

1/11/55

Irradiation effd  $\bar{c}$  WCG-1-S. Used overnight aerated culture of grow in Ag 1 broth, room temp.

Irradiated  $10^{-4}$  dilution; cfed, washed, resuspended in saline, then diluted.

90 sec irradiation  $\bar{c}$  big lamp.

Diluted further to give  $10^{-7}$  and  $5 \times 10^{-7}$  dilutions of original culture on plates.

Inc.  $25^{\circ}$  incubator

1/13/55

9:00 AM: Irradiated culture plated 1/11: Dilutions used too high:  $5 \times 10^{-7} \Rightarrow$  about 20/plate. Colonies still very small; inc. another day before replicating.

Started culture for second irradiation: 2.1 ml Ag 1 broth culture (not aerated)  $\rightarrow$  Ag 1 broth aerated, room temp.

1/14/55

Replicated 5 plates of  $5 \times 10^{-7}$  dilution to minimal. 6th plate contaminated, incl molds; pick <sup>colonies</sup> spot to complete for later replication. Also pick the few colonies from  $10^{-7}$  dilution to complete.

1/15/55

Exposed plates repl. to min from 1/11 irradiation: 9 possible auxotrophs from 245 colonies. Picked, spotted on complete. Some colonies very rough; use caution on identity. If possible, concentrate on typical colonies.

Second UV run, WCG-1-S:

Culture: aerated at room temp overnight; stored

3 aeration at room temp another 24 hrs.

Cfgd, pellet washed in saline; diluted to 10 ml in saline.  $10^{-4}$  dilution irradiated 90 sec.

Plating: 0.1 ml and 0.2 ml  $5 \times 10^{-6}$  dilution.

1/17/55

Second UV run: Very few colonies visible at 48 hrs!

Continue incubation at  $25^{\circ}$ . By 2:30 pm colonies on  $10^{-4}$  dilution large enough to pick from plates for replating. 2 by 5-12 and 20 plates.

"Autos" from first run: Of the 9 spotted on complete, three are gummy; look like typical Agrobacterium growth. Other six rough & rather leathery. Replicate all to minimal.

Of cols. picked from 1st run plates <sup>(plates which were contaminated or too dilute for replating)</sup> & spotted, then repl to min, 1 of 66 appears to be an auxotroph. Test along with other 9.

Made up 2 minimal tubes. To check medium, aux (drop water suspension from WCG-1-S stab into <sup>1) complete, not aerated</sup> 2) min +  $\text{Na}_2\text{S}$  aerated Used 1 drop 1% sterile  $\text{Na}_2\text{S}$  / tube (10 ml) <sup>3) " " not aerated</sup>

1/18/55

"Autos" from first UV run: 4 of 10 grew well on min. in 24 hrs. These included all the 3 "typical" gummy growers. 2 of the rough type show poor

growth on <sup>replica</sup> ~~replication~~ both to men & complete at 24 hrs.  
 Remaining 2 replicated show good growth on complete,  
 poor growth on men. The 10<sup>th</sup> "copy", brushed  
 on plates to which others were replicated, shows fair  
 growth on compl at 24 hrs, no growth on men.  
 Continue incubation of these plates until early pm.

~~Several~~ Two of the "rough" spots  $\rightarrow$  mixed  
 growth when replicated to men & complete; center  
 of spot rough sparse growth; outer ring of heavier,  
 gummy growth. Especially marked on complete.  
 So it appears these rough colonies may be Agrobacterium.

Colo picked from 2<sup>nd</sup> UV: Overnight growth  
 poor; continue incubation until early pm before  
 replicating to men.

More colonies now visible (3 days) on  $5 \times 10^{-6}$  & on  
 $10^{-7}$  plates from 2<sup>nd</sup> UV run. Pick; spot on  
 complete for repl. to men.

Check on Ag 2 minimal: No growth overnight in  
 men tubes  $\bar{5}$  or  $\bar{5}$  aeration. Un-aerated Ag 1 tube  
 (compl) slightly turbid. Room temp probably quite  
 low during night.

1/19/55

Check of minimal liquid (see 1/17): At 2 days, very good growth of WC8-1-S in min ( $\bar{c}$  Na<sub>2</sub>S) aerated. About the same turbidity in un-aerated mineral as in ~~un-aerated~~ complete.

Why Na<sub>2</sub>S & aeration needed for best growth? Seems contradictory. Effect of aeration may be largely agitation rather than O<sub>2</sub> availability; but at pH slightly below 7 should think sulfide would be rapidly lost.

Set up: min + Na<sub>2</sub>S } aerated  
min  $\bar{c}$  Na<sub>2</sub>S }  
min  $\bar{c}$  Na<sub>2</sub>S } un-aerated  
min  $\bar{c}$  Na<sub>2</sub>S }

1/20/55

Possible culpas from 1st UV run: The rough type grew up eventually on minimal. Check these on a couple of other sugars; could poor growth & lack of green be due to mutation involving utilization of sucrose?

#10 picked from first run appears to be auxotroph. Stab; prepare random tubes. Growth is rather scant even on complete. How be sure of identity ????

Second UV run: Of 53 col. picked, 10 are poorly growing, rough type. If this is a contaminant, it certainly appears

consistently.

One possible escape from 2nd run. Stab,  
streak on complete.

1/21/55

Effect of aeration:  $\bar{c}$  &  $\bar{s}$   $\text{Na}_2\text{S}$ : (48 hrs)

Not aerated: Little difference  $\bar{c}$  &  $\bar{s}$   $\text{Na}_2\text{S}$ . Poor growth.

Aerated, no  $\text{Na}_2\text{S}$ : better growth than  $\bar{s}$  aeration.

Aerated,  $\bar{c}$   $\text{Na}_2\text{S}$ : Very good growth; est.  $10^9/\text{ml}$ .

1/22/55

Inoc Ag 2 run-downs  $\bar{c}$  UV1-10, UV2-1, and WCG15  
from water suspensions from Stals.

UV2-1 grows very poorly when streaked on complete.

"Rough" strains streaked on compl.  $\bar{c}$  glucose & mannitol  
look the same as on sucrose.

1/24/55

Run-downs (2 days)

	<u>WCG15</u>	<u>UV1-10</u>	<u>UV2-1</u>		<u>WCG15</u>	<u>UV1-10</u>	<u>UV2-1</u>
O	++	-	-	HC	++	+	-
A1	++	-	-	YNA	+	±	-
A2	++	-	-	VITS	++	-	-
A3	++	-	-				
A4	++	-	-	compl	++	++	++
A5	++	-	-				

1/25/55

Rundrums (3 days)

	WCG15	1-10	2-1		WCG15	1-10	2-1
0	+++	-	-	AA5	+++	-	-
AA1	+++	-	-	HC	+++	+++	++
2	+++	-	-	YVA	+++	-	+
3	+++	-	-	VITS	+++	-	-
4	+++	-	-	compl	+++	+++	+++

try amino acid single omissions. [The HC used is contaminated, so growth in HC tubes not necessarily due to HC components].

UVrun (#3)

24 hr aerated culture (Ag 1, room temp)

90 sec. Pl. 0.1 & 0.2 ml  $10^{-5}$  dil. (Irradiated  $10^{-4}$ )

1/28/55

UV 3: Replicated to minimal from plates  $\bar{c} > 10$  col. Picked from plates  $\bar{c} < 10$  col & spotted on compl.

One yellow (or creamy) colony. Picked to stab, labelled UV3-1. Colony was same size, shape, height, & texture as others on plate. Check motility; growth  $\bar{c}$  &  $\bar{s}$  sulfide,  $\bar{c}$  &  $\bar{s}$  aeration.

1/28/55

Rundowns, single omissions of AA groups (2 days)

	<u>UV1-10</u>	<u>2-1</u>		<u>1-10</u>	<u>2-1</u>
0	0	0	-5	+	+
-1	+	+	all	+	+
-2	+	+	all + vits	+	+
-3	+	+	HC	-	+
-4	+	+	confl	++	++

Fresh PC used (taken from freezer)

Very similar to behavior of previous mutants. New vit mix not inhibitory. Try various carbon sources & N sources

1/29/55

Rundowns, single omissions, 3 days

	<u>UV1-10</u>	<u>2-1</u>		<u>1-10</u>	<u>2-1</u>
0	0	0	-5	+++	+
-1	++	+	all	+++	+
-2	++	+	all + vits	++++	++
-3	++	+	HC	±	+++
-4	++	+	confl	++++	+++

3rd UV run: replicated to mini. col. picked to complete 1/28.

Direct replerations: 2 possible auxos out of ca 80 col. One is the yellow colony.

1/31/55

UV3: Col. picked, repl. to mini 1/29: 2 possible auxos (both rough, so possibly just poor replerations). { UV3-3 & UV3-4 }

Screen as follows: 1) min. 2) min + HC. 3) min + all AA groups together. 4) Co 3, + VITS. 5) YNA.  
 Also include UV3-1 + 3-2 in tests.

2/2/55

Randoms of UV3 aupos: 2 days

	<u>HC</u>	<u>AA</u>	<u>AAV</u>	<u>YNA</u>	<u>Ag 1</u>	<u>0</u>
3-1	+++	+++	+++	+	++	-
2	+	++	+++	+++	+++	-
3	±	+	-	-	+++	±
4	-	-	-	-	+++	-

2/3/55

UV3 randoms, 3 days.

	<u>0</u>	<u>HC</u>	<u>AA</u>	<u>AAV</u>	<u>YNA</u>	<u>Ag 1</u>
3-1	±	++++	++++	+++	++	++++
2	±	+++	+++	+++	+++	++++
3	±	++	++++	-	-	++++
4	-	-	-	-	-	++++

These were kept for several more days; no further growth in minimal tubes; no change in other tubes.

2/8/55

Inoc. AA randoms of UV3-2 + 3-3. (limpo, single additions)

2/9/55

Set up randoms of UV3-4 as follows: HC, HC+V, HC+YNA, YNA, YNA+V, V, HC+YNA+V, YX. Grew only in YX



2/10/55

Runs, UV 3-2 + 3-3 (2 days)

	<u>3-2</u>	<u>3-3</u>		<u>3-2</u>	<u>3-3</u>
0	-	-	A4	-	-
A1	-	-	A5	-	-
A2	-	-	VTS	+	not included
A3	-	-	compel + vito	not included	++ to check for toxicity
			compel.	++	++

If these results check out after another 24 hrs, try 3-2 = individual vito + 3-3 = AA single omissions

2/12/55

Set up runs = individual vito = UV 3-2. (B-12 may be contaminated).

2/13/55

Run UV 3-2 vit runs.

2/14/55 UV 3-2

Vit runs; At about 28 hrs, fair growth in vito mix, trace of growth in broten.

Would pay to do individual vit = all axes; behavior has been constant = vit requirement, but mix may be toxic to some. Also: 3-3 = ~~AA~~ AA groups.

2/15/55 Vito, UV 3-2; fair growth in broten; other vito 0

UV3-3 Inoc. randoms, AA groups.

2/17/55

UV3-3 no growth in AA groups or in all AA's (2 days)

2/20/55

UV1-10 & 2-1 in vito: no growth in individual vito or mix. (Inoc. 2/18)

2/22/55

UV1-10 No growth in individual vito, but some growth in vit. mix (4 days) May have double requirement, or conc. of requ. vit in individual tube may be too high, since some are higher than in mix. Also may be reversal - see UV2-1

UV2-1 Growth in the following tubes (very slight)

Riboflavin	paba	Reversins?
B-12	vit mix	Dirty tubes?
Pentothenate		

2/23/55

(5 days)

UV1-10 Growth in: Biotin, Dirty tubes choline, nic, paba, vit mix

UV2-1 " " : trace in those tested previously. Very scant even in vit mix.

2/24/55

Set up min, min + glycine (1 mg/10 ml), min + vito, & min, vito, & glycine to see whether glycine will speed growth in vito & itself permitting growth (1-10 & 2-1)

WCG Inc. 2/25

3/1/55

UV 1-10 & 2-1 in glycine, vits:

At 3 days, 1-10 ± in glycine, glycine + vits.

4 days 0 -

glyc ±

vits +

vits+gl ++

2-1 at 4 days: 0 -

glyc +

vits +

vits+glyc - (!?!)

3/4/55

UV 3-3 (2 days) 3 days

0 - - A4 - -

A1 ++ ++ A5 - ++

A2 - - all AA's - ±

A3 - + compl +++ +++

3/6/55

(UV 3-3)

A1 runs down in 3/4. Possibly trace of growth in lysine

3/7/55

A1 runs down - 3 days (at room temp; cold at night)

0 -

meth -

Lys +

A1 -

Arg -

compl ++

Cyst ±

WCG

Try ± & 5 lysine ± aeration. If really Lys - should give more clear-cut result because of more rapid growth.

3/8/55

Start culture of WCG 1<sup>(S)</sup> for another UV exp<sup>(#4)</sup> to try to get a readily identifiable auxotroph.

Ag 1, room temp, aeration

NOV 11 1955  
24 hrs

TRANSFER STABS

3/10/55

Excellent growth of WCG 1-5 for irradiation. Irradiated 90 sec. ( $10^{-4}$  dil)  
Plated 0.2 ml/plate of  $2 \times 10^{-5}$  dilution (Ag 1 medium)

UV 3-3 in min, lys, Ag 1 aerated; In 3 days, faint growth in all tubes.

3/14/55

UV 4 : 6 possible auxotrophs - 1 is ~~the~~ a light yellow similar to UV 3-1.

3/15/55

UV 4-1 (yellow) is an auxotroph; all others grew in min.

3/18/55

UV 5 : 48 (approx) hr culture, Ag 1 broth, aerated. Irradiated  $10^{-4}$  dilution for 90 sec. Plated 0.1 ml/plate of  $10^{-5}$  dilution. Inc.  $25^{\circ}$ .

3/19/55

UV4-1 & UV3-1 (yellow): Struck on complete. 3-1 rough, 4-1 smooth. Appearance  $\cong$  rough & smooth WCG-1 except for color. Smell like Agrobacterium.

Try running down requirements

Spread 4-1 broth (5 days old) on minimal; look for possible "reversions" to prototrophy which should appear as papillae. If any, what color? (I.e. is yellow color result of a metabolic block.)

Also - compare growth in complete broth  $\bar{c}$  &  $\bar{s}$  aeration  $\bar{c}$  WCG-1.

3/21/55

UV 3-1 & 4-1 Both show scanty growth in <sup>complete</sup> broth  $\bar{s}$  aeration, good growth aerated (but not so heavy as WCG-1) 4-1 spread on minimal - No papillae at 2 days.

Reundowns:

UV 3-1 ++ in A4, + in A5,  $\pm$  in o/s } 2 days  
UV 4-1 0 in all reundown tubes.

3/22/55

UV 3-1 & 4-1 reundowns

4-1 no growth except in complete

3-1	A1	0	A5	+++	Imp. 2 distinct
	A2	0	VTS	++	Comp. top of A2
	A3	0	0	0	(4) hist. to no, glud prod. paper
	A4	+++	compl.	+++	5) along, paper, ser, OH prod

3/24/55

UV 5: Of 7 possible cups picked, all show some growth in  
mini at 2 days. Two have only very faint turbidity; keep these,  
check especially for vits. low second check smaller inoculum, no  
growth in 4 days UV 5-1 & 5-2

(3/19)  
UV 4-1 Spread on Ag 2 plate: 1 papilla, which appeared  
in about 3 days. Appears to be slightly buff-colored. Pick, streak  
on complete, check a mini for color & phototrophy.

3/26/55

UV 3-1 re-inoculation - 3 days (run temp - cold at night)

BI	0	pyri	0	A4	++	A5	±
ribo	0	pent	0	hid	0	alon	0
mic	0	<u>chol</u>	++	threo	0	<u>glyc</u>	±
paba	0	vits mix	0	glut	0	ser	0
bio	0			pyr	0	<u>OHP</u>	++
PGA	0			<u>aspart</u>	++		
<small>6-11% for maybe ppt at 24h</small> inos	++					compl	+++
K	0						

Streak out; recheck individual colonies.

3/28/55

Papilla from UV 4-1 - picked from mini, streaked on  
complete - Colonies yellow. Pick several, check for  
anoxytrophly

3/31/55

UV3-1 : Randoms inc. from 2 individual col. (2 days)

	(1)		(2)			(1)		(2) (3 days)	
o	o	o	o	±	A4	o	o	++	+++
nts	o	+	++	+++	hist	o	o	o	+
chol.	o	o	o	+	threo	o	o	o	o
inc	o	o	o	o	gluta.	o	o	o	++
A5	+	+++	o	±	prol.	o	o	+	+++
alan	±	++	o	+	aspart	o	o	++	+++
glyc	o	o	o	+++	campel	+++		+++	
Ser	o	o	o	o					
OHP	o	o	o	+					

Indicates erratic results prob due to reversion?

5-1 & 5-2 randoms: grew in min. However, there are apparently some inhibitions.

	<u>5-1</u>	<u>5-2</u>
o	+++	++
A1	+	±
A2	o	+
A3	±	o
A4	+++	++++
A5	o	o
Vits	+++	++++

4/2/55

Colo. from papilla from UV4-1; autotrophs: However, 1 streak → faint growth after 5 days, c. 1 large colony, many small. On min, large col appears white. Pick, streak on complete.

4/5/55

Final comparison of randoms on two col. from UV 3-1 (inc. 3/29)

	(1)	(2)		(1)	(2)
o	±	+	A4	+	+++
ure	±	+++	hist	++	+
chol	±	++	threo	o	±
inos	±	+++	gluta.	++	++
A5	++++	+++	prol	+++	+++
alan	++++	++++	aspart	++	++
glyc	+++	++++			
ser	o	+++			
OHP	±	+++			

UV4-1 Picked from large col. on mini: Strain mixed, yellow col as before, also a white, rough col. type - latter extremely rough - probably a contaminant.

Might try plating UV4-1 on 3-1 together on WCG 16. Main purpose would be to look for colonies on yellow & white sectors. First check nutrition of WCG 16; also make sure UV4-1 (on 3-1) does not respond to broken.

4/12/55

Spread water suspensions (from slants) of WCG 16 & UV4-1 on plates of mini on 5 broken: Also cross-brushed ea. on SK4.

4/14/55

WCG 16 - <sup>confluent</sup> growth on mini on 5; none on 5 bis.

UV4-1 - no growth on either plate

WCG 16 S<sup>S</sup>

UV4-1 SK

WCG



4/22/55

Results of attempted cross of WCG 16 & UV-4-1 (4 days 25°)

Controls: WCG 16 can grow  $\bar{c}$  SM at 200  $\mu$ g/ml

" " no growth  $\bar{c}$  protein.

UV 4-1 Some background  $\bar{c}$  SM, none  $\bar{c}$  SM,

though culture appeared to be S<sup>R</sup>

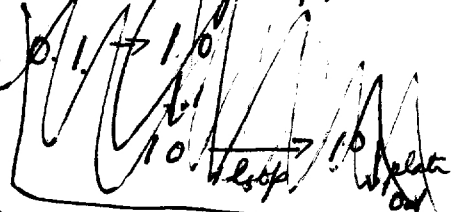
With mixtures, confluent growth on all media. Growth  
is white, i.e. WCG 16

1 min  
min + SM  
min + SM + protein.

# 3-3 AA groups

~~Platt~~  
~~Start~~ crosses of 33-48

In loop dilutions to be plated in  
 ml/cy. 0.1, 1.0, 10.0, 100.0



1-10 HC<sup>-</sup>; in AA single group omissions, grew well in all tubes except mini central. AA<sup>+</sup> sing additions all -; late growth (4 days) in some into

2-1 HC<sup>+</sup>; poor growth in all AA single omission tubes. Late growth (4 days) in some into.

3-1 (yellow) HC<sup>+</sup>; AA's together +; poor in YNA

3-2 HC faint+, AA's together + better in AAV; good YNA  
BIOTIN

3-3 HC +, AA's together strong +, AAV -, YNA -

3-4 Grows only in complete or mini + YX  
 (Have tried HC, HCV, HCYNA, YNA, YNAV, V, HCYNAV)

Possible schemes involving WCG16 + UV4-1  
 See whether both more or less S<sup>S</sup>.

If could obtain:

yellow S<sup>R</sup> x<sup>-</sup> B<sup>+</sup>  
 white S<sup>S</sup> x<sup>+</sup> B<sup>-</sup>

Plate on

- 1) mini
- 2) mini + A
- 3) mini + biotin + L M

What could grow?

- all x<sup>+</sup> B<sup>+</sup>
- only x<sup>+</sup> B<sup>+</sup> S<sup>R</sup>
- all x<sup>+</sup> S<sup>R</sup>

unselected:  
 color, S  
 color  
 color, B

Loop any with Cyst