



19

June 5, 1958

REF: 1420 A

	1	2	3	4	5	6	7	8	9	10
1										
2		Plate		Gal +	Wct +	T ₁				
3		counts								
4										
5	1	310		1/50	5/50	39/50				
6	2	340		3/45	13/45	35/45				
7	3	298		5/50	8/48	30/50				
8	4	380		4/50	17/49	36/50				
9	5	310		4/50	14/48	36/50				
0	6	298		3/50	4/50	35/50				
1	7	320		3/50	17/48	36/48				
2	8	280		2/50	11/45	35/45				
3	9	355		2/50	10/48	31/50				
4	11	474		28/100	45/100	70/100				
5	12	365		14/100	39/100	64/100				
6	13	"								
7	14	127		0/80	3/80	35/80				
8	15	390		5/100	19/100	67/99				
9	16	370		6/50	23/50	35/50				
0	17	400		13/50	23/50	37/50				
1	18	148		0/50	1/50	18/50				← interrupted
2	19	430		0/50	15/48	33/50				
3	20	0								

Conclusions.

pH has practically no effect on transfer; same also on mating unless no recombination takes place in these conditions. Range 4.5-9.

Thioglycolate has practically no effect at 1/50
 Ferriodate M/10000 shows partial interruption

Addition of living F- cells interrupts, that of killed F- cells does not.

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June 1 1958

REF:

Kinetic experiments based on pulsed mating have been unsatisfactory until now on account of an unsatisfactory separation of recruitment and entry phases. In previous control experiments, extent of recruitment varied as approximately the 1/2 power of cell density (instead of expected square), so that subsequent recruitment accounted for 5-25% of total yields. This is quite unsatisfactory.

In this experiment, screen for reagents which may impede mating without preventing further entry.

Following are suggested:

1. pH range in .5 units from 4.5 to 9 Adjust D(0) by HCl or NaOH
2. tris
3. versene
4. thioglycollate
5. ethanolamine
6. dead and live S^S F⁻
7. fluorouracil deoxyriboside ↓ not available today
8. periodate M/100 M/1000 M/10,000
9. D(0) / Mg

Experimental design: Mate ORC W-3060 and W-3064 (washed and resuspended in cold penassay) at sat'd cell density for 5 minutes at 37°. Dilute 1:10 into indicated solutions (made up in D(0) unless indicated, and incubate to 60'. Dilute 1:100 for plating .05 ml on SGalThSm. The pulse matings should gave a baseline count of ~~SRP~~ SRP including some 10% Gal+.

Toxicity will be indicated by a loss of SRP

Inhibition of entry will be indicated by failure of Gal+ among SRP

No inhibition will be indicated by marked rise in SRP

Inhibition of mating / inh. of entry will be indicated by stabilization of SRP with establishment of Gal+.

Controls needed: Plate recombinants (plate mixture after dilution) }
 Early interruption: plate mating at 5'; }
 1:1000 dilution at 5'; plate at 5' 15' and 60' } Dilute in D(0).....
 1:10 dilution: plate at 5' 15' and 60' }

Dilution only 1:10 from penassay!
criticism: may have already been pulsed by high density. Luca's conclusions:
 pH 5 to 8.5 no effect. 4.5 gradual reduction or slight 4 no effect. 2,3 not tried
 D(0)-Mg no effect. periodate suggestive (partial interruption and null-recruitment)
 6A no effect! But 6B



June 4, 1958.

REF:

1420 B

19

1 2 3 4 5 6 7 8 9 10

PULSATION WITH PERIODATE

Parents: W3060, W3064

ORC cultures, spun, susp in chilled water 20x until used

then diluted in: Penafay, 1/20,

DM (D(M) + a parafine 1%)

DO (D(O) + " + glucose .5%)

α1 Pulse 2' in Penafay, mixing equal amounts of parents: 1ml each in a flask, starting from chilled susp mixed while chilled. Dilute after 2' in DM periodate M/20000, 1/100, incubate and sample every 5' until 40'; dilute sample of 0.2ml in 0.8ml, blend and plate on D(Sm B₁) 0.05 ml.; for 20' onwards also further 1/5 dil. ^{plated}

α10 From pulse dilute in D(M) 1/100, without periodate; sample only after 40' and plate 1/5 and 1/25 .05 → D(Sm B₁) -

There were no colonies except in α10 (when there were 4 colonies).

F+ reversion of W3060?

Moreover while this part of the experiment showed contamination with staph, the second part of this same exp., which used the same cultures, showed no or less contamination, but was also staph.

June 4 1958

REF:

Matings. Premix chilled matings (ORC W3060, W3064 in chilled penassay, resuspended 1x. Pulse = time
 1 a Pulse 2 min in penassay 1 ml each in 10 ml tubes held at 37°.
 2 B Pulse 0 " " " Dilute 1:200 each parent (not ml ~~thru~~ in 10 ml tubes.
 3 Y " 2 " " D(o) 5 ml in tube
 4 Δ " 2 " " D(m)
 5 E " (a) 0 kept chilled for plate recombinants
 6
 7
 8 add periodate to chilled D(o, m) tubes.
 9
 10

sample 5 ml		Dilute in	# Periodate	
	α	D(m)	2000	→ time series.
all others	β	"	0	
are plated	β	"	500	
at 40' (600)	β	"	1000	
	β	"	2000	
Dilute 1:10	β	D(o)	0	
in chilled	β	"	500	
water.	β	"	1000	
	β	"	2000	
	α	D(m)	0	
	β	add penassay just	D(m) + P ₁₀	0
	β	prior to cells.		500
	β			1000
	β			2000
5 minutes	β	thioglycollate 0.1%		500
	Y 7 pulse	D(o)		0
	Δ 5 pulse	D(m)		0
	E Mixed at 0' up just to 40' plated 1:1000			
	E - mix post dilutions	1:100 plated		
	"	1:1000 plated.		

Does dilution into periodate give a pulse? Possibility that 1/2000 periodate is toxic if not neutralized? Should try adding thioglycollate at various times, providing thioglycollate does neutralize. 1/2000 periodate was not toxic in 15 minutes with a higher cell suspension (14/12). Agar will neutralize per periodate.

No colonies! Presumably reversed then!

June 9, 1958.

1420 C

	1	2	3	4	5	6	7	8	9	10
1										
2			Pulsation with periodate.							
3										
4										
5	#	3060	(?)	3064.	Sheet giving details of cultures missing.					
6										
7										
8										
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Pulsation with periodate.

3060 (?) ; 3064. Sheet giving details of cultures missing.

Pulsed 2', diluted 1/100 in

P. DM + asparagin 1/1000 + yeast 1/1000 + periodate 1/2500

C. " " " " —

B. Penallay broth

Samples taken at 20', 30', 40', 50', 60' (or parts), diluted 1/5, and blended and plated on D Sm B₁.

Plate counts. (log colonies; also some minute colonies).

	P	C	B	
20'	3, 9	7, 5	12, 8	All streaked on B kc 6/11
30	1, 4	10, 3	8, 14	
40	6, 4	2, 2		
50	8, 5		6, 9	
60	7, 10	8, 12	28, 24	

1420 C - Lac strokes

P 20 4+ , 9-
P 30 1+ , 4-
P 40 1+ , 9-
P 50 3+ , 9- , 2 w
P 60 6+ , 10- , 1 w

C 20 7+ , 6-
C 30 6+ , 7-
C 40 4-
C 60 9+ , 11- (some w?)

B 20 6+ , 14- , 1 w
B 30 10+ , 15- , 1 w
B 50 7+ , 9-
B 60 21+ , 34-



19

June 11, 1958.

REF: 1420 D

	1	2	3	4	5	6	7	8	9	10
1										
2										
3										
4										
5	AC.	W 3870	0.1 ml	} 10' incub → 0.1 + 0.5 ml W945 + 0.9 broth (A)						
6		W 3064	0.1 ml							
7		Broth	1.0 ml							
8										
9										
0										
1										
2	B.	W 3870	0.1 ml	} 10' incub → 0.1 + 0.1 W3064 + 0.9 broth (B)						
3		W 945	0.1 ml							
4		Broth	1.0 ml							
5										
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INHIBITION BY LIVING CELLS. INTERRUPTION

Cell suspensions 10x (same as in exp. 1422)

AC. W 3870 0.1 ml
 W 3064 0.1 ml } 10' incub → 0.1 + 0.5 ml W945 + 0.9 broth (A)
 Broth 1.0 ml } → 0.1 + 1.4 broth (C)

B. W 3870 0.1 ml
 W 945 0.1 ml } 10' incub → 0.1 + 0.1 W3064 + 0.9 broth (B)
 Broth 1.0 ml }

A, B, C : at 5', 15', 30' 0.2 ml + 1.8 chilled water;
 0.05 D (Sun B)

Plate counts.

	A	B	C
5'	91,72	-	164,128
15'	162,182	77,85	456,410
30'	160,155	343,510	460,486



	1	2	3	4	5	6	7	8	9	10
1		Gal'/hr	Lac'/hr	T ₁ '/tot						
2										
3										
4										
5	A 5	0/50	1/50							
6										
7	A 15	0/50	1/50							
8										
9	A 30	0/50	2/50							
0										
1										
2	B 15	0/50	0/50							
3										
4	B 30	0/50	5/50							
5										
6										
7										
8	C 5	0/50	0/50							
9										
0	C 15	0/50	12/50							
1										
2	C 30	0/50	12/50							
3										
4										
5										
6										
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7										
8										
9										
0										

Addition of W945 cells to the cross inhibits further mating, interrupts. Compare with control C.

No accelerated Gal detected in the second mating.

Note. W945 is added in some excess over physiological conc. of cells. Interruption may be due to transfer to over-saturated flagell. cell concentration. Retest with normal cell conc.



19

June 25, 1958

REF: 1420 E

1	2	3	4	5	6	7	8	9	10
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♂ INTERRUPTION BY LIVING FEMALES.

3^{hrsp} R.C. W 3064, W 3870, W 945.

Spun, resusp. in fresh broth.

A ♂ : ♀ = 10 : 1 ratio.

2 ml 3870 + 0.2 ml 3064, incubated 20'
then:

AC: 0.9 ml transferred with 1 ml ^{warmed} pipette to empty, warm tube, incubated 20', diluted 1/20 & 1/100 (chilled) plated on 1/2 gal sm B₁ (.05).

AD: 0.1 ml transferred to 0.9 ml W 945 ^{warm} suspension and incubated 20', diluted 1/2 & 1/10 chilled. plated as above

B ♂ : ♀ = 1 : 1 ratio

1 ml 3870 + 1 ml 3064, incub 20'
then:

BC: 1 ml. transferred with 1 ml. warmed pipette to warm tube, incubated 20', diluted 1/100 & 1/500 chilled, plated on 1/2 gal sm B₁ (.05).

BD: 0.1 ml transfers to 0.9 ml warmed W 945 susp. incubated 20', diluted 1/10 & 1/50 chilled, plated on 1/2 gal sm B₁.

Purpose: to test if addition of living ♀♀ interrupts even when the whole experiment is kept at the same total conc. of cells throughout, and this concentration is within physiological limits.

Periodate

1421A

Jan 1 1958

REF:

u3060 + u3064
Same cells as 1420. Harvest 5ml into .5ml water.

1ml + .4ml periodate : 1/200; 1/2000; 1/20,000 and H₂O.

after 10 minutes treatment at 37° add 4.5ml peptone + 1% glycerol.

Mix equal volumes 15' at 37°, dilute 1:100 and plate on D-13; 522.

Concn	♂ treatment	♀	8P2. Pulm.	A3 Count
0	0	0	+	149
2000	2000	2000	-	2
200	200	200	-	0
20	20	20	-	0
2000	2000	0	-	2
200	200	0	-	0
20	20	0	-	0
0	0	2000	+	45
0	0	200	+	1
0	0	20	-	0

Toss by chilling

1-6 diluted, in 0.1 ml rather than 0.1 ml.

at 1/2000, interaction maybe differential.

Preliminary results are very promising (for periodate as a de-sensitizing agent). Could the aerobiosis phenomenon have been oxidation of surface receptors? If so should see a + phase.

See 1420 for origin of this experiment. Periodate is of course known to affect the receptors of animal cells for influenza virus. (Will it agglutinate any bacteria?)

Try other oxidants? Ferricyanide?

Encubation (within limit of test): sexual competence is destroyed by periodate, ♂♂ more sensitive than ♀♀. [Used kinetic and viability controls.]

1421B shows 1/200 periodate is toxic; 1/2000 is not.

If they're inactive, will periodated bacteria deactivate ♂♂?

June 3 19 58

REF: 1420;1421

1 Harvest OMC W-3060, W-3064, 3x7 ml each into 1 ml water. (Keep chilled during day
2 before beginning experiment about 7PM. 1421 suggested differential effect of periodate.
3 This is essentially a repetition with some added controls for viability and reproduc-
4 ibility.

5 In each case, add 0.1 ml cell suspension to 0.5 ml treatment solution (in D(m)). Treat
6 10 mins. at 37; neutralize 5 mins. by addition of 4.5 ml penassay-15 glycerol. Mate
7 for 15' by mixing .5 ml each suspension. Then chill, dilute 1/100 and plate 0.1 ml
8 on D-B; on agar: duplicate plates. Also dilute 10^{-6} and plate ml 0.1 ml on EMB Lac
9 for viability.

Treatment: (periodate conc.)

1	Viability		SRP counts	
	♂	♀	Lac+	-
2	-	-	28,29	60,38
3	M/10000	-	49	78
4	5000	-	32	85
5	2000	-	30	71
6	200	-	2	104
7	-	10000	107	84
8	-	5000	30	62
9	-	2000	36	52
10	-	200	53	1
11	2000P	-	43	68
12	-	2000P	22	71

(A few SRP counts were slightly distorted by smearing)

These counts reflect a substantial pipetting error, but not enough to weaken the experiment.

12 Tubes 10-11 (P) had periodate neutralized by penassay-glycerol/prior to addition of the cells for 5'

CONCLUSIONS: M/200 periodate is toxic; M/2000 and less is not.

M/2000 periodate differentially inactivates fertility of ♂♂, with no perceptible effect on fertility of ♀♀ or viability. M/5000 has a partial inhibitory effect; M/10,000 is doubtful. Must explore possibility that mild treatment of ♀♀ enhances fertility; try especially neutralized M/2000 or M/10000. This might be explained if there is a phase of periodate treatment with slows the separation of the pairs.

Also need a more careful study now of effects on interruption and on speed of effects.

June 3 19 58

REF: 1420;1421

1 Harvest ORC W-3060, W-3064 3x7 ml each into 1 ml water. (Keep chilled during day
2 before beginning experiment about 7PM. 1421 suggested differential effect of periodate.
3 This is essentially a repetition with some added controls for viability and reproduci-
4 bility.
5 In each case, add 0.1 ml cell suspension to 0.5 ml treatment solution (in D(m)). Treat
6 10 mins. at 37; neutralize 5 mins. by addition of 4.5 ml penassay+1% glycerol. Mate
7 for 15' by mixing .5 ml each suspension. Then chill, dilute 1/100 and plate 0.1 ml
8 on D-Bism agar: duplicate plates. Also dilute 10^{-6} and plate ~~at~~ 0.1 ml on EMB Lac
9 for viability.

Treatment: (periodate conc.)		Viability		SRP counts	
♂	♀	Lac+	-		
-	-	28,29	60,38	27	27
M/10000	-	49	78	20	15
5000	-	32	85	7	8
2000	-	30	71	0	0
200	-	2	104	0	0
-	10000	107	84	25	60sm.
-	5000	30	62	47	23
-	2000	36	52	38	37
-	200	53	1	0	0
2000P	-	43	68	102	55
-	2000P	22	71	50 _{sm}	23

Hfr keto on lac+
0/10 from 1
0/2 from 2.
Valid keto? or
cultures attenuated of
Hfr.

(A few SRP counts were slightly distorted by smearing)

These counts reflect a substantial pipetting error, but not enough to weaken the experiment.

Tubes 10-11 (P) had periodate neutralized by penassay-glycerol/for 5' prior to addition of the cells

CONCLUSIONS: M/200 periodate is toxic; M/2000 and less is not.

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Also need a more careful study now of effects on interruption and on speed of effects.

Periodate
Effects on F⁺

1421C

5 June 1958.

REF: 1421

1 ORC W6, W3064 necessary \rightarrow water. Chill 10 min.
 2 α : .2 ml + .8 ml D(m) + asparagine + 1/2000 periodate } W6
 3 β : .2 " " D(m) + " }
 4
 5
 6 Incubate 10 minutes. Add .2 ml W3064 and
 7 Plate .05 ml samples on DB, sm.

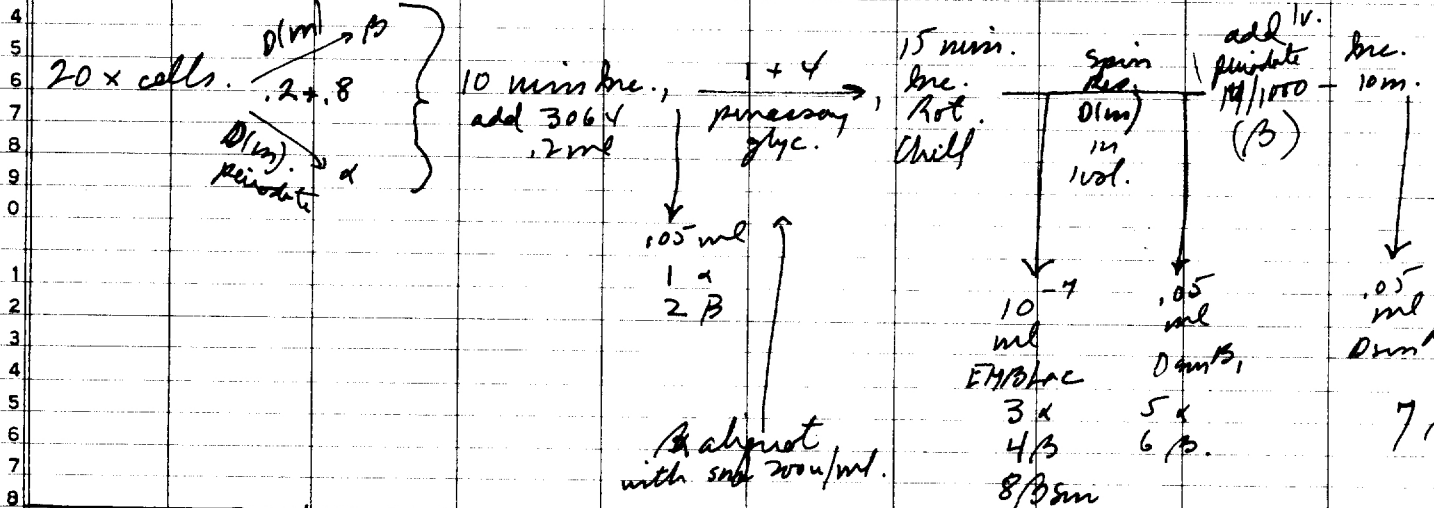
1 α Crowded
 2 β Crowded

3 dilute 1:4 in necessary glycol, rotate 15 minutes, chill:
 4 dilute and plate for F transfer. Add sm 200u to aliquot of 4 \rightarrow 8.

5 α uncountable
 6 β 200-, 130+
 7 13 sm. 290-, 2+ 120-, 0+

8 Spin and resuspend in 1v. Plate .05 ml samples on DB, sm
 9 α 18, 23 days
 10 β 107 days, ✓
 11 13 sm

12 (Donby) add periodate to 1/2000, incubate 10 minutes and
 13 β plate .05 ml 26, 30 days,



14 Pretreat F⁺ with various concentrations of periodate.
 15 incubate 20x W6 1:1000 in D(m), 1:100 in treatment tubes (10 min.)

16 plate .05 ml, directly and 1:100 (duplicate).
 17 Viability

Conclusions: 1 & 2 shows that periodated F⁺ still gives overblowing yield of plate recombinants (key requirement of receptors?)

- 12 1/2000
- 13 1/1000
- 14 1/500
- 15 1/200
- 16 1/100
- 17 into plates prepared with .05 ml 1/200

Periodate and other oxidants
effect on Hfr.
Regeneration of receptors

1421 D

5 June 1958

RE 1421

As 1421A. - Pre-treatments of W 3060.

1 incubated at 37°C unless otherwise stated
2
3 20x cells diluted .1 + .4 ml into treatments. Neutralized by penicillin-glyc
4 10 m. 5 m.

5 Mate + equal volume W 3064 15 min., dilute 1:200 and plate .05 ml.
6 (30S 1:1000)

Counts

1	Periodate 4/2000	37	10 m.	7/15	0
2	" "	at 0°C.	30 m.	4/3	4
3	" 30(m)	37°	10 m.	4/3	43
4	" 4/2000	37°	5 m.	2/3	2
5	Ferricyanide 1/10000	37	5 m.		36
6	" 1/100		10		24
7	" 1/10000	"	"		22
8	Dichromate 4/10000		"		15
9	" 1000		"	20	2
10	" 10000		"		2

1	Bubble oxygen	37	10 m.	? Any survival? (some) not possible.
2	"	37	30 m.	
3	"	0°	30 m.	
4	"	0°	30 m.	

Regeneration: Use samples of ① after neutralization.

This is now at $1/5 = \frac{1}{50} \times 20\% = .4\%$ (i.e. 4% saturating concentration).

dilute further 1:10 in penicillin. F(E) take 0.2 ml + .1 ml of 1x 99, incubate 12 m.
(test receptors) dilute 1:10 + plate

0 minutes postpone.

Control counts not very satisfactory and no control on plate recombination.
Conclude: periodate effect is byre; incomplete at 0° and < 10 minutes
Ferricyanide has no effect on fertility at 1/100. (look for viability!) (Regenerate a useful indicator). Dichromate effective only at 1/10000! (presumably viability).
Bubble oxygen for 10 minutes induces F-phenotype! Should also be reported.

1 ml
20x cells
2 ml (10m)
These
2.5 ml
penicillin
glycyl
compare with
others.



19

June 13 1958

REF: 1421 E

	1	2	3	4	5	6	7	8	9	10
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DIFFERENTIAL EFFECT OF
VARIOUS AGENTS ON MATING
ABILITIES.

ORC W 3870, W 3064, spmy conc. 20x in chilled water.

	A	B
1. DM	1 3	1
2. Nutrient Broth + $CaCl_2$ 1%	4 1	0 3
3. RDE + $CaCl_2$ 1%	3 1	4 0
4. Lysozyme .5%	2 0	2 0
5. Dichromate M/1000	2 2	0 3
6. Ferricyanide M/100.	1 0	2 4

M: 0 0

0.4 ml of each agent added to

A. : .1 ml of ♂ susp. 20x conc.

B. : .1 ml of ♀ " "

Treat for 10' at 37°. Chill, add 4.5 chilled broth to every tube.

From tube A₁ (control ♂), .5 in ^{new} tubes A₁, B₁-B₆; from tube B₁ (control ♀) .5 ml in A₁-A₆, B₁.

20' incubation then dilution 1/100, mating .05 Dfm B₁.

7. Plate recombination control. Parental susp. diluted 1/100 → 1/50
dilutions mixed, .05 Dfm B₁-

Control not enough recombinants! Culture 11.5.?



June 11, 1958

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- RDE - inhib. of ♂♂ & mating.
- ORC, spun, resusp in 1ml 10x, chilled. ♂ = 3870, ♀ = W3064.
1. RDE 1 ml + 0.1 ml ♂, 10' incub. → 0.1 ml + 1 ml broth + 0.1 ♀: 20' incub. → 1/100 dilution → 0.05 ml D(Sm B₁).
 2. RDE 1 ml + 0.1 ml ♀, 10' incub → 0.1 ml + 1 ml broth + 0.1 ♂, 20' incub. → 1/100 dil. → .05 ml D(Sm B₁)
 3. Same as 1. but broth used instead of RDE
 4. Same as 2. " " " " " "

Plate counts:

1.	201, 181	Sum	382
2.	185, 164		349
3.	277, 215		492
4.	171, 216		387

5 : 2 ml broth + 0.1 ♂ + 0.1 ♀, 25' incub → $\begin{matrix} 5A \\ 5B \end{matrix}$

$\begin{matrix} 5A \\ \swarrow \\ 5B \end{matrix}$ → 0.1 in 1 ml RDE for 15' → 1/10 → .05 D(Sm B₁)

5B → 0.1 in 1 ml broth " " " "

About 400 colonies in every plate of the 5A, 5B series.

Conclusions:

No effect of RDE detected on mating ability of cells.

Note: no Ca⁺⁺ added.

June 12, 1958

REF: 1423A

³ DELAY⁴ EN⁵ ENTRY -
Chloramphenicol.

Testing possible delay of spontaneous interruption.

ORC W 3870, W 3064, spun and resusp. in water 20x concentrated. Chloramphenicol solution 1%.

- A. Penallay 1ml + ♂ 0.1 ml + ♀ 0.1 ml + 0.05 Chloramph (= 40 g/ml)
- B. " " " " + — (control)
- C. " " ♂ 2.5% 0.1 ml + ♀ 2.5% 0.1 ml + 0.05 Chloramph.
- D. " " " " + — (control)

A, B: at 30', 60', 90' and 135' dilution 1/500 (0.02 in 10 ml), plating of .05 on DSmB₁.

C, D: at same times dilution 1/10 (1 in .9 water) and plating.

E. Control of plate recombination at various concs. of cells.

E. From same mother suspensions, .025 ♂ + .025 ♀ + 10 ml chilled water; → .05 in DSmB₁ = E₁
 ↓
 → 1 ml diluted serially + 2.16 chilled water = E₂-E₇

Plate counts:

E 1	2, 9
E 2	1, 0
E 3	1, 0
E 4	0, 0
E 5	0, 0
E 6	0, 0
E 7	0, 0



	1	2	3	4	5	6	7	8	9	10
1		Gal	Lac							
2										
3										
4	A 30	0/7	2/7							
5										
6	A 60	0/5	1/5							
7										
8	A 90	0/9	0/9							
9										
0	A 135	0/6	0/6							
1										
2										
3	B 30	0/13	0/13							
4										
5	B 60	0/10	1/10							
6										
7	B 90	1/10	2/10							
8										
9	B 135	1/25	2/25							
0										
1	C 30	0/23	7/23							
2										
3	C 60	1/20	11/20							
4										
5	C 90	1/10	5/10							
6										
7	C 135	0/7	4/7							
8										
9										
0	D 30	2/50	16/51							
1										
2	D 60	4/50	19/50							
3										
4	D 90	2/50	24/50							
5										
6	D 135	9/50	22/50							
7										
8										
9										
0										

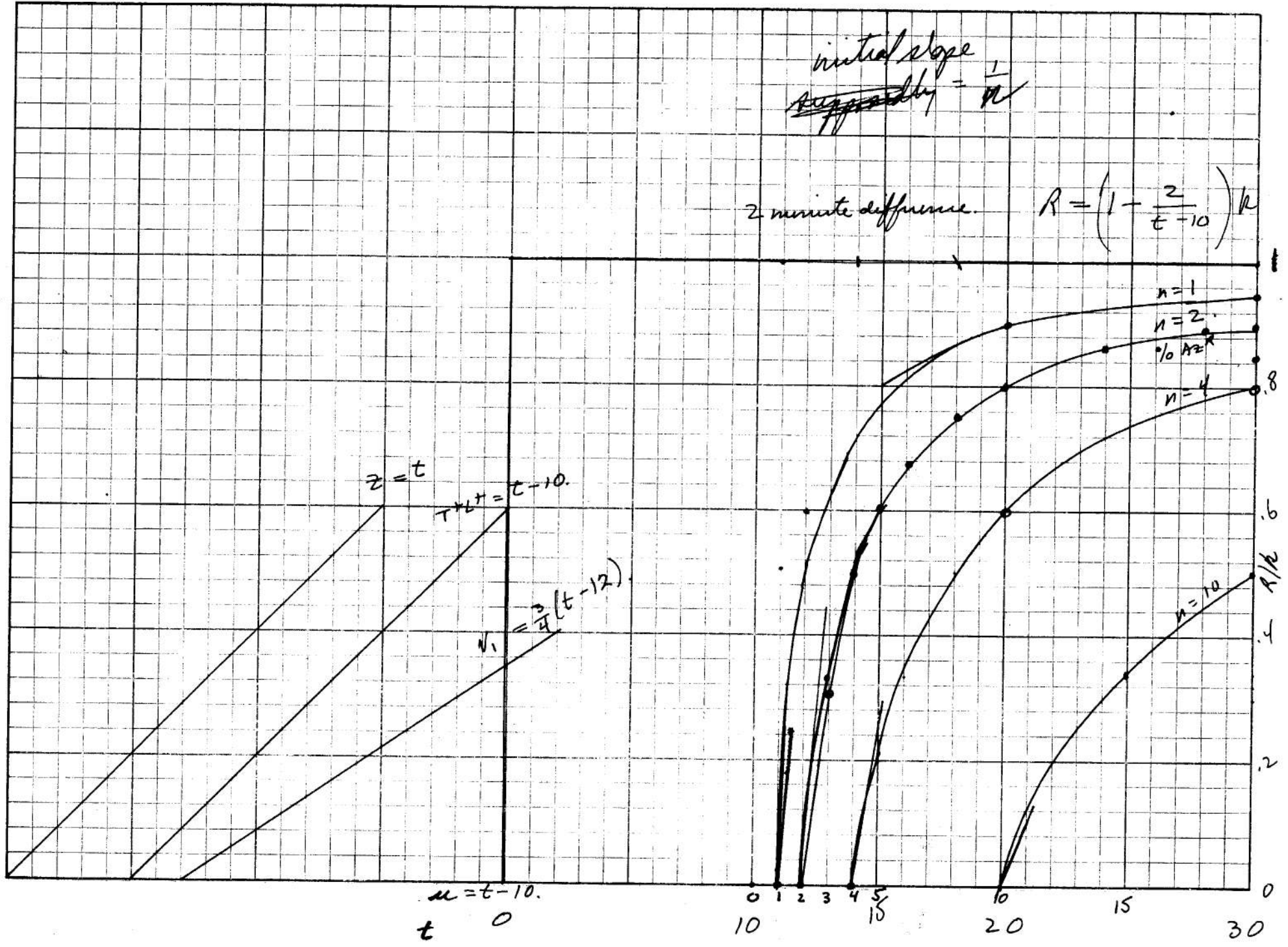
Plate counts.

	30	60	90	135
A	7	5	9	6
B	13	10	10	25
C	23	20	10	7
D	65	218	320	520.

Conclusions. : A, B in unphysiological conditions, low motility (excess of cells or poisoned culture?). In C, chloramphenicol may have slight bactericidal effect; at any rate does not permit multiplication, as judged from prototroph count. Chromosome entry possibly undisturbed.

initial slope
~~approximately~~ $= \frac{1}{n}$

2 minute difference. $R = \left(1 - \frac{z}{t-10}\right)^n$



$$Z = t \cdot n_p \cdot n_q$$

$$TL^+ = \frac{t-10}{t}$$

$$\frac{\frac{d}{dt} TL^+}{TL^+} = k \frac{t-10-n}{t-10} = \frac{u-n}{u}$$

Note: slopes have to be multiplied by k .

Note here: curves are independent of rate of recruitment! (in these assumptions).

