 0 0	33 1 0 0	57 5 0 0			
16	Σ	0 0 0 0	0 0 0 0	1	21 0	78 2	108 8			

Survival count	Lac	+	-	+	-	+	-	+	-	+	-
10 ⁶ ml		4	54	6	64	8	51	3	73	6	71
		3	27	5	67	6	76	7	63	9	93

fertility %
ca. 0.1% ca. 0.1% ca. 0.5% ca. 1%



Comparisons of the fertility of exogenous and endogenous segmt.

11959

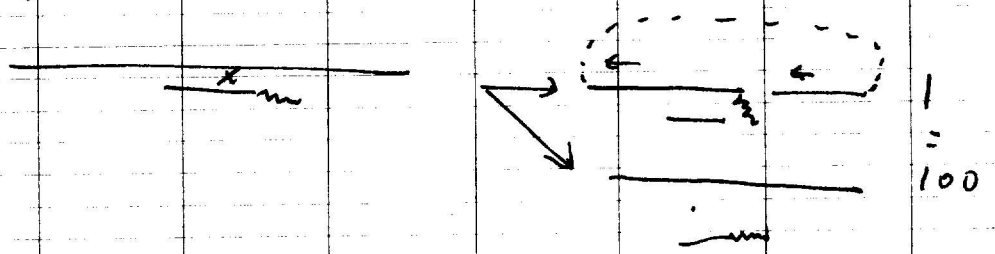
REF:

	1	2	3	4	5	6	7	8	9	10	
	Cultural age: overnight culture.										
						fertility		Survival (10^6 ml)			Rec./Host
						10^{-3} ml	10^{-2} ml	Lac ⁺	Lac ⁻	+ -	
1	W4121	61/ex 2			W4629	79	1214	8	38	8 71	
2						110	1097	5	45	8 75	
3						Ca. 100	Ca. 1000	Ca. 5×10^8	4×10^7		Ca. 10^{-2}
4											
5	W4121	61/ex 2			W3112 (F ⁻ Lac ₂ S ^R)	0	12	1	27	18 156	
6						0	20	3	32	23 155	
7						0	Ca. 10 ¹⁰⁻²⁰	Ca. 2×10^8	Ca. 3×10^7		< Ca. 10^{-5}
8	-----										
9						10^{-3} ml	10^{-2} ml	+	-	+ -	
0	W3747	4/ex 2			W4629	1231	68	248	35	124 14	
1							101	262	79	138 20	
2						Ca. 1000	Ca. 100	262	41		Ca. 10^{-2}
3								Ca. 2×10^8	4×10^7		
4											
5	W3747	4/ex 2			W3112	1261	52	170	46	240 70	
6							113	262	79	291 68	
7						Ca. 1000	Ca. 100	Ca. 2×10^8	5×10^7		Ca. 10^{-2}

Conclusion:

Fertility of
exogenous \cong endogenous $\times 100$.

This may be explained by crossing over between F' and host-chromosomes about 1%.



Transformation of Gal in k-12.

20/11. 1959

REF:

Experimental condition:

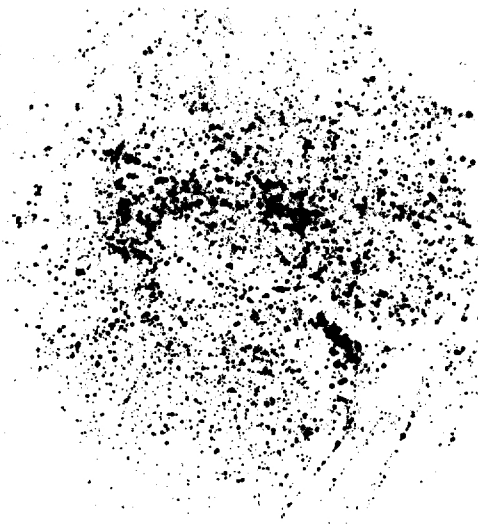
- 1 1 Conc. of Dugard (Na-lauryl sulphate) : 10%.
- 2 2 Time of Treatment : 3 hrs at 37°C. - H₂O.
1:45 ~ 4:45.
- 3 3 Bacterial culture : overnight; ca 10⁸ ~ 10⁹ cells/ml. washed once with H₂O.
re-purified, grown. suspended into H₂O.
32 mls of ^{wbfb} culture was used. (4 tubes of 8 ml per.)
↓
concentrated into 2 mls.
↓
1 ml / treated with dugard. final 10%.
20% dugard soln 1 ml +
Bact. suspension 1 ml
↓
1 ml / untreated
H₂O 1 ml +
Bact. susp. 1 ml.

4. Method of transformation.

DNA	dilution of original culture	10 ⁻¹	10 ⁻²	10 ⁻³	10 ⁻⁶
• # of recombinants.		521	521	43	
• survival cells in 0.5 ml		-	-	2,5 ↑	
Bacteria		10 ⁻¹	10 ⁻²	10 ⁻³	10 ⁻⁶
• # of recombinants					41
• survival cells in 1 ml		-	-	-	435 585 ↑

Next step: Filter this DNA sample.

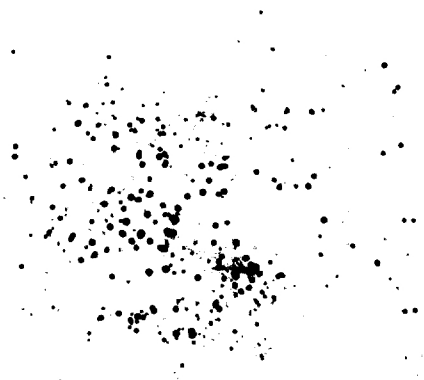
DNA 10^{-1} dil.



Intact.
 10^{-2} dil.



DNA 10^{-2} dil.



DNA 10^{-3} dil.



Transformation of F₈ Gal.
(filtered material).

Negative result.

23/ix 1959

REF:

	1	2	3	4	5	6	7	8	9	10	
1	Isolation of Sample.	1. W6 F ₈	M- F ₈			W4573	La ₉ Gal ₂				
2		4 tubes, 8 ml. → suspend in 4 ml of H ₂ O → add 4 ml of deposed 20%.									
3		2. Ex treatment with deposed: Na-borohyd-sulfate.									
4		(final conc. 10%.)									
5		in dist. W.									
6		37°C 4hr.									
7		3. Filtered through millipore filter.									
8		4. Mix:									
9		0.2 ml of W4573 + 0.2 ml of broth + 1 ml of DNA.									
0		: old culture (2 days) deposited sample.									
1	and incubate it for 30'.										
2	5. Seed it on M Gal.										
3	Result:										
4	1) survival count: 0.2 ml of the extract was seeded onto EMB/acc.										
5	No colonies are observed.										
6	2) Recombination activity.										
7											
8	Extract : 1 ml (1/10 diluted) (1/100 diluted)										
9	0.1 ml 0.01 ml										
0	<hr/>										
1	# of Recombinations. 0 0 0										
2	Conclusion:										
3	Mating activity of W6 F ₈ is in insoluble part.										
4											
5											
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7											
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0											

Action of propamidine isethionate to F and F' (E)

(P.I.)
4:4'-diamidino-diphenylpropane-di(β-hydroxyethane sulphonate) 5 gr.

May & Baker Ltd. Dagenham England.

16/ix. 1959

Joshua gave me. on 11 (sept. '59.

1	2	3	4	5	6			
1	Stock solution: 100 mg / 1 ml soluble in water.							
2	(Weigh 100mg of P.I. and dissolved into sterile H ₂ O - 1 ml.)							
3								
4	1. Test survival & bacteriostatic concentrations of P.I.						Control.	
5								
6	Conc. of P.I.		0.5 mg/ml	1 mg	100 μg	10 μg	1 μg.	
7	(cc. of stock sol.)		0.25 ml	0.05	10 μg	1 μg	0.05	
8	W6 Fg		-	-	-	+	+	
9	W6.		-	-	-	+	+	
0								
1	make survival stability test of P.I. soln.						100 μg	10 μg.
2	Media 5 ml Penassay broth, pH. 7.0.						-	-
3								
4	2. Streak these treated cells on Bhae agar.							
5	100 μg. P.I.							
6								
7	3. Test their compatibility by cross-brushing method.							
8								
9	Result:							
0	P.I.-treated			untreated control.				
1	100 μg/ml Km.							
2	F ⁻	F ⁺	70 F ⁻	F ⁻	F ⁺	70 F ⁻		
3								
4	7	24		3	20			
5								
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7								
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9								
0	W6 P.I.			W6 Cont.				
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24/10, 1959

REF:

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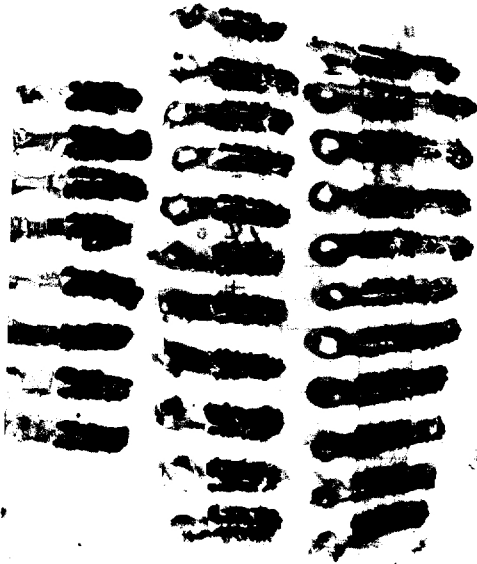
1	Med. ²	3 inoculum ⁴ size:	5	Strain ⁶ :	7	8	9	10
	Penicillin	ca. 10 ⁸ cells/ml.		W6.			(x/100)	
						Conc. of P.I. Soln used:	1 mg/ml.	
	Conc. of P.I.		0x	100x	200x	500x		(5 ml plaq.)
			0 ml	0.05 ml	0.1 ml	0.25 ml		
	Ratio of conusions							
	F ⁻ /total treated		0/30	0/28	0/28	4/29		
	(%)		0	0	0	14.0		
	effect		-	-	-	+		

Purpose: Is ~~the~~ P.I. active for removal of F? (P.I. can remove chloroplast of E. coli.) In former experiment, 10x/ml of P.I. looks active. Increase the concentration of P.I. and see the increase of that activity.

Conclusion: P.I. seems effective at 50x. in penicillin, inoculum size, 10⁸ cells/ml.

cont.

W6 P.I. 08.

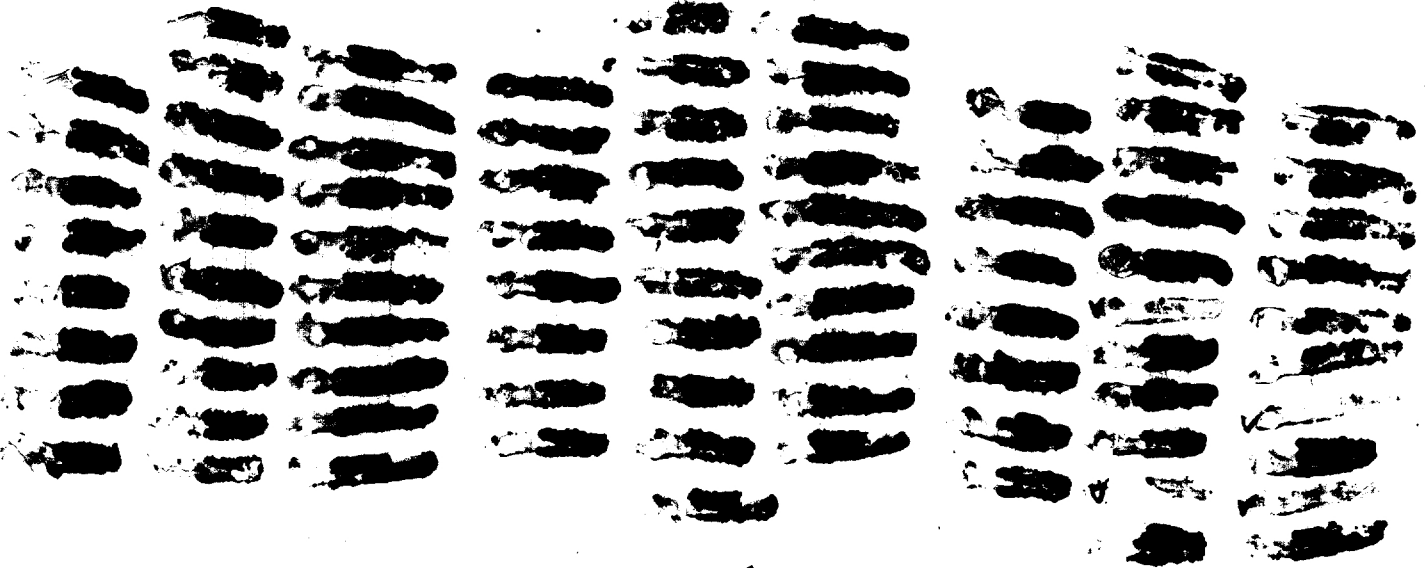


MLac
X 4573

PI 108

P.E. 208

P.I. 508



MLac X 4573