

4/2/49.

Compare carbohydrate utilization by cell suspensions harvested from 20 hour lac Y2 broth, unshaken of (A) W760 and (B) W815.

Add 10 mg sugar to 1ml cell suspension and 1ml buffer BCP.
(uM)

	A	10m	15m	5m	10m	B	15m	20m.	25	60
1	glucose	+++	---	-	-	±	-	±	±	+++
2	galactose	+++	---	-	-	±	±	±	±	+++
3	lactose	+++	---	±	++	+++	+++	+++	+++	+++
4	butyl galact.	+++	---	±	++	+++	+++	+++	+++	+++

Butyl galactoside is fermented much more quickly than galactose.
(ca 3x)

Is glucose accumulated from lactose? cf. W255 and W815 grows on lactose. Also W1089L3+. J.

Query? does galactose permeate the cell? Use inhibition of galactosidase.

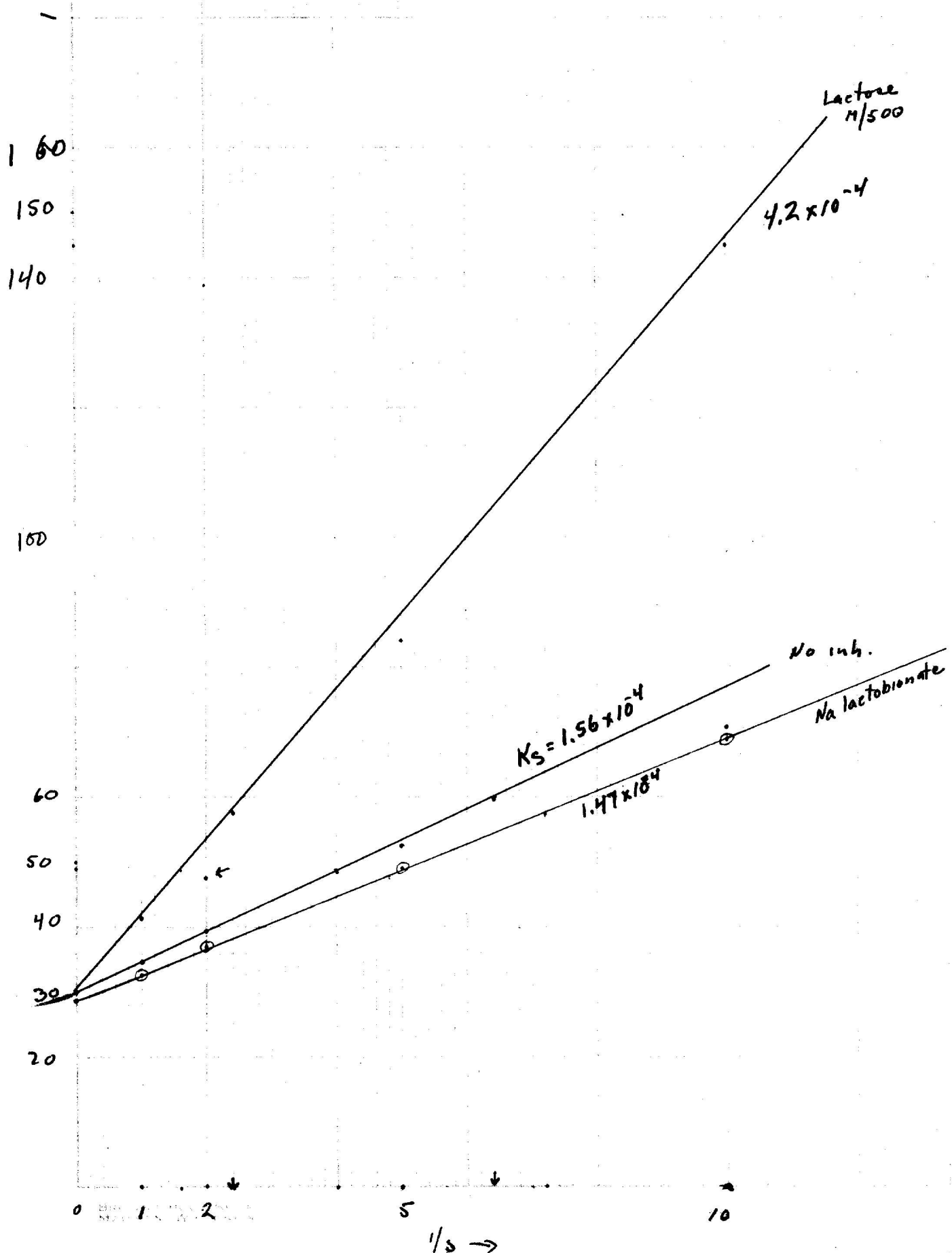
Competitive inhibition of galactosidase

4/3/49

Extract 399 dry cells. Dilute 1% aqueous extract 1:200 and use 1 ml aliquots.
 NaP buffer pH 7.5 M/50.

ONPG M/.		Di	D _f	D _{cor}	1/v
1 100		020	307 ✓	289	34.6
2 200		010	263	254	39.4
3 500		002	193	191	52.4
4 1000		-003	129	132	75.7
11 100	Lac M/500.	020	261	243	41.1
12 200		013	221	210 209	47.8
13 500		003	122	119	84.0
14 1000		-001	68	69	145
21	Lba M/500	021	338	319	32.4
22		013	281	269	37.2
23		005	209	204	49.0
24		003	147	144	69.4

Lba = Ca lactobionate; Ca replaced by Na i oxalate, benzin and Na₂SO₄.
 Make substrate etc. to 9ml. Add 1ml enzyme dilution at to. 36°.



	D_i		D_f
1. —	177 177 177		550
2. Azide	178		520
3. lac	180		540
4. Azide + lac	190		570

Glucosams.

	D_i		D_f
1. —	178	160 + 010	165.
		= 170	

Competitive Inhibition

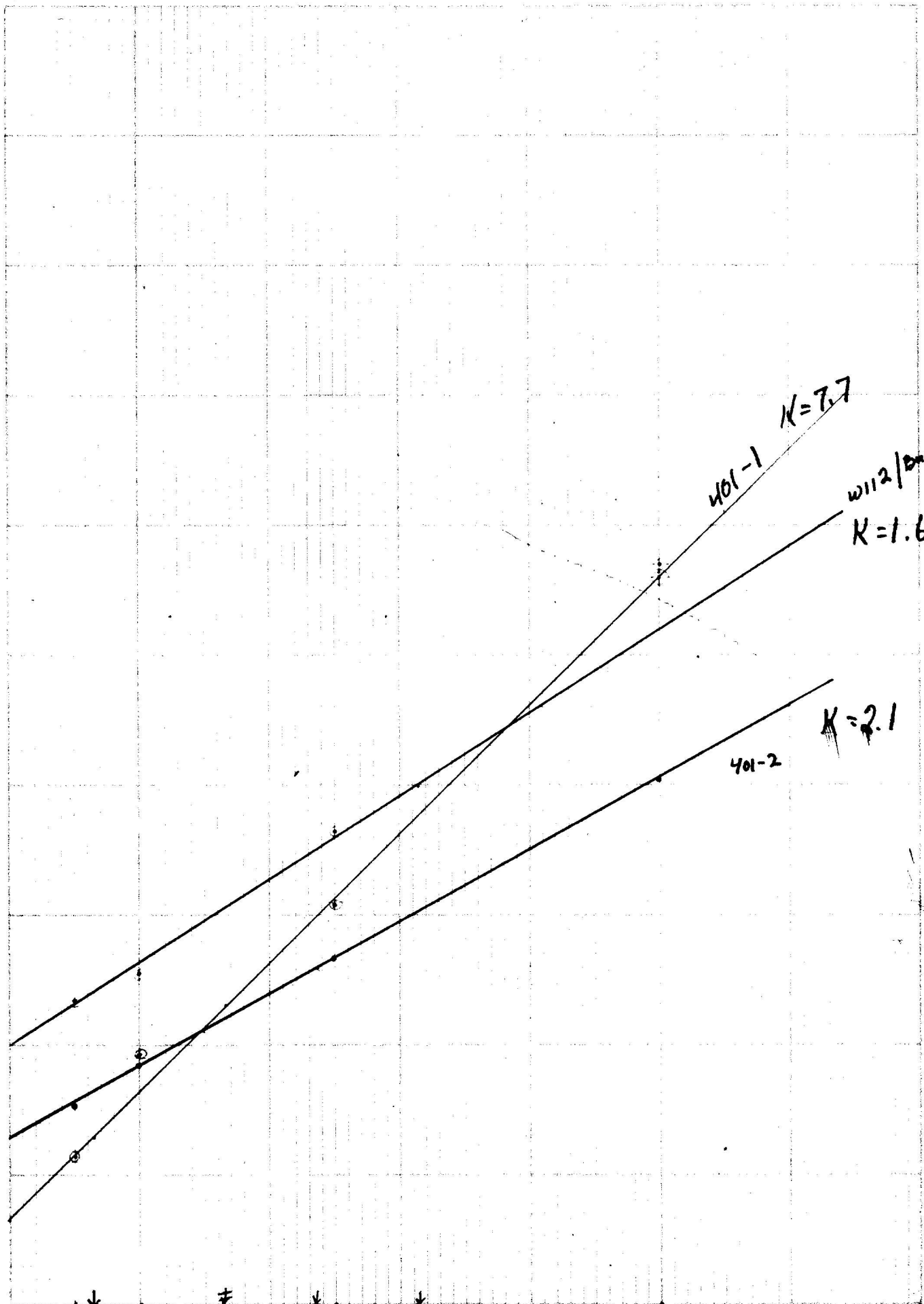
4/4/49.

	[399:200]		NaP 7.5 M/50 + Na ₂	SO ₄ M/50 · 1/4	
A	1	—	023	387	368 27.2
	2		010	339	330 30.2
	3		003	259	256 34.1
	4		—	180	180 55.6
		Lactitol lactobionate			
	11	M/400	021	360	343 29.1
	12	"	012	301	290 34.4
	13	"	001	210	209 47.8
	14	"	002	141	139 71.9
	21	Bugal	027	278	256 39.1
	22		017	203	189 378 52.9
	23	M/500	009	109	102 18.0
	24		005	62	58 17.2
	31	Megal	024	379	359 27.8
	32		017	330	316 31.6
	33	M/500	007	244	238 42.0
	34		004	173	170 58.8

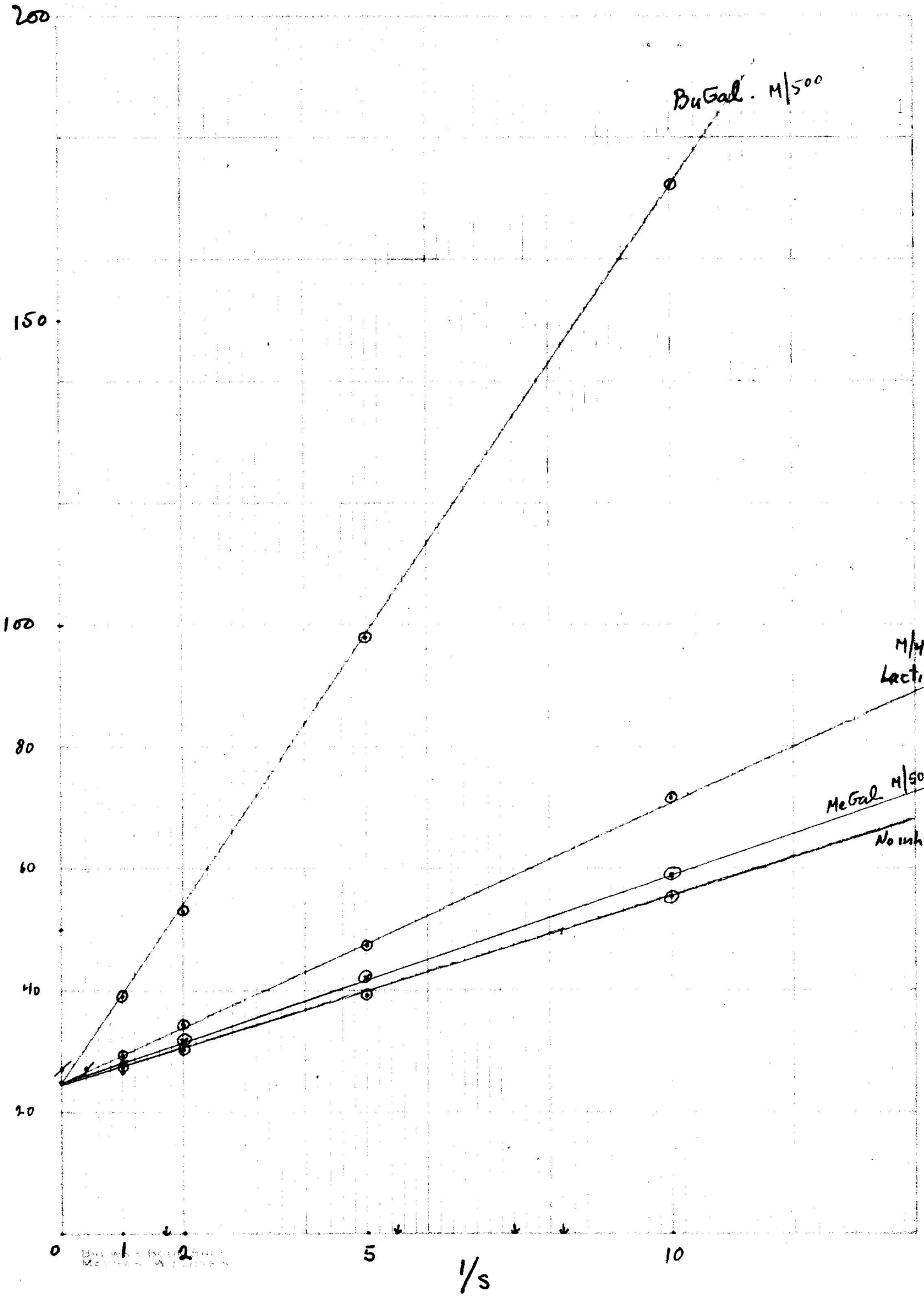
count is increase from 9 to 11 ml. Subtract 0.9/11 of Di from Df.

Apparent Km : $\times 10^{-4}$

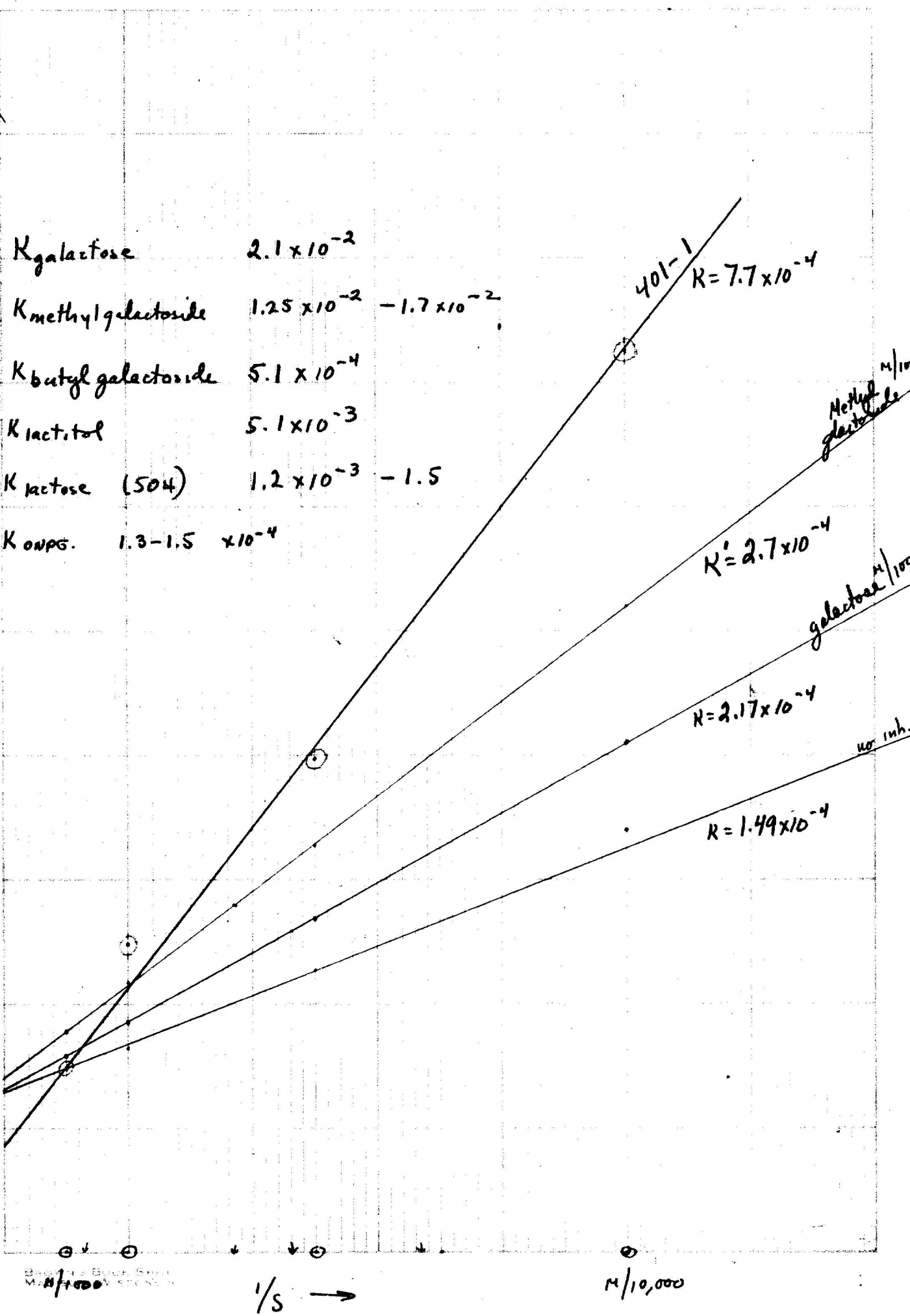
- Blank: 1.22
- Megalactoside 1.35
- Bugalactoside 5.9
- Lactitol 1.82



1/V →



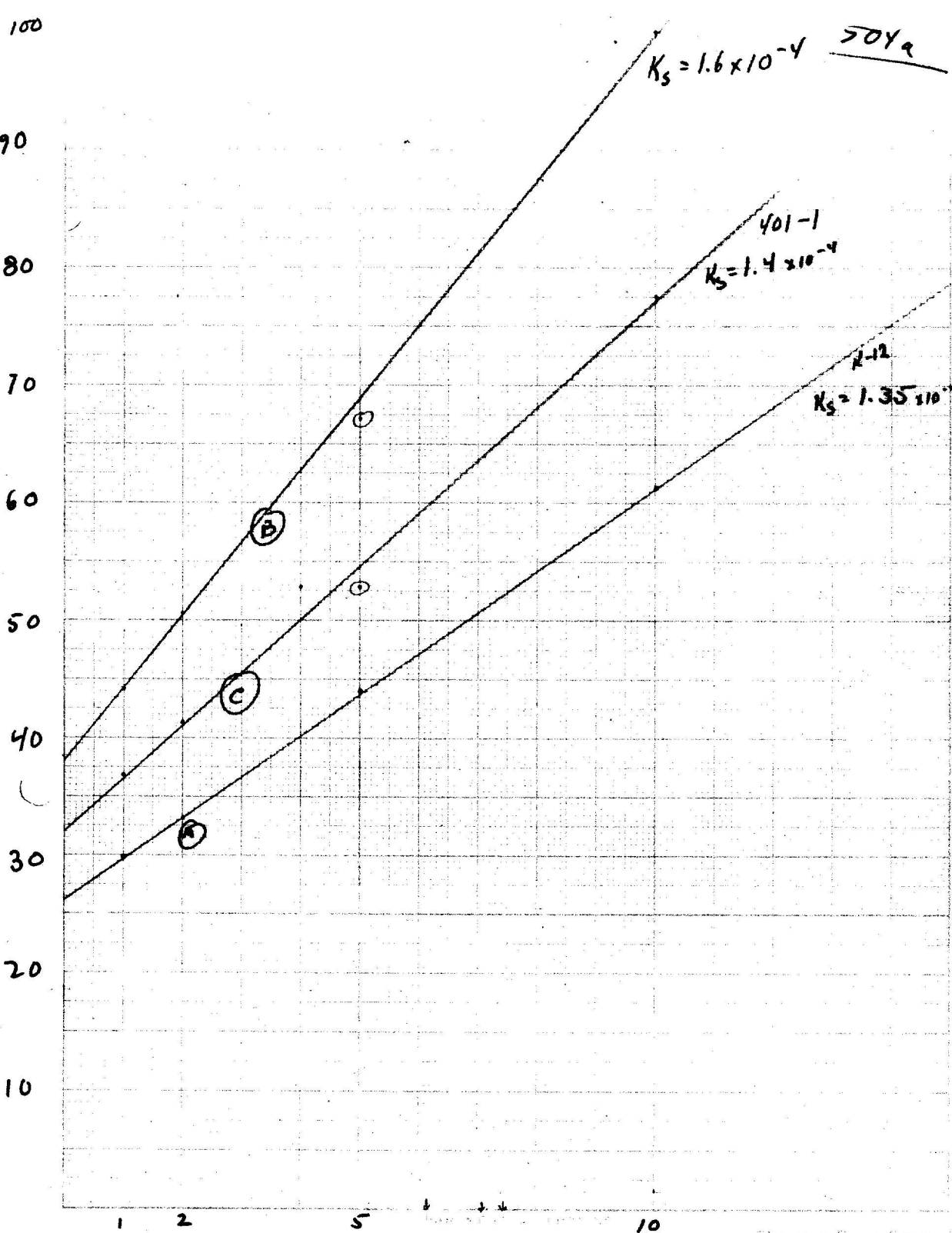
1941 W. H. B. ...
M. ...



see 384

100

Beckman DU-40 Spectrophotometer



Example B. 1. Sheet

Kinetics of suppressor lactases 504a

4/7/49

1/5 ONPG NaP 7.5 M/100.

	100/M	D _i	D _f	V _{con.}	1/V	K _s	V _{max}
A 399 (K-12) 10 mins.	1	017	358	344	29.9	1.35	
	2	009	309	302	33.1		
	3	003	230	227	44.0		
	Y	0	163	163	61.3		
B (Vol-1)	1	023	240 245	226	44.2	1.6	
	2	013	209	198	50.5		
	3	006	154	149	67.1		
	Y	—	100	100	100. —		
C (Vol)	1	022	240 290	272	36.8	1.4	
	2	011	251	242	41.3		
	3	003	192	189	52.9		
	4	006	134	129	77.5		
D with bungal. prep. 10 mins.	1	019	760	744		excess enzyme	
	2	013	680	669			
	3	003	500	500			
	Y	006	331.	325			

These determinations show no unusual deviations!
and are consistent with 504

4/5/49.

Grow K-12 overnight in 200ml 42 Megalac. 12%
Harvest P5 and dry over P₂O₅.

Yield: 85 mg dry cells.

Triturate and extract 40mg / 10ml H₂O for extract 506A.
Extract potency ca. 600 u/ml.

4/5/49.

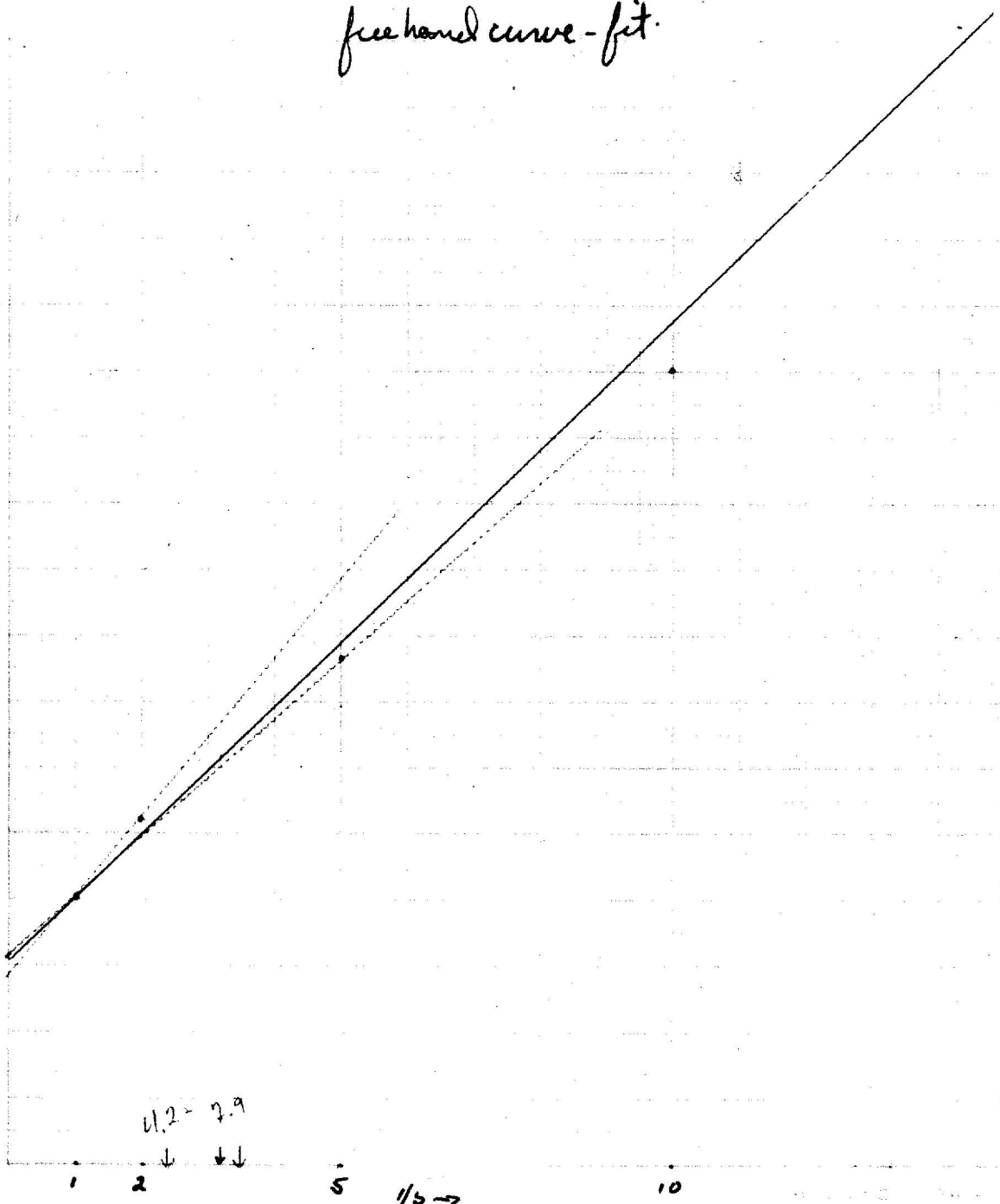
Grow K-12 in 2 x 50ml 1/2 Malt 1%	harvest and
dry over P ₂ O ₅ .	Yield: 29 mg.

Intracellular lactase.
free hand curve-fit.

150

100

50



$1.2 = 7.9$
↓ ↓

1/5 →

Kinetics of cellular lactase

508

4/6/49.

K-12 harvested from Y2 Lac. 5X. then .2ml in 10
NaP buffer 4/100 pH 7.5

ONPG	Adg.	Est. Cou.	V + 0L. 7	V	1/V
4/1000	346	370-358	338	249	40.2
2000	283	299-271	281	192	52.1
5000	220	222-221	219	130	76.9
10,000	167	178-173	172	83	120.5
12500	260		240		
∞		0	89		

(stirred vigorously).

$$V_{max} = 322.$$

$$K_m = 3.2 \times 10^{-4}$$

Stirring does not stimulate enzyme action!

K_m is here at least twice that of isolated enzyme.

Kinetics of enzyme from lactose
and fungal grown cells.
Temperature coefficients at enzyme saturation

4/7/49.

1+2 at 37° 3+4 at 22° 0.145 M/1000 NaP M/100

1,3 K-12/lac cells, Di controlled.
2,4 K-12 (399) extract.

	Di ^{4:41 PM}	D ₂₀		V _{con}	D ₃₁
1	22		461.111	342	612
2	25		307	287	457
3	23		262	142	319
4	20		159	143	231 231
	cells		101		

$Q_{15} \text{ extract} = \frac{287}{143} = 2.01$

$Q_{10} = 1.6$

$Q_{15} \text{ cells} = \frac{342}{142} = 2.41$

$Q_{10} = 1.8$

or calc. $Q_{10} = (Q_{15})^{2/3}$

Note: Q_{10} cells is higher than Q_{10} extract at this high substrate concentration.

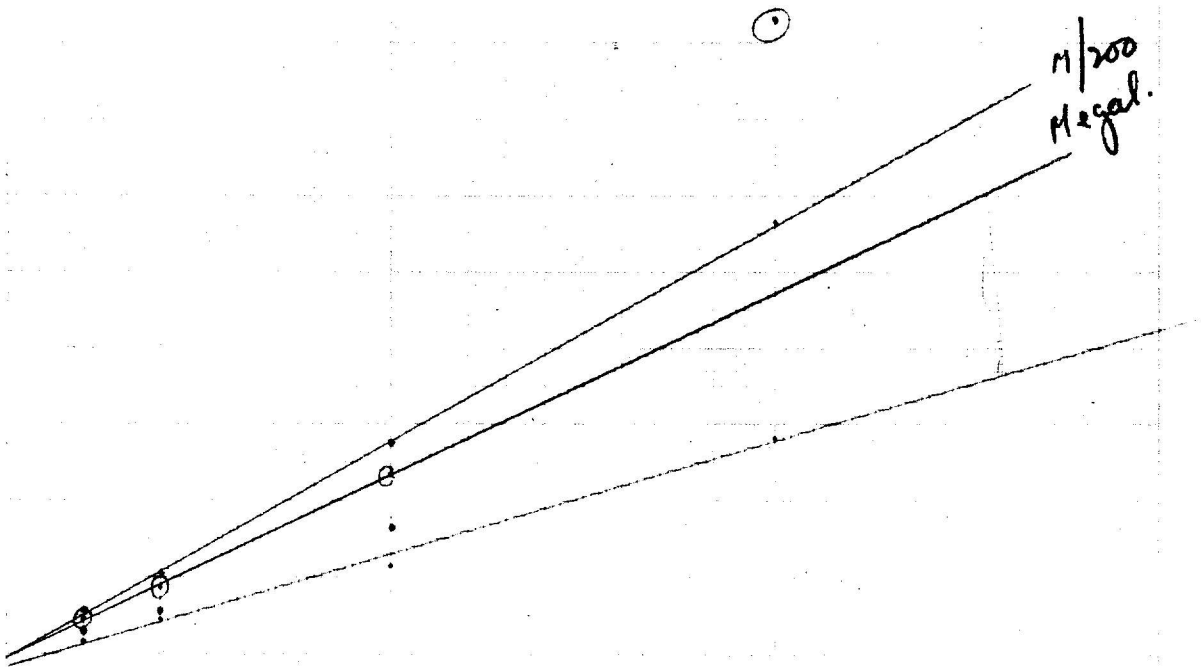
2:25 K-12 + W-349 gram on lactose tested on lactose; lactitol.

K:12: +++ on lac ++ on lol in 5 mins. Blue+++ + lol in 10 mins.
W349. — —

Add a glucose pair at 20:30 2:55: W349 ---

~~510~~

510



4/7/49.

NaP M/100 7.5 20mmis 37°

Time	Run	Substance	Code	Value 1	Value 2	Value 3	Value 4
399 1:200	1	/	020	478	462	1/4	21.6
	2A		018	421	406		24.6
	3		—	319	319		31.3
	4		-5	203	208		48.1
506 1:150	1	Megal	019	421	406		24.6
	2B	M/100	009	353	346		28.9
	3		-001	229	230		43.5
	4		003	200	97		103.1
506 1:150	1	M/100	020	451	435		23.0
	2C		011	400	391		25.6
	3		007	281	275		36.5
	4		-004	204	208		48.1
506 1:150	1	Megal	020	404	388		25.8
	2		010	337	329		20.5
	3	M/100	008	217	210		30.4
	4		-003	128	131		47.6
							76.3

??

Data n.g. expt. needs repetition.

Test for induction period in cellular
utilization of ONPG.

514

4/8/49.

Harvest K-12 from Y2 lac. Conc. 5x. Use .2 ml/10.
Make up in NaP buffer ± ONPG 1/2000. Run ext in cuvette.
Temperature: 22.5 initially. Add substrate at to.
24° at 16m. at 20m.

Time	D	10	2	.
-	134		30	263
to calc.		131		
0	121		60	270
20	121 (mixing).		90	271
30	137			
40	141			
60	146	12	120	282
70	148			
100	149	<u>1m</u>		
120	159	13		293
140	162			
160	167	14		307
180	170			
200	177	15		319
220	180			
240	184	16		330
270	190	17		341
300	197	18		357
330	202	19		369
360	209	20		380
90	212			24°
120	219			
150	225			
180	231			
210	239			
240	245			
270	252			
300	259			

6m.

Correction: -13.4 for dilution of cells. + 10 for substrate. ∴ -3.4 + 134 gives initial.
= 131.

April 10, 1949

W349 is listed as lactitol + B-4-. Inoculate 2 x 500 ml
12 lactose and aerate 24 hours. Harvest + dry over 205.
Yield: 672 mg.

Lumina delutem respase
galactosidase

4/14/49

399A, ca. 1:100	1 ml/tube	NaP 4/50 7.5	ONPG 4/2000.
Di	Df	V _{cor}	
.1 003	41	25	
.2 004	72	56	
.3 0	96	80	
.4 0	127	111	
.5 003	158	142	
.6 003	189	173	
.7 002	212	196	
.8 010	267	242	} non lumina
.9 001	318	302	
1.0 001	354	338	
0 Substrate	016		

Substrate (~~1/100~~) 016 from all sec.

and Di from #8. - .025 on this one.
for V_{cor}

4/29/49

Grow K-12 shebeen overnight in $M/100$ hba Sml. Harvest and compare with lac $M/100$ adapted cells, etc.

		7:10PM Di	2:00
1	lac	119	800
2	hba	106	126
3 (↗)	glu(42)	157	172
4 (↘)	NSB	162	170

These tubes were made up from Stodola's purified lactobromate. Either the pure prep. is inactive or $M/100$ is too dilute.

App. increases of 2-4 probably artifacts; no visible color. ~~most yellow~~
 hba does not adapt to ONPG. after 1 hour, progressive color ~~after 1 hour~~
 in controls, ca 50. probably adaptation.

Effect of azide on pH sensitivity.

Compare activity of lac adapted cells above in $KPM/50$ buffer pH 5.0 and 7.0 all tubes receive $M/50$ Na_2SO_4 and $M/2000$ ONPG.

pH	Azide	Di	Df.	R.A.
7.0	-	54	340	
7.0	+	60	361	
5.0	-	53	94	
5.0	+	51	145	

SIC!

Azide stimulates cells!

should use KNO_3 to eliminate Na effect

April 17, 1949.

Prepare M/10 Na Lba. from Link's crude material with Sod. Carbonate equiv. to pH 9.5

1. Make up YZ- M/40 Lba. Grow K-12 and Y-53 in 5 ml. ea. overnight with shaking. From these suspensions, inoculate 180 ml aerated flask for dry-cell prep'n.

Test galactosidase activity of washed suspensions. ONPG M/2000; NaP M/50 7.5
20 mins. 37 C.

	D_i	D_f	R.A.
K-12	229	880	300
Y-53	229	222	008

This prep. of lactobionate certainly elicits a very active galactosidase, but not from Lac₁-/

The cells harvested fermented glucose, lactose very very slowly.

2. Inhibitions. Make up tubes with .01 ml 399A lactase, M/1000 ONPG, NaP as above.

To 3,4 add M/100 Lba.

	D_i	D_f	
1	002	251	250
2	0	252	
3	010	169	
4	014	164	155

$\frac{1}{v_0}$ $\frac{1}{v_i}$
40 ↙
64.5

Taking K_{onpg} as 1.3×10^{-4} , K_{lba} can be calculated:

$$K_I = \frac{I}{I} \left[\frac{\frac{1}{v_0} \left(1 - \frac{S}{K_s + S} \right)}{\frac{1}{v_i} - \frac{1}{v_0}} \right]$$

$$= \frac{M}{100} \left[\frac{40}{24.5} \left(1 - \frac{10^{-4}}{2.3 \times 10^{-4}} \right) \right]$$

$$= \frac{M}{100} \left[\frac{40}{24.5} \left(1 - \frac{1}{2.3} \right) \right]$$

$$= \frac{(40)(.57)}{24.5} \times 10^{-2} = .93 \times 10^{-2}$$

$$\frac{K_I}{I} = k_i = \frac{v_i k_s}{(v_0 - v_i)(1 + k_s)}$$

$$= \frac{155}{95} \frac{1.3}{2.3}$$

$$= .83$$

$$= 8.3 \times 10^{-3}$$

(crude lactobionate)

3. Inoculate dried cells from 180 ml aerated YZ-Lba. Yield 160mg. Well aerated culture was very dense

Lactobionate.

523a.

4/17/49.

399A 15/100

M/1000 ONPG

Repeat \bar{c} purified lactobionate from F. Stodola. NaPM/50, H7.5

<u>Lba.</u>	<u>D_i</u>	<u>D_f</u>
-	003	400
M/200	004	367
M/100.	010	359

ONPG added. Concentri. 010

$$\frac{K_I}{I} = \frac{V_i k_s}{(v_0 - v_i)(1 + k_s)} = \frac{359 \times 1.3}{(41)(2.3)} = \frac{367}{23} \cdot \frac{1.3}{2.3} \cdot M/200$$

$$= 4.9 \times 10^{-2}$$

$$4.5 \times 10^{-2}$$

$$\text{use } \bar{m} = 4.7 \times 10^{-2}$$

523a

Concentration effects on adaptation

4/20/49.

Lactose 3.6% stock. Make up $\frac{1}{2}$:		2 each.		
	$\frac{1}{10}$	$\frac{1}{10}$	Δ	R.A.
1. M/50	041	432	400	1000
2. M/100	044	570	530	>1000
3. M/500	056	395	350	650
4. M/1000	053	477	430	900
5. M/10,000	045	120	4 80	170
6. M/100,000	048	77	35	75

Harvest K-12 grown overnight in $\frac{1}{2}$ + each of above conc. (10 ml shake flask). Conc. ca 5%; use 1 ml / 10 ml tube in assaying for galactosidase.

Repeat adaptation to galactose (1%) and Lba purified (M/40 in $\frac{1}{2}$)

Gal	087	139	60	75
Lba	063	97	40	55

The cut off of adaptive response appears to be much lower than for combination of the enzyme!

The response to lactobionate is undoubtedly due to lactose impurity. If M/40 lactobionate is used, an impurity of 1% will give M/4000, in the range of effective response!
 = Check if Lba potentiates adaptation!