

September 9, ff., 1949.

2 l. ~~activated~~ K12/Y2 Lac washed and concentrated to 30 ml.
 Aliquots of 15 ml ea. mixed \bar{c} A) 15 ml H₂O ; B) 15 ml NaP 7/5.
 and incubated 1 hour at 30°. After removal of 1 ml, 29 ml
 samples were dried^{over: over 20s}, and subsequently found to yield 1.642 and .560 g.
 respectively after washing, or 22.1 and 19.3 mg/ml respectively.

Assays of A and B before and after drying were (u./mg.)

	wet	dried
A	5.1	104
B	44.5	146

After ~~the~~ benzene treatment, an activity of 157 u./mg was recovered.

2. Can dried cells be further activated? Relate these activities to V_{max} .
 pH characteristics of activated cells. Rb responses.

September 9, 1949

Assay aliquots of A and B. $\frac{1}{10}$; $\frac{1}{10} = .01 \text{ ml}$

	Di	20m.	R.A.	^{20m.}
A	089	193		113
B	080	329 ^{6min.}		$257 \times \frac{10}{3} = 858$

1 ml A = $\frac{1642 \text{ mg}}{29 \text{ ml}} = 22.1 \text{ mg}$, assuming complete recovery.

4:30 -
5:10
m A cells.

	Di	3 min		
Benzene	067	530 $\times \frac{20}{3}$	3860	3500 = 3500 μ /ml A
Toluene	048	430 $\times \frac{20}{3}$	2500	$\div 157 \mu$ /mg.

\therefore autolysis strongly activates galactosidase.

.01 ml samples of A, B suspensions have activities of 113; 860 μ respectively, $\div (113, 860) \mu$ /ml. Total samples should be 29 x ... or

	Total.	grams dry. wt.	μ /mg.
A	3280	560	1642 g.
B	24800	aqueous	560 g.

Use .02 ml samples of 1% suspensions of dried cells for comparison.

	Di	De	T.	R.A.	μ /ml	μ /mg.	μ /mg prod.
A	.02 ml	040	560	5min. 2080	560 1040	1040	5.1
B	.01 ml	014	380	5min. 1464	380	146	44.5

Benzene: 157

This drying has resulted in optimal activation of E. coli lactase.

Lactase activation

September 9, 1949.

Harvest and water wash K-12 from 2 l. aerated 37° Y2 Lac 1 1/2 %.

Suspended in 35 ml. Remove (5 ml), and separate 15 ml portions of remainder: A) + 15 ml H₂O B) + 15 ml NaP M/5 pH 7.5. Incubate in stoppered flask at 30° 1³⁰ to 2³⁰, for subsequent dry cell preparation. At 2³⁰ Remove 1 ml aliquots, and sediment + dry remainder

[Dilute 1/100; 5/10 = 1/2000 for assays.]

A) assay in dil (M/50) and conc. (M/10) buff. Do latter in colorimeter.

Use cells & ONPG as blanks.

ONPG O: 034.
Cells D: 200

M/5 buffer NaP. 8.5 ml
cells (add at T₀) .5 ml
onpg 1 ml

Time.		D.
215	20 s	036
	60 s	034
	1 30	035
220	2 40	039
	5 M	040
	6 M	
	7 M	
	8	
	9	
	10	
233	18	052
239	24	087
242	27	100
250		
245	30	112
253	38	146
305	50	191

605

Kinetics of activation
NaP buffer .17M 30+°.

Activation ratio: $\frac{82}{18} = 4.5$

200

100

75

50

25

0

5

10

15

20

25

30

35

40

45

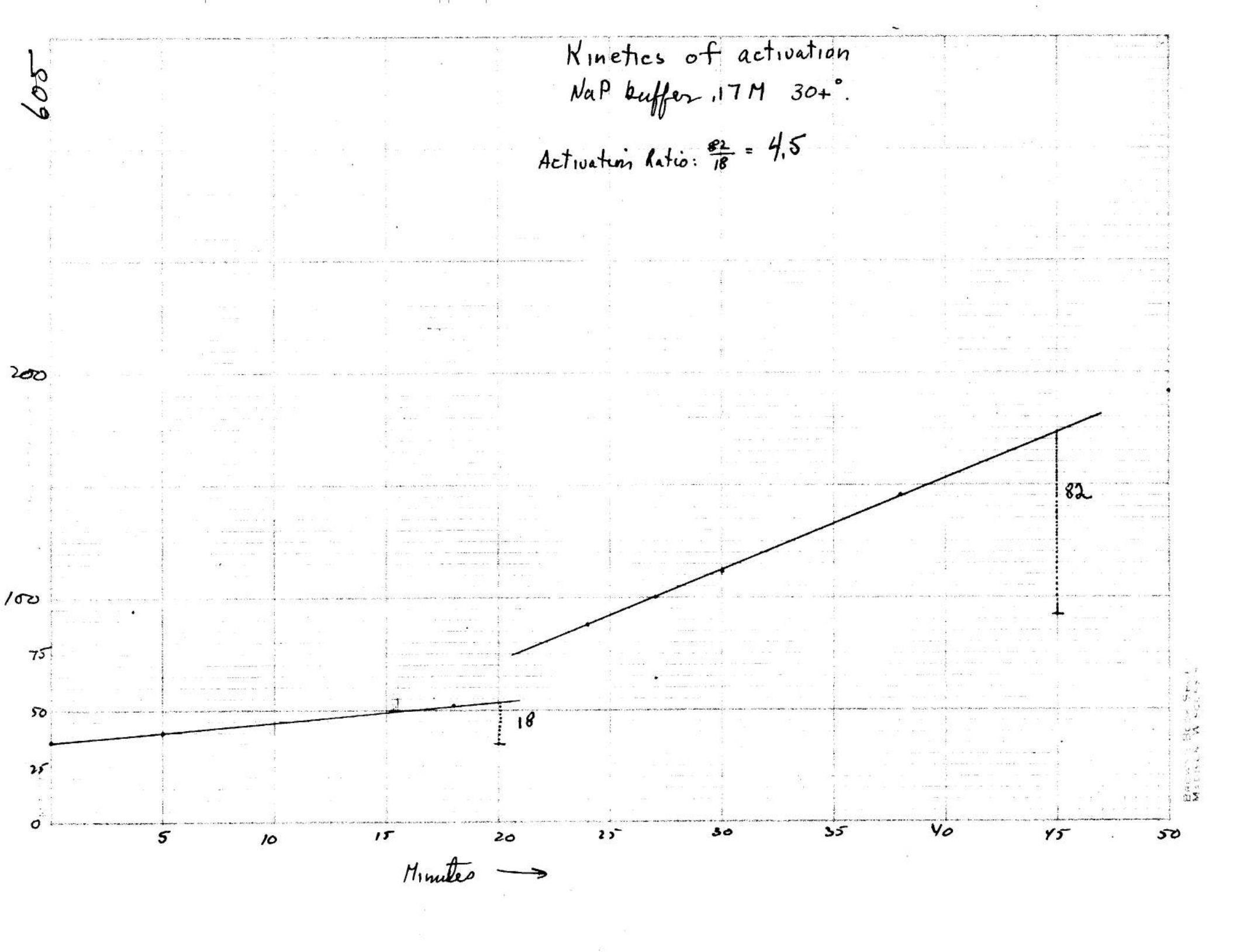
50

Minutes →

82

18

BRUNNEN
MILWAUKEE, WISCONSIN

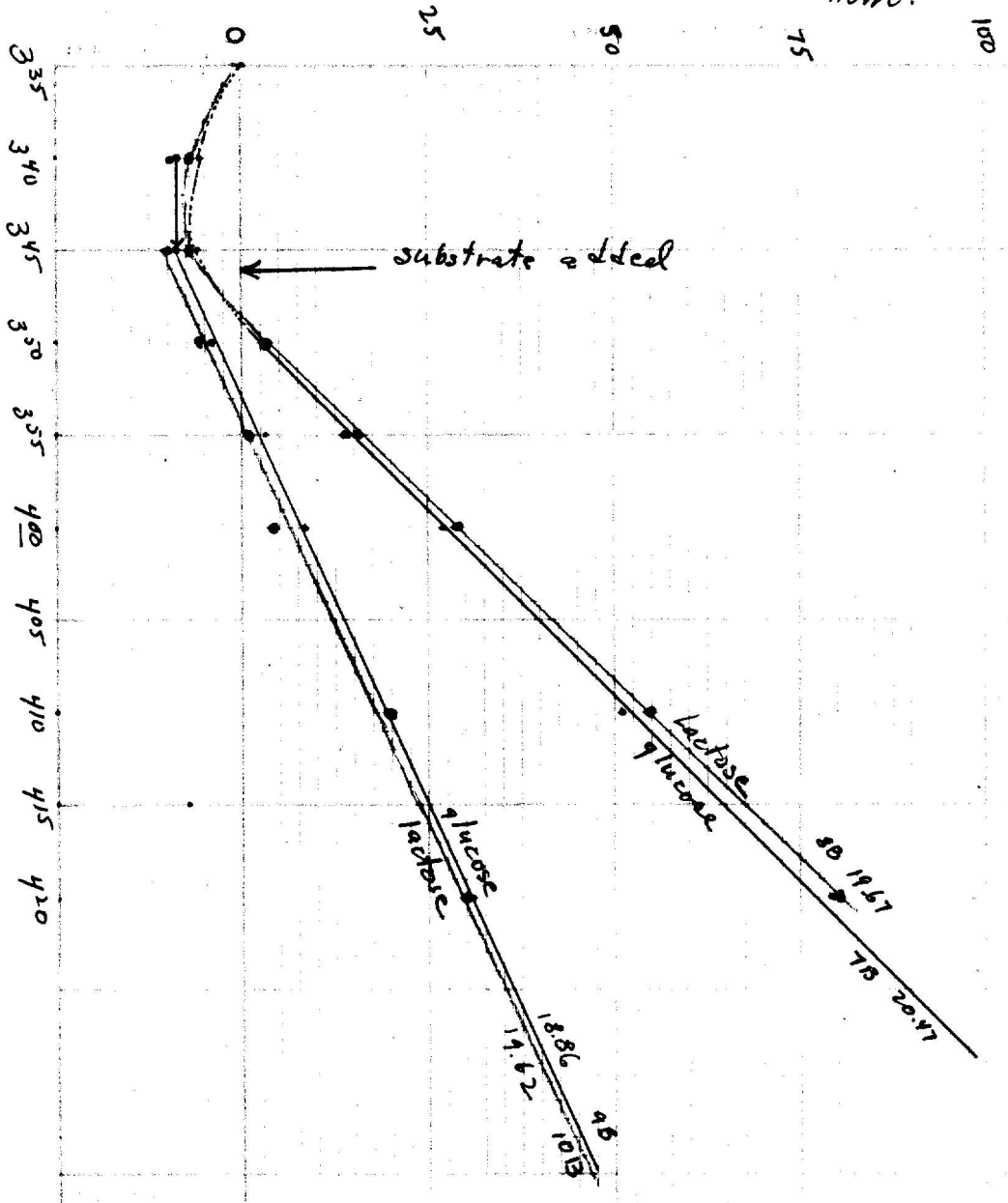


	Thermoc	7-7B	6-8B	9-9B	10-10B
main vessel.	.1cc	← 2cc $M_{10} NaHCO_3$ - $M_{10} Na_2PO_4$ →			
		← Untreated coli →		← treated c. $M_{10} Na_2PO_4$ buffer! →	
side cup.	.1cc 10% glucose	lactose	glucose	lactose	
3:35	161	104	68	89	23
3:40	148 +3	95 -6	58 -7	78 -8	61 -9
3:45	147 +4	93 -7	57 -1	77 -8	59 -12
3:50	150 +6	106 +10	70 +10	84 +4	66 +4
3:55	149 +2	119 +4	84 +15	90 +7	72 +7
4:00	147 +4	127 +10	93 +11	93 +5	73 +3
4:10	149 +2	153 +24	121 +26	107 +12	91 +15
4:20	153 +2	185 +28	157 +26	122 +11	105 +10

Nanometric tests on "activated" cells.

ca. 50% inactivation of buffer treated cells.

mm.



Unconverted.
Km = 0 given.

Utilization of Isomaltose

September 8, 1949.

	0/g	2/m	4/g	4/m	4/m	4/-			
	9A	2A	4A	8A	10A	7-10A	T		
150	0	14 0	07 0	12 0	15 0	04 0	450	152	0
210	5	18 4	07 0	05-7	10-5	01 -3	450	152	0
217	10	14-3	15	13-2	16-2	06 -1	491	155	-3
225	(35)	11-6	12	09 0	14-4	03 -4	47-1	155	-3
3mm →									
230	(40)	13-4	09-1	19 4	18 0	03 -4	42-6	155	-3
	5	18 0	10 -1	36 20	28 9	03 -5	42-7	156	-4
	10	25 6	12 0	58 41	43 23	08 -1	47-3	157	-5
	15	33 14	11 -1	72 55	49 29	04 -5	42-8	157	-5
	20	38 19	08 -1	92 77	62 44	09 +2	48 0	155	-3
308	25	51 29	14 -1	111 91	71 48	08 -4	46-7	160	-8
	30	56 34	12 -3	126 106	77 54	04 -8	42-11	160	-8
338	60	101	14	218	133	06 -9	43-13	163	-11
	X		X	X					
503		09				03 -13	42-15	164	-12
		X			X	X			

K12 *Celle grossis in glucose or maltose (D, M)*

2ml cells, 1ml substrate 10% = 10mg. g. ml.

NaHCO₃ M/20

CO₂

NaP M/1000

9A D, g

2A S, m

4A M, g

8A M, m

10A M, isomaltase

7A M -

Isomaltose not utilized by maltose-adapted K-12!

606

100

80

60

40

20

00

-20

Utilization of isomaltose

150
10

20
2¹⁰

30
2²⁰

2³⁵

2³⁰

240

250

3⁰⁰

4⁰⁰

5⁰⁰

Substrate

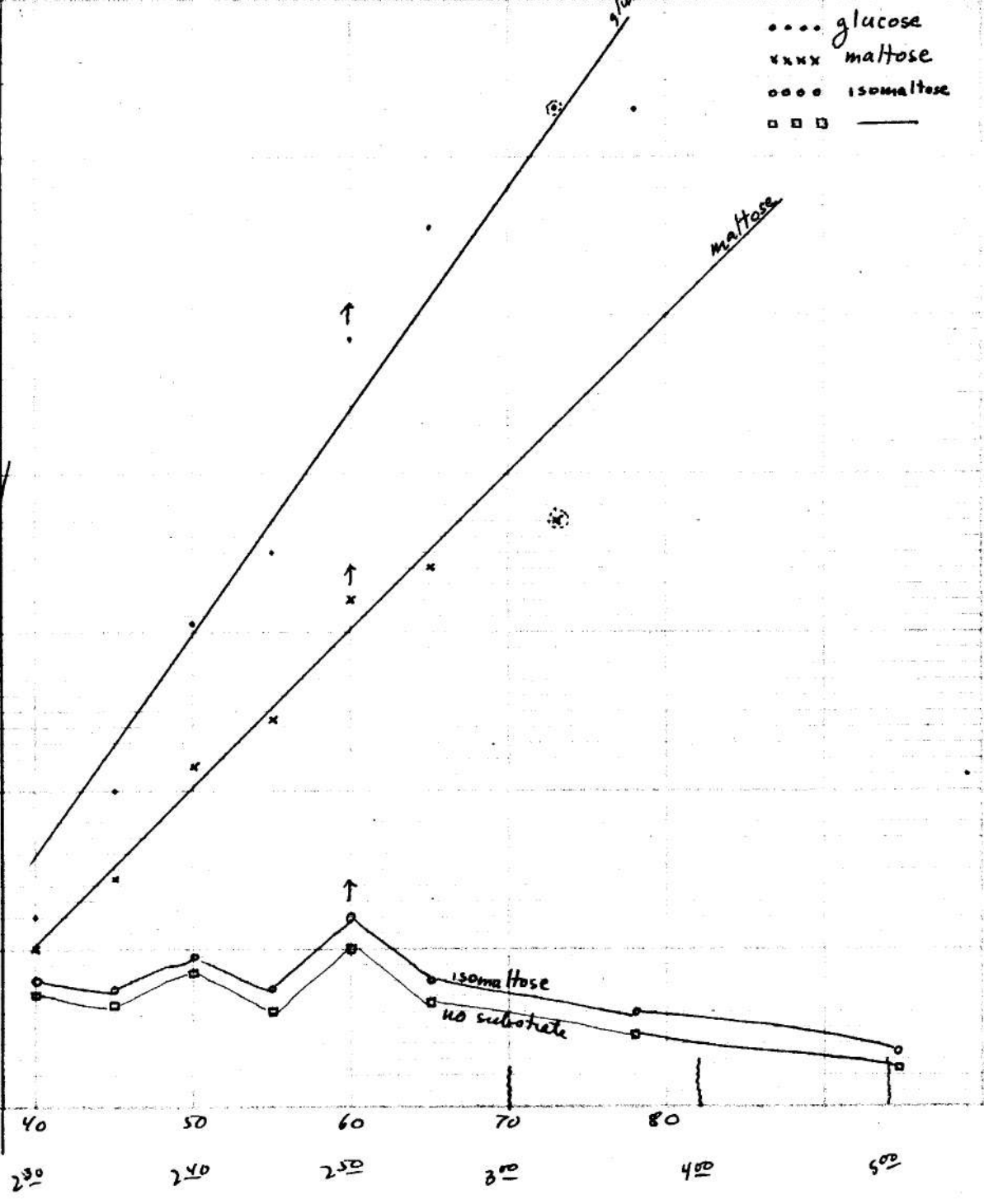
- glucose
- xxxxx maltose
- ooooo isomaltose
- ———

glucose

maltose

isomaltose

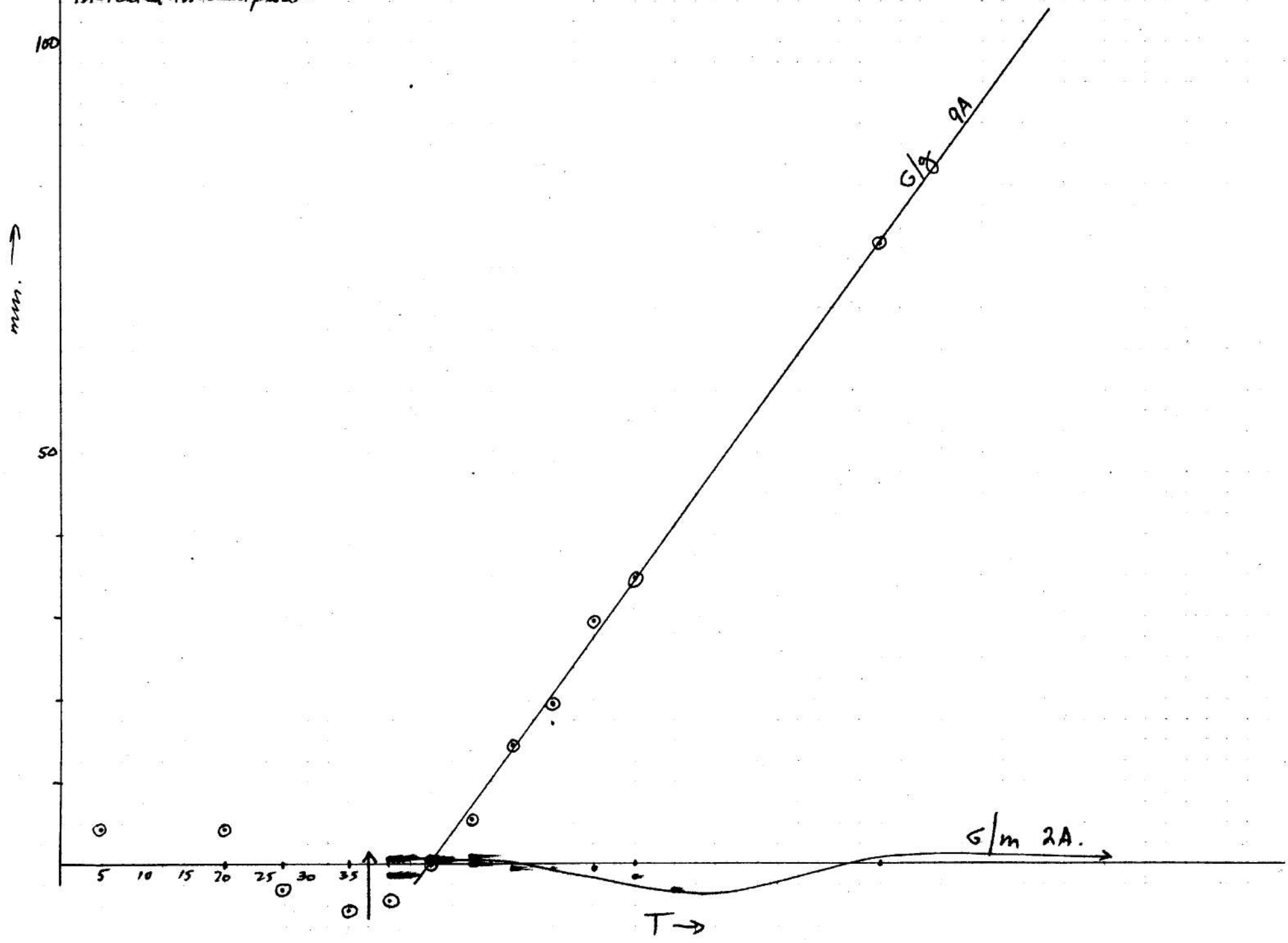
no substrate



J

Some autogenous
 O_2 - or CO_2 - removal
indicated! (or alkali prod).

Utilization of isomaltose



Cross adaptations.

September 8, 1949

	1A	1 6A	2 5A	3 3A	4 13A	5	12A	6 8A	7	T	
	T/T ₂	T/gl	T/mal	T/ar	S/gl	S/ar	S/-				
11:50	0	17	9	18	19	43	18	8		151	
11:55	5	13-4	5-4	14-4	16-3	40	17	9		151	
→											
12:01	11	20 3	23 14	21 3	17-2	46	9	4		151	Stoppfenstück
12:05	15	33 16	56 47	25 7	21+2	66	15	9		151	
12:10	20	43 26	86 77	22 4	19 0	82	13	7		151	
12:15	25	54 37	118 109	19 1	18 -1	100	13	7		151	
12:20	30	73 56	160 151	25 7	26 +7	121	22	14		151	
12:25	35	93 76	202 193	32 14	35 16	131	27	17		151	
		X	X		X						
12:48				38 20	46 27		29	16		151	
1:12				51 32	72 52		36	14		152-1	
1:33				51 31	72 56		36	14		153-2	
2:20				70 43	152 124		46	19		160-9	
3:17				92 56	281		55	23		169-18	
				x							

K12 grown overn. in 1% Trehalose 1/2% (T) or Galactose 1% (D).

Test on maltose, glucose, trehalose, and arabinose

Cells 5x, 2ml in NaHCO₃ 1/20 NaP 1/1000 set 20² 32°

.1ml 10% sugar at →

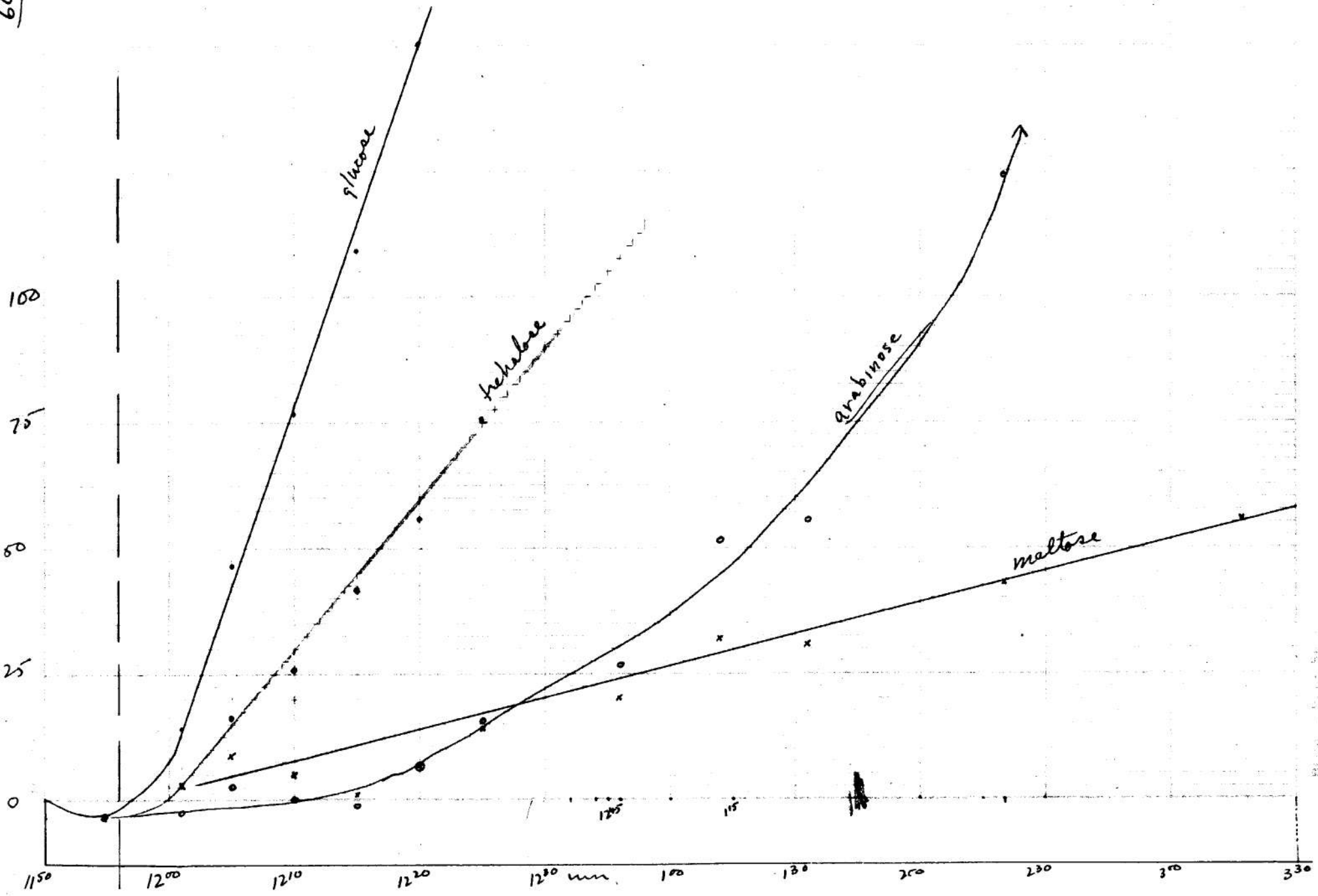
Trehalose // maltose. Need autoferment. control.

Note rapid adaptation to arabinose (30 minutes)

175

Cells grown on trehalose

607



Trehalase in maltose-adapted

607'

		glucose 4	maltose 8	trehalase 10	Thermo
Yml cells in NaHCO ₃	910	27	08	10	148
CO ₂	915	23	06	09	146
M/20					
K12/ Tip Sub.	*				
maltose	920	20	09	16	149
32°	926	38 + 18	29 + 20	21 + 5	149 0
	930	54 + 15	43 + 13	22 0	150 - 1
Bubble later	936	79 + 26	71 + 29	21 0	149 + 1
indam.	940	97 + 16	91 + 18	23 0	151 - 2
	945	118 + 23	116 + 27	25 + 3	150 + 1
	950	137 + 19	140 + 26	27 + 2	150 - 1
	1005			32 + 3	152 - 2
	1102			60 + 26	154 - 2
	1107			61 + 1	154
24m.	99 98		109	04	
hour				26	

Arsenate inhibition of galactose fermentations.

September 9, 1949

K12/lac. 10mg gal in one side arm; 10mg glu in 2d.
 2ml diluted cells from exp. , in NaHCO_3 - $\text{NaPM}/1000$ / CO_2 32°

KAsH_2PO_4

— M/50 M/100 M/200 M/500

1 9A 2 7A 3 4A 4 2A 5 10A T

930 47 18 16 30 03 151

935 47+1 17 0 ~~12~~ 27-2 02 0 150+1

→ galactose 940 95 47 27 8 22 5 39 8 10 6 152-1

945 181 133 45 26 41 14 61 30 58 54 152-1

X → glucose

58 40 61 45 83 53 98 95 151 0

75 58 79 67 105 76 130 128 150 +1

→ glu

89 88 74 119 91 149 148 149+2

111 112 96 X X 149+2

1000

1005

950
95T

Cellozym on lactose

arsenate inhibition

175
608

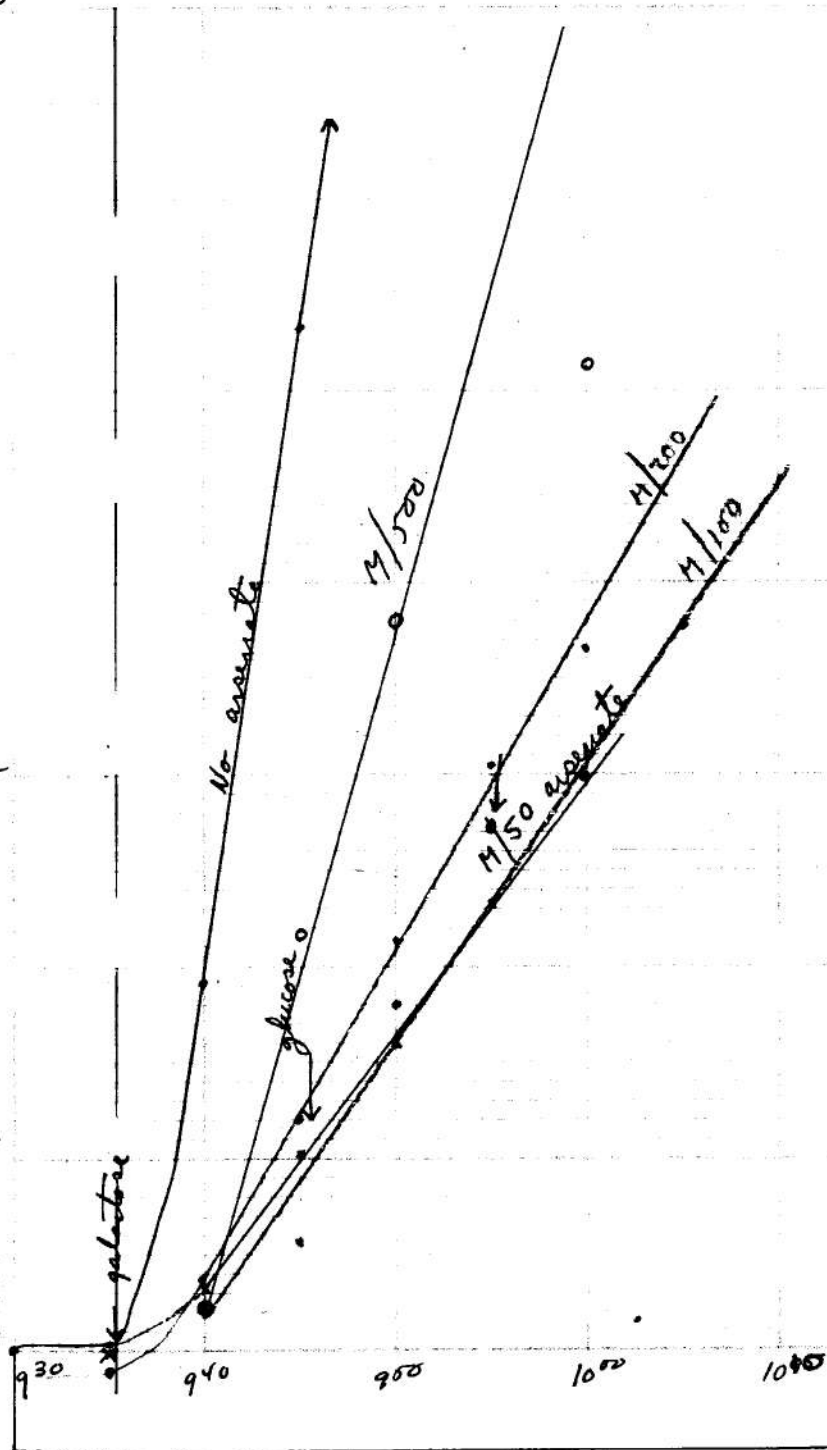
100

75

50

25

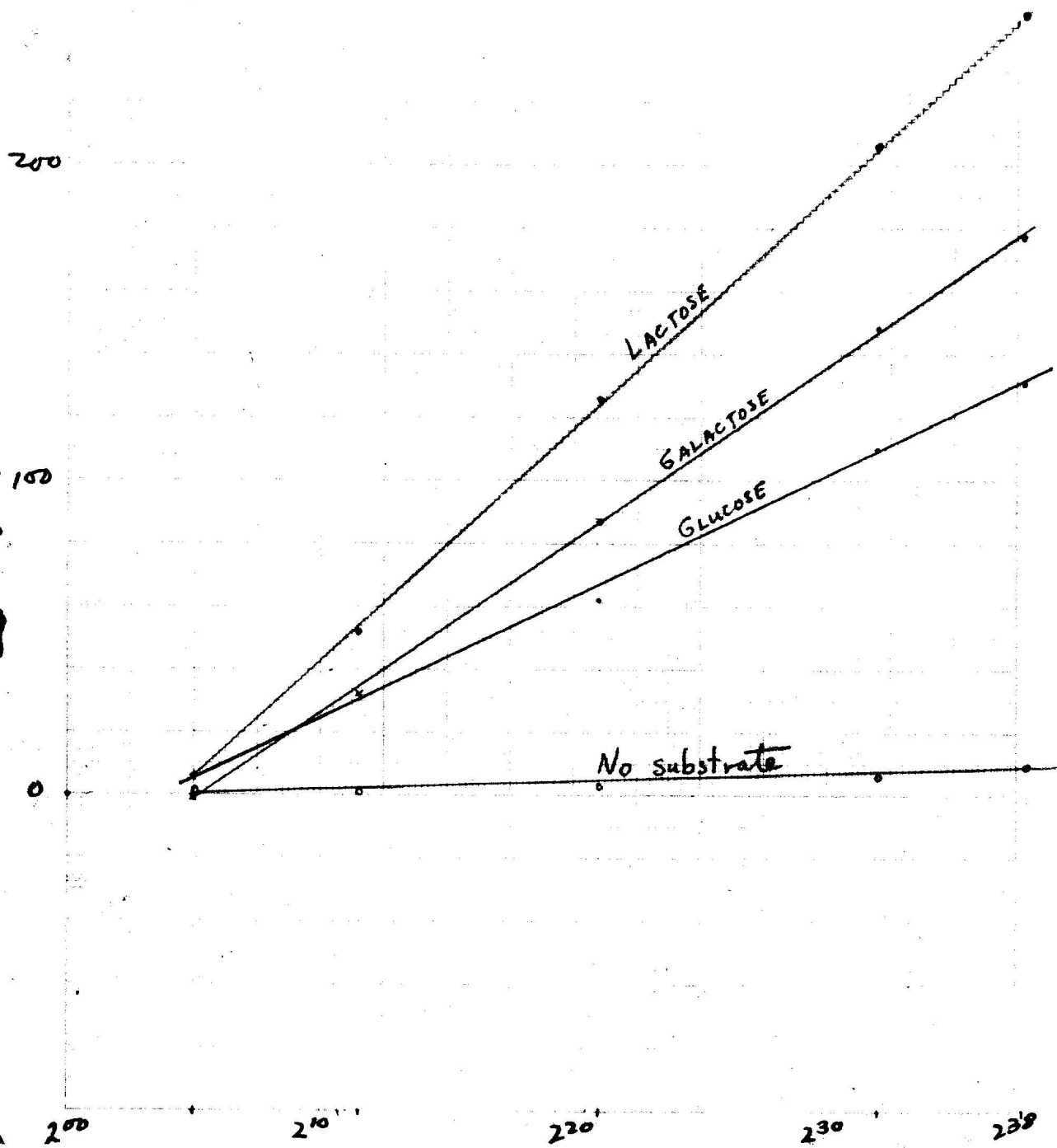
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arsenate appears to inhibit
glucose and galactose glycolysis
indiscriminate.

T.

609a



9/13/49

See

glucose

galactose

lactose

Equilibrate ca 1 1/2 hrs!

	1	1B	2	8B	3	3B	4	5B	T
200	06	04	01	01	09	108			
205	20	5	12	-1	16	6	19	1	117-9
240	43	32	41	32	58	52	15	1	113-5
221	80	62	101	87	137	126	22	3	118-10
232	125	110	161	148	216	206	24	6	117-9
238	143	131	187	177	249	242	24	9	114-6

Stuck out 1B: ca 30% Glu+!

Stuck out culture 1

9/11/49

15 ml cells 1 ml 10% sugars NaHCO₃ 4/20 NaP 4/1000
 Cell suspension in 50 ml x 2 lac overnight \bar{c} aeration. However, the medium, evaporated to ca 15 ml. This may acct. for the poor lactose activity seen here.

glucose galactose
 Fructose
 lactose
 D-glucose

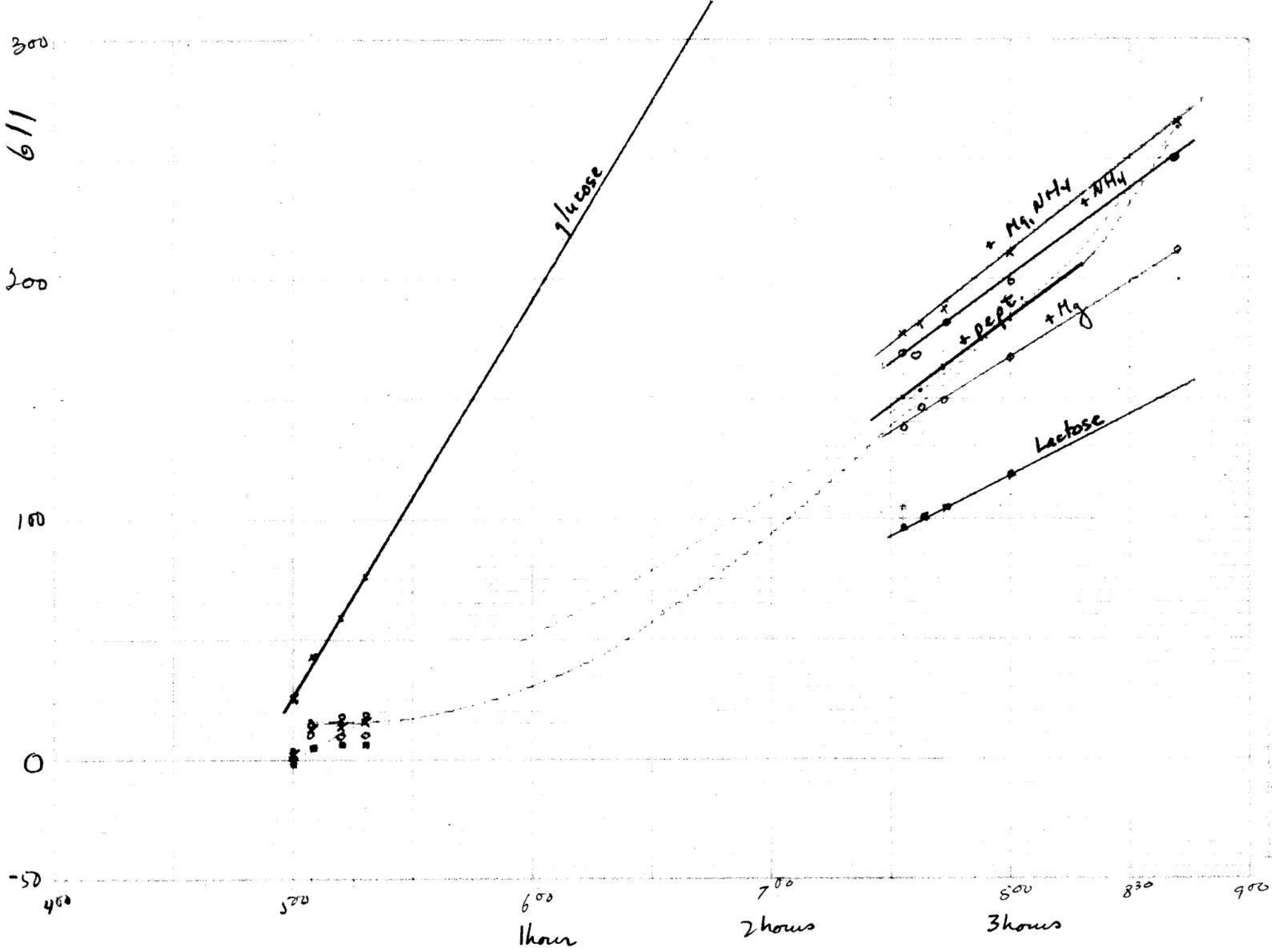
1240
 1245
 1246
 1250
 1255
 100
 110
 115
 121
 131
 150
 205

	1 5B	2 2B	3 6B	4 4B	T
	-2	18	-3	16	147
	-1	23	04	27 (153)	147
	05	28	08	27	153
	10	32	12	31	157
	04	28	08	25	151
	04	30	10	20	148
	14	40	19	31	153
	22	48	25	32	150
	28	52	27	29 (151)	154
	41	69	42	35	156
	62	84	51	27	152
	89	114	68	29	152

$$\alpha_{32^{\circ}}^{CO_2} = ca .63$$

Subtract

	Volume	$R_{0ml 32^{\circ}}$	1ml	2ml						
A	1									
	2	21.12	1.88							
	3	19.51	1.74							
	4	20.19	1.80							
	5	19.97	1.78							
	6	18.20	1.62							
	7	18.43	1.64							
	8	18.99	1.69							
	9	19.02	1.69							
	10	18.44	1.64							
	11	19.60	1.75							
	12	18.86	1.68							
	13	19.61	1.74							
	14	18.26	1.63							
1.82	1	19.81	1.76							
	2	19.88	1.77							
	3	20.45	1.82							
	4	20.85	1.86							
	5	19.85	1.77							
	6	18.95	1.69							
	7	20.47	1.82							
	8	19.67	1.75							
	9	18.86	1.68							
	10	19.62	1.75							
T	19.11	1.70								
			subtract							
			.0891							
			subtract							
			.178							



5

September 23, 1949

540

		D _i	D _{OMP} 10 MIN.	Δ
M1	L1 water	007	438	384
M2	L2 glucose	002	217	166
M3	L3 lactose	0	165	116 !
M4	L4 water	001	072	022
M2	glucose	001	058	008
M3	lactose	0	071	022
M4	Mg	0	074	025
-	lactose	-007	042 = 49	0

Cell density L 19.9
M 13.3

Cells incubated from 3³⁰ PM
in indicated supplement:

- 1 ml cells
- 12 ml 1% sugar
- .1 ml KP buffer pH 7.0 M/5.

(4) + .1 ml MgSO₄ 4/5.

K12 / mal and / lac

showing decrement of activity
when incubated with lactose or glucose!

800

- L1
- L2
- L3
- M1
- M2
- M3
- M4

vs OMP 6
flambs
mg. exp. 66

- D_{OMP}
- 10 M
- 438
- 125
- 54
- 20
- 20

September 24, 1949.

2 PM

A) 5 tubes each receive 1 ml K12/vac. # 6 the same, dil. 1:10.
Add 1 ml benzene / tube.

5 PM add 9 ml H₂O to 1-5. (# 2 merib.).

Assay 2 ml samples.

	Di	8 ^{pm} D ₅₀₀ 9
1	010	400 71c!
2		157
3		173
4		163
5		172
6		205.

Too erratic to be used in present stage of development.

Effect of N-supply on lactase deadaptation.

611a

September 24, 1949.

12 hour cells aerated
mice washed

Hawest K-12 from Y2 Mal and Y2 Lac.

Add NaP 7.5 to M/50. 1ml cells + 1ml supplement

incubate from 12⁵⁰ to 3⁵⁰ PM = 3 hours. 37°

Add .1ml benzene to activate.
on pg M/2000 in M/100 NaP 7.5 37°.

cell density (before after 1:1)
mmio. 6.7

Suppl.	Di	10 m. Dongy	A'
1 Y2	029	387	
2 Y2 lac	027	590	
3 lac 1%	028	217	
4 H ₂ O	024	236	
5 lac .2% + (NH ₄) ₂ SO ₄ M/10 .1ml	022	286 264	?
6 (NH ₄) ₂ SO ₄ M/10 .1ml	023	264	
7 lac .2%	022	364	
8 —	003	011	014

M/1 Na₂CO₃ 1ml added

W251a/lac

	T	1 SA	2	2A	3	4A	4	13A	5	3A	6	12A	7	6B	8	10A	9	9A
150	155-53	30	22		32		44		39		33		57		46		45	
155	154-48	32	24		34		43		35		29		51		41		38	
200	157-53	32	24		34		45		38		32		56		45		43	
201																		
205	158-54	30	24		33		44		36		33		57		45		43	
210	157-48	28	24		32		42		35		34		57		45		40	
215	158-53	28	22		31		41		34		36		59		49		42	
220	150-58	27	21		30		44		38		39		66		60		49	
225	150-59	30	26		34		46		44		53		74		67		51	
230	161-60	35	30		39		53		51		63		82		73		52	
235	161-61	48	31		40		53		51		63		80		75		52	
240	154-58	44	39		48		64		62		76		88		81		51	
245	157-60	51	46		51		67		66		80		90		85		51	
250	162-58	57	48		54		70		68		84		93		89		51	
255	162-56	60	50		55		71		71		89		96		92		49	
260	161-57	62	51		57		73		75		93		100		97		50	
265	162-55	62	52		57		71		80		101		104		103		47	
270	158-55	70	53		58		79		84		106		107		108		46	
275	160-52	75	57		62		83		90		116		113		116		44	
280	158-60	90	71		73		100		119		150		141		148		55	
285	155-58	102	76		75		104		128		164		150		161		50	
1	LAC	1 mg.																
2	GLU+GAL	.5 ea.																
3	GLU	.5																
4	GAL	.5																
5	LAC	10																
6	GLU+GAL	.5 ea.																
7	GLU	.5																
8	GAL	.5																
9	—																	

530	163	127	108	99	135	182	219	191	209	155
	1 ml cells	W251a/lac aer.			in NaHCO ₃	4/20		.05 - .10	in sidearms.	
645	179	127	127	110	137					55

Strained out on EMB glucose: essentially pure Glu -!
 (99% -) But note overall slow fermentation.
 Culture may have gone too acid.

Gal'ase activity in unadapted cells.

Sept. 30, 1949.

Harvest K-12 from 12 hour aer. 42 - 50ml. conc. to 5ml (10x)
 Leave water suspensions on table top 10A - 7:32 P 30.

1 ml aliquots incubated in benzene 7³⁰ - 9¹⁰ PM (90 mins.)
 Test samples per standard ONPG (1/2000 mg; 20 mins; 37°; NaP 7.5
 7/50

Untreated samples: (.1 ml / 10)

	Di	Dampg (12 min.)	R.A./ml	R.A./ml / Di, 100 / 10 ³	
K/lac	250	800	94	38	
K/Hcl	307	475	19	6.2	17
K/glu	118	119	0.2 ±	.0.2	1 ±

TREATED

(.01 ml) K/lac	017	540 (7 min.)	1.5×10^3	.58	100
(.01 ml) K/Hcl	027	380	$.36 \times 10^3$.12	21
(.1 ml) K/glu	070	269	$.02 \times 10^3$.02	3

Activation of ca $\frac{1500}{94} = 16x$ fairly consistent here, but
 1 1/2 h. may not provide maximal activation with benzene.

Lactase is present in glucose and especially in maltose-adapted
 cells.

Gal'ase activation in K12
Octyl alcohol, thymol, benzene

Oct. 1, 1949.

Hewlett K-12 12 hr. aer. 42/- 50 ml. Wash 2x and conc. 10x.

1 ml aliquots to small tubes and incubate in given reagent.
Assay standard mpy.

Intact Cells.

A.	Concn	Dint.	Di	D _{comp}	R.A. / Di = 100	
1	1 Lac	.025	043	140	2/0	(100)
2	1 Mal	.1	140	167	21	10
3	1 Glu	.2	141	130		
4	1 Lac	.1	054	218	293	100
5	1 Mal	.2	129	193	51	17
Benzene treated tubes?						
1	Lac	.01	007	310	292	171 (100)
2	Mal	.01	007	046	29	21 (12)
3	Glu	.1	040	062	1.5	(2)
4	an. Lac	.01	0	169	158	292 (100)
5	an. Mal	.01	0	072	61	43 (17)
Octyl alc.	1 Lac	.01	007	418 (11m)	750	

Note superiority of octyl alcohol activation.

P1.	Di	D _{comp}	R.A. / Di	
Octyl Alc (.1 ml)	1 012	268 (5)	573	(100)
	2 003	061	34	6
	3 043	052	2	< 1
	4 001	367	657	(100)
	5 -002	110	85	13
Benzene	1 012	230 (5)	484	(100)
	2 005	061	32	7
	3 042	056	5	1
	4 -001	230	418	(100)
	5 -001	073	49	12
Thymol (crystal)	1 029	419 (5)	932	

Octyl alc. > Benzene

Thymol >> Octyl alcohol.

Test O₂OH; Thymol for partition of NO₂OH at pH. 7.5.

Octyl alc 1:70
Thymol

D_{comp} 11/50,000
100
88
99.

Neglig. diff. even if carried over

Kinetics of thyroxine activation
Gal'ase in W842. (test).

Oct. 2, 1949.

K/Lac of 10/1/49.	A) 1ml unshaken, 37°.	B) 5ml in 10ml cent. tube	
Add a crystal (10-20mg) of thyroxine at 4 ¹⁵ PM. .005ml samples C = phenol. 1ml start at 4³⁰			
T.	Mins.	Donpy.	
420	5		Note: slow process. Needs >> 1 hr.
	A } B }	No visible color	
440	25		
	A } B }	251 103	
500	45		
	A } B }	444 126	
	(30 MIN) C	132	
700 800	130 215		Some evaporation possible.
	A } B } (185) C	> 1000 650 231	

P2. Hawate normal W842/Mal; W842/Lac K-12/Lac.

	Dist	Di	Donpy	RA	
<u>Zello.</u>					Activation = $\frac{655}{178} \times \frac{5}{2} = 9.2$ fold
K/L	.02	052	149	178	
W/L	.05	172	150	-	
W/M	.05	130	109	-	
<u>Thyroxine</u>					No activity!
K/L	.005	004	670	655	
W/L	.005	019	018	0!	
W/M	.005				

Consistency of Gal'ase activation by thymol, octyl alcohol.

1: .5 ml susp.

all in duplicate

~~2: .5 ml + 1.5 ml H₂O~~

2: .5 ml " 4.5 " "

A thymol

3: .5 ml ($\frac{1}{10}$) + 4.5 ml "

B ~~to~~ octanol

C benzene

1/2 hour tests

Make up to 10 ml (exc. 3)

Test .1 ml samples 1, 2; .5 ml of 3 ($\frac{2}{3}$)

Nuglet D: (.007 ± 0.3). Add NO₂, CO₂ to terminate Rx.

	A (Thy)	B (oc)	C (B ₂)
1	318	131	200
1	359	118	171
2	062	054	054
2	060	057	062
3	082	069	067
3	093	053	064

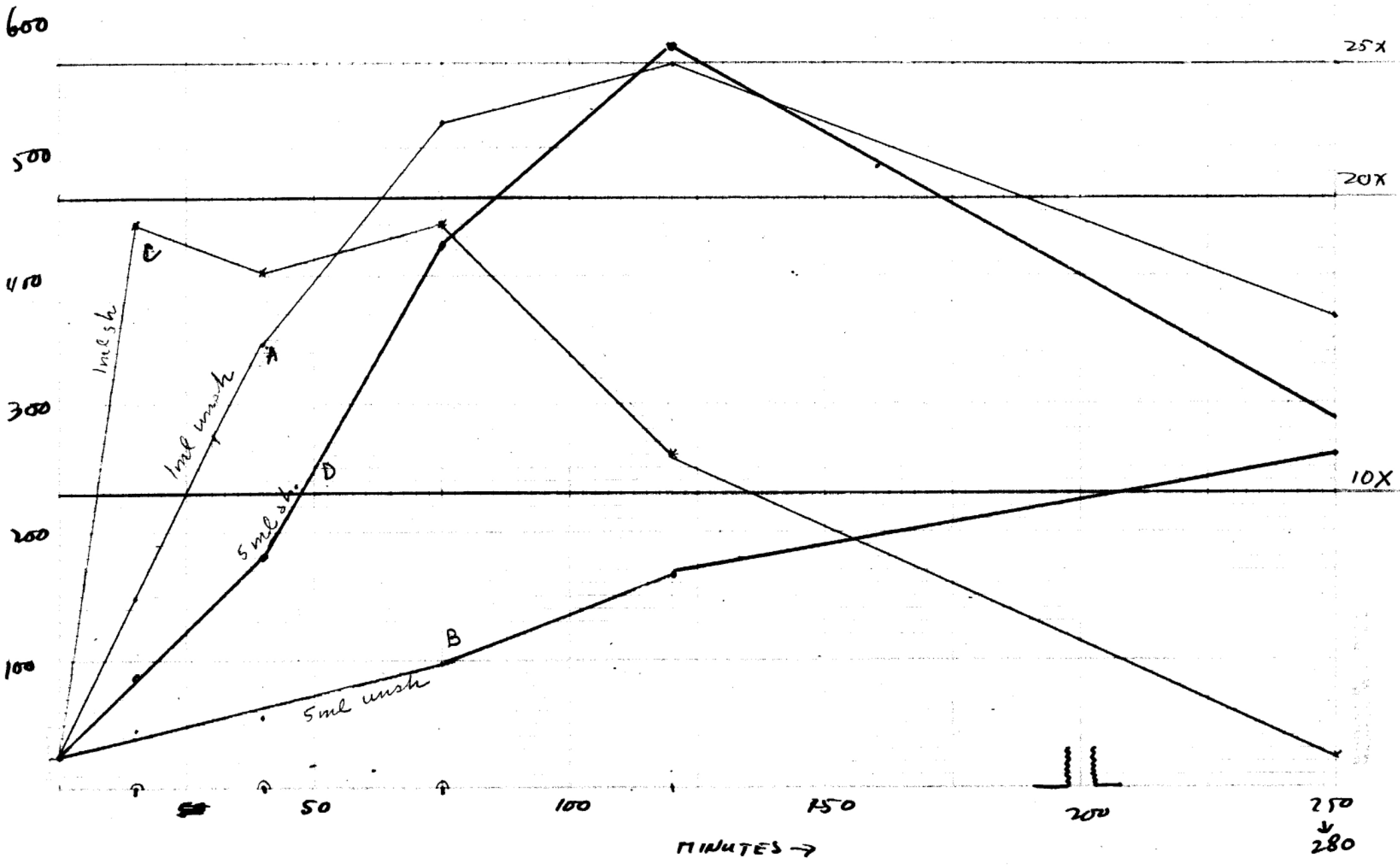
Time may have been insufficient for complete activation! Thymol seems to act most rapidly. I try Phenol, other ϕ -OH's

Re assay 1, 3 4P2.

A: ~~4~~ 1773.

Kinetics of gelase activation
by thymal

624



3

Kinetics of Gal'ase activation. Effect of shaking

10/3/49.

Harvest aer. K/Lac conc 50/20. H₂O. Add Thymol: $3 \frac{PM}{20}$.

A) 1ml unsh. B) 5ml unsh. C) 1ml sh D) 5ml sh.

Remove 1ml samples from time to time; dilute in water 10ml and assay.
Terminate with Na₂CO₃, etc. to fix cells. mid. units.

335 T MINS

15 Di 20 318 23

1ml 0 089 318 23

10ml 003

15 A 149

15 B 46

15 C 460 (440)

15 D 87

400 40

1/2 A 173 x 2 = 346

1/2 B 53

1/2 C 201 x 2 = 402 (!)

1/2 D 180

10% < above!

435

1/2 A 260 x 2 = 520

1/2 B 98

1/2 C 220 x 2 = 440

1/2 D 423

520

120 A 281 x 2 = 562

120 B 88 166

120 C 150 260

120 D 289 x 2 = 578

800

A 369

B 260

C 025

D 289

2 hours optimum for unshaken cultures.

	2 1/2 h.	184.	2 1/2 h.	1ml. treated 545 - 815	test + compare:
Thymol	479	178			
phenol	016	—			
benzene	466	685			
octanol	369	222			
			Repeat overnight.		

Gal. use of adapted + unadapted cells; Lac, -

624a

October 5, 1949.

a) W112 harvested from 1/2 Lac; 1/2 Mal; K-12/Lac. as above.

① Inert cells.	Di	Don pg
K/L ^{10/ml}	131	710
W/L	98	—
W/M	124	—
② Benzene 24 hours		
K/L ¹⁰¹	006	590 (12.5 min; Na ₂ CO ₃)
W/L ¹¹	073 068	261
W/M ¹¹	068 073	092

b) K12 from 1/2 Lac; Mal; Idler. Ser. [Benzene from 12N ± I. ca 8 hours.]

Inert:	K/L ¹¹	130	520	A corr	RA	n/mg
	K/M ¹¹	129	151	392	3.02	14
	K/G ¹²	204	182	24	—	0.9
	—	—	007	—	0	0
	—	—	± 004	Counter = +11		
Benzene	K/L ¹⁰⁰⁵	-004	410	403 ^{x20}	62	297
	K/M ⁰¹	+004	074	59 ^{x10}	4.6	22
	K/G ⁰¹	074	060	—	—	—

Antart

L
M
G
-

1
.05
.1
.1
.1

Di
087
157
130
214

Dong
246
189
124
250

Bz
4h.

L
M
G
-

.005
.02
.1
.02

0
018
069
032

530
194
074 (535 PM)
267

K12 harvested from yeast - peptide (VP) / sugar. 50ml/10ml.

	Di	Donp9	A	R.A.	u/mg
Lac	173	408	231	134	6.4
Map	181	177	8 ⁴⁰	2	0.1
-	122	125	10	4	0.2

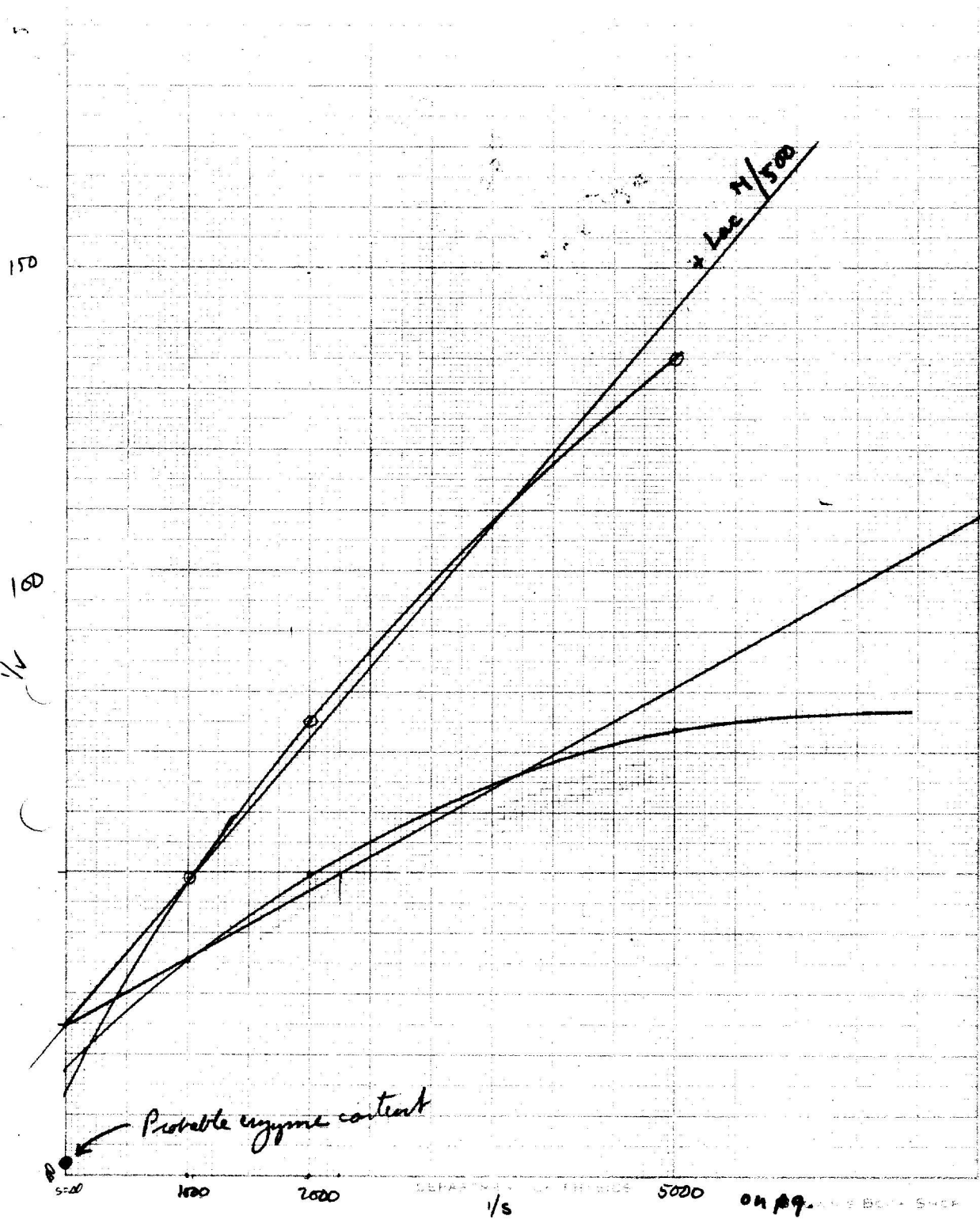
182⁴⁰⁴
131
maybe inaccurate

	Di	CHMS.	A'	R.A./Di	R.A.	R.A./Lac	u/mg
Lac ¹⁰⁵	005	212	196	380		100	408 300
Map ¹¹	104	174 ²⁰⁴	69	38		10	1.8
- "	080	141 "	58	47		12	2.3

Activation: $20 \times 196 \times 3\frac{1}{3}$
231. 57x !!

cell

with.



Probable enzyme content

$\frac{1}{2} \frac{1}{500}$

1/s

on pg.

Kinetics of Gal'ase in intact cells.

Oct. 7, 1949.

		K-12 harvested from Lac Y2				
K _{onpg} and K _{lac}					M/100 NaP	
	K _{onpg}	lac	Di	Na ₂ CO ₃ Donpg		1/V
cells	100			352 ✓	282	35.5
	200			274	202	49.5
	500			183	136	73.5
cells	100	M/500		274	204	49.0
	200	500		185	133	75
	500	500		121	74	135
cells		—	089	050	047	
no cells.	100			020		
	200			002		
	500			-003		

Graph calc: $V_{max} = 1/25 = \underline{400}$

$K_{onpg} = M/2000 = 5 \times 10^{-4} M$ ✓ per meas.

$K_{lac} = [Lac] = 2 \times 10^{-3}$

Note: In extracts + cells, cf: (K_s): (x 10⁻⁴)

onpg	cell	ex
	5	1.3
lac	20	14

i.e., transport block to lac
 << onpg. But still note
 that the 1/5 : 1/v plot do not
 extrapolate to the full V_{max} for
 extracts! Possibility of
 bending needs to be rechecked.