

Inhibition of galactosidase by carbohydrates.

Sept. 15, 1948

Galactosidase from *E. coli*

M/50 citrate buffer
7.5
20m. 37°
M/500 ONPG
M/10 (ca.) Sugar

Strain	Control	% inhibition	Sugar
K-12	0	0	Control
	Maltose	- 80	Maltose
	Galactose	- 83	Galactose
	Glucuronate	+ 15	Glucuronate
	L-Trehalose	76	L-Trehalose
	L-Fructose	85	L-Fructose
	D-Xylose	55	D-Xylose
W 211	0	-	
	Maltose	87	Maltose
W 33	0	-	
	Maltose	82	Maltose
Y 10	0	-	
	Maltose	85	Maltose
PM	0	-	
	Glucose	79	Glucose
	Fructose	53	Fructose
	Mannose	11	Mannose
	Raffinose	100	Raffinose
	Trehalose	5	Trehalose
	Dulcitol	79	Dulcitol
	Sorbitol	29	Sorbitol
Melibiose	100	Melibiose	

Sept. 15, 1948.

Due to paucity of material, the following tests were done in 1.0 ml volumes. $100\mu\text{M}$ was dissolved in .9 ml bacterial suspension in buffer as above, then .1 ml $1/500$ ONPG was added after temp. equilibrium. Color read as + or - :

H-12

	Color	
Maltose	-	
Galactosan	+	
Lactitol	-	\pm
d-xylulose	+	
Ca lactobionate	+	
Ca maltobionate	\pm	Original color makes this reading doubtful

Adaptation Expt: Preliminary

3502
4 13
1 19

Sept 17, 1948

		$\lambda 420$ 1000	$\lambda 650_{00}$	$\lambda 650$ vs water	Y20 H ₂ O	3 hour exposure opt. color
L	-					
glucose	1	.000	.018			+
"	2	.100	.017			+
lactose	3	.012	.014			+
"	4	.215	.019			+
Lac + Azide M/100	5	.028	7.000.0			+
"	6	.012	.017			+
Blu + Az	7	.040	7.012			+
"	8	.100	7.010			+
water	9	.187	7.010			+
G	-					
glu	1	.069	.047	1159	2/6	-
"	2	.046	.047			-
lac	3	.0614	.048			+
"	4	.133	.047			+
lac azide	5	.100	.047			+
lac ATP 5mg	6	.035	.038			+
" " tyrid	7	.035	.042			+
2ml M/1000	8	.044	.043			+
SMT M	9	.017	7.010			-
water	10	.11.1	7.010			-

concentrate cells from Y2 Lac (L) and Y2 Blu (G) 5:1

Adaptation system: 2ml cells: 2ml 4% sugar in M/5 buffer + 1ml (supplement if any). Centrifuge once + resuspend in 4ml H₂O Test ϵ ONPG in M/10 citrate buffer as above, 1ml: 9 ONPG + buffer.

SMT, Azide apparently inhibit adaptation, benzimidazole does not at this concentration.

Conditions for opt. adaptation

9/18/48.

Core 100 ml H-12 from Y2/Plu to 20ml (5:1)
 Add 7ml cells to 7ml sugars 4% in M/5 buffer pH 7.0. Add H₂O or
 suppl. to 5ml volume 1130 A18. Incubate 5 shaking at 37°.

- | | | | |
|----|---------------------|-------------|--|
| 1. | Sugar | Suppl. | |
| 2. | " | " | |
| 3. | " | Peptone 1% | |
| 4. | " | V. Extr. 1% | |
| 5. | " | Glucose 1mg | |
| 6. | Glucose + Galactose | 1/2 | |
| 7. | — | — | |

ONPG as above, but use total volume of 9 ml rather than 10, and use 8/9 ONPG perously.
 Read tubes a) against water suspensions of same cells, and b) the latter against water, all at 420.

8	Lactose Hydrol. Casein 1%		
	a (activity)	b (cell dens.)	R.A. % L.
1	.160	.207	.77 100
3	.499	.334 .279	1.79 233
4	.551	.279 .334 .310	1.78 231
5	.022	.200	.11 14
6	T=101	.230	0 0
7	.000	.200	0 0
8	.519	.334	1.55 202

V. Extr., Peptone + H.C are definitely stimulatory to adaptation.

<.005

Sept. 20, 1948

System as above (except 2x for anaerobic expts.)
 All fishes contain lactose etc. 1.

	Suppl.	Relative activity
✓ 1.	-	03
✓ 2.	lac	35
✓ 3.	" Glucose 1mg	07
✓ 4.	" (W.H.P.) ₂ Soy 1ml 10%	32
✓ 5.	" " ✓, glucose	11
✓ 6.	" copracazine	43
✓ 7.	" TL	75
✓ 8.	" 4, anaerobic	28
✓ 9.	" 5, anaerobic	21
✓ 10.	" 4, Vits. ✓ 1ml	33
✓ 11.	" 2, Am. Ac.	125
✓ 12.	" M.C.	120

	D_{420}^i	D_{420}^+	$D^i \times \frac{230}{251}$	Δ	$\frac{\Delta}{D_{420}^i}$
1	251	237	230	007	03
1 \checkmark	230				
2	229	282	209	073	35
3	236	231	215	016	07
4	220	265	201	064	32
5	221	225	202	023	11
6	176	230	161	069	43
7	220	351	201	150	75
8	180	210	164	046	28
9	215	238	196	042	21
10	213	260	195	065	33
11	297	610	271	339	125
12	309	620	282	338	120

Sept. 11, 1948.

Effects of amino acids on adaptation.

K-12 harvested from Y2 Glu as above.

Irradiated supplements ca 1 mg ea. in 1 ml.

	A	B	C	D.	coefficient
O	242	224			-
Lac	230	218			-
Megal	246	231			+++
Buylal	319	310			+++
CNPS	240	219			-

A = Duro tract, suage, V = 9mC
 B = Duro + substrate suage, 10mC
 C = 4mC (K. 90)
 D = B - C = Δ
 E = D/C = relative activity

	A	B	C	D	E	% var	% inter action	
A12	1	274	277	205	95	47	-	69
A3	2	246	370	216	104	48	-	71
A4	3	249	335	224	109	49	-	72
A5	4	273	429	246	173	70	+	103
A6	5	249	380	224	156	70	+	103
Arginine	6	239	291	215	76	35	-	52
Methionine	7	263	400	237	163	69	+	103
Adenic	8	243	356	232	124	53	-	79
Galan	9	230	348	207	141	68	+	102
prol	10	155	371	232	139	60	-	90
lys	11	246	366	222	144	65	+	97
arg	12	235	409	214	195	89	+	133
meth	13	231	383	208	175	84	+	125
thc	14	231	377	226	151	67	-	100
-Lac	15	230	217	207	10	5	-	7.5
H.C.	16	351	870	326	584	176	++	263
H.C. + Typ.	17	347	860	312	548	178	++	266
T+Linc	18	263	409	237	182	73	+	109.

only arginine and methionine showed significant stimulatory effect for K-12 adaptation.

Sept. 22, 1948.

5ml system for adaptations above. All Lac. K-12.

	A	B	C=A _{con}	D(p-c)	E $\frac{B-C}{C}$	% of Lac(i)
1. -	228	305	205	100	56	100
2. HC	310	700	279	421	151	
3. AA of HC	296	650	266	384	144	
4. Σ HA	271	520	244	276	118	
5. AA - A12	229	309	206	103	50	
6. " A3	249	410	222	257	116	
7. " A4	259	520	333	187	80	
8. " A5	241	477	217	212	98	
9. " A6	250	460	225	235	104	
10. Arg+Meth.	239	371	215	156	72	130

.2ml each AA group in 4-9.

.5ml ca. 10.

.1ml HC 10% 2.

1ml $\frac{10}{100}$ HC 3.

Sept 25-26, 1948.

K-12 grown 24 hours in Synthetic + Lactose 1%, 10 tubes.

25g. cell paste recovered. ca 24g. + 10pc 7.5 P₀₄ buffer shaken 24h. under toluene. Remove debris & collect supernatant in ca 30cc buffer. Deep yellowgreen fluorescence. ca 1ml/gram bacteria.

(A).
(B). ca 1g. washed in acetone and dried at room temperature. Considerable loss by spattering yellow coloration only of final product.

See 316

see 325 for assay.

Sept. 25, 1948.

K-17 grown in 200ml 1/2 lactose. Harvest to
5cc. 7.5% buffer & autolyze under vacuum & shaking
24h & 48h.

- (A) 24h. 1ml withdrawn, debris sedimented & supernatant diluted to 4ml.
- (B) 48h. Remainder (4/5) removed, etc. dilute to 16ml

Each ml corresponds to 10ml original culture & should have
an activity of ca. 10x bacterial suspension. (i.e. .05ml should give
ca 100% hydrolysis of 10ml 1/5000 ONPG in 20 mins). I.E., calculating
2g/liter, corresponds to 20 mg/ml

See 3/6

Sept. 27, 1948.

(A) K-12 grown 36 hours in 10 liters S(Lac). 9.4 liters of supernatant were removed leaving 31 grams wet Sharples paste. Make up to ca 45 ml \bar{c} PO_4 buffer pH 7.5 and grind 75 minutes in Booth-Drum mill. Combine efflux \bar{c} washings. ~~Inorganic~~ milky opalescent supernatant is obtained, in ca. 100 ml, i.e. 31 grams/ml.

(B) 10 ml sample of culture was taken. Resuspended in eg. H_2O + measure turbidity at 1:20 D_{420} .
1:50 dilutions.

1 Unit = A of .100 in D_{420} .
for cell free prep.

Assays:	A	B	C	D	Act./ml.	
1	008	290		283	14315A	.2 ml
2	002	205		205	10 10 B	.2 ml
3	007	260		254	25 314A	.1 ml
4	001	043		042	40	.01 ml
5	010	020		021	90	.001 ml
6	032	1500		1500	150+ 316A	.1 ml
7	002	980		980	980	.01
8	000	290		290	2,900	.001
9	360	1900		1600	(445) 316B(cells)	1 ml
10	079	880.		809	(1100.)	.2 ml
ONPG.	012	012.		0	0.	

v. high activity!
for non-enzymatic (non) hydrolysis!

In prep. 316, 1 ml being \approx culture medium 10 liters/100 \approx 100 ml cells.

and .001 ml should be equivalent to .1 ml cells, which it is, very nearly. (Therefore a large proportion of the cellular activity is present in extracts. Hydrolyses are nearly as effective with smaller volumes.)

Sept 28, 1948.

K-12 grown on 100 ml T(0) glucose ^A & do. + H.C. (^B 1/2 ml/100)
 shaken 16 hours. Adjust densities:

- (A) 1:10 dilution D_{650} 259
 (B) H.C. 319

ratio of 1:23 : 1.

Supplement the ~~main~~ (0) culture ²⁰ with 5 ml H₂O; the HC culture in 24.6 ~~ml~~ ml H₂O to adjust initial densities.

The adaptation system consists of 1 ml cells + 3 ml T(0) lactose + 1 ml complement. Adapt 3 hours, in duplicate. Resuspend in 4 ml H₂O + 8 ml buffer for A. Add 1 ml ONPG solution for B.

	A	B	C	D	E
1. Lactose buffer pH 7.0 M/50.	.201	745			
	.196	641			
2. T(0) lac	.248	669			
12	.260	720			
3. T(0) lac + 1 ml H.C.	.256	710			
13	.260	731			
(B) 4. Lac buffer	.177	169			
14	.184	175			
5. T(0) lac	.187	171			
15	.188	170			
6. T(0) lac + 1 ml HC	.189	153			
15	.172	168			

	A.	B.
A. 25/9	.232	.219
B. 25/9	.239	.200

Negligible activity of unadapted culture and of B series.

Sept 28, 1948.

(N2) W478, W583 on Lac B₁.

20 colonies *stuartii* on
LacEMB: All++.

Fractionation of galase 31B.

319

Sept. 28-9, 1948.

Original extract (316) consisted of 2900 u/ml in 100cc or 2.9×10^5 units all together.

To fractionate remove 50ml and dilute in 50ml H_2O . (1.5×10^5 units; ~~1500~~ 1500/ml).

"316" is fraction 0. Add Na_2SO_4 in 4 aliquots of 17.5g. each in ice bath to give $1/4$ sat'd fractions. Take up sediments in 10ml $4/50 PO_4$ ^{app. activity.} except for the final fraction.

0	hop. fract.	Act.	Prop. Act.	Assay	.01	.001
	1.00.		1.00.		615	1089

1 ($1/4$ sat.)	5.00			129	019
2 ($1/2$ sat.)	5.00			390	055
3 ($3/4$ sat.)	5.00			194	023
4 (sat.)	10.00			101	015
5 Supernat.	1.00.			060	015.
					<u>.140</u>

Assay at the equivalent of .01 and .001 ml of the ~~original~~ fraction 0. 1ml $4/50$ ONPG in $4/50 PO_4$ buffer.

Enzyme activity is probably not quite linear. Fractions have higher total activity than the original "soup".

Pool fractions ~~1, 2~~, 2 + 3 (40ml) and add Na_2SO_4 AS ($3/4$ sat.). Take up ppt in $4/50$ citrate buffer, 70-80 319A

→ P30. To remaining 50ml (1.5×10^5 units) add 250ml collacetone, let stand, and filter off 330mg. dry powder. 319B. This should have an activity of about 500 u/mg. Take up 10mg in 10ml phosphate buffer.

Effect of phosphate on lactase

Sept. 29

Lactase preparation 319A is suspended in 1/50 citrate buffer.
pH 7.5 (Ethylene diamine - citric acid) = (EDC buffer), and should
have a potency ca. $[100/20] \times (.58 + .23) \times 10^3$ u/ml. = 4000 u/ml.

Assay

.001 ml in citrate and in phosphate buffer 1/50. pH 7.5.

Triplicate series.

Phosphate seemed to be
mild. After 7 mins, use

7ml EDC + 1ml Phosph. + 1ml
enzyme + 1ml ONPG.

1	EDC PO ₄	.371
2	"	.369
3	"	.390

11	PO ₄ EDC	0.12.
12	"	0.13
13	"	0.12.

ONPM/5000m

21	EDC	640
22	"	640
31	PO ₄	750
32	"	745

41. (7 mins later).
EDC + PO₄. 0

may be due to inhibition by citrate.

Sept 30, 1948.

K-12 in A) T(0)
shake overnight.
1:100 dilution:

Days
A 119
B 119
C 052
050

5ml 1mg/ml. 5ml 1% H₂C=

1) T(Prol) C) T(AA) 2ml
Resuspend in 5ml H₂O. Turbidity at

dilute A and B to 11.9 ml to equalize c.

Adaptation system: 5ml. 3 hours 37°. 10³⁰A - 1²⁰P

A. B.

1ml cells
3ml substrate.

① Phosph M, 50 1.5 + 2% tae

② T. (2%) Lec.

③ ② + supplement prolinc 1mg% 2ml

④ ② + H₂AA. 1% 1ml

A	1	176	220
	2	259	331
	3	162	218
	4	160	291

B	1	169	215
	2	167	206
	3	186	226
	4	174	272

C	1	150	281
	2	190	310
	3	226	589
	4	249	778

T(0) cells did not adapt!! T(AA) cells were stimulated by T(0).
& further by amino acids.

4/7/70

A B C = .9A D = B - C E = D/C % Lac - Suppl.

	A	B	C = .9A	D = B - C	E = D/C	% Lac - Suppl.
1	257	368	231	137	59	120
2	248	329	223	106	48	98
3	241	335	217	118	54	110
4	242	329	218	111	51	104
5	241	300	217	83	38	77
6	259	402	233	169	73	149
7	242	366	218	152	70	143
8	241	367	217	150	69	132
9	249	372	224	148	66	135
10	250	427	225	202	91	110
11	240	410	216	194	82	153
12	240	451	216	235	109	174
13	269	489	242	247	102	208
14	211	452	190	262	138	173
15	264	441	238	203	86	167
16	230	352	207	145	70	—
17	319	715	287	428	149	49
18				508	177	362

Σ AA
 AA-A12 + arg
 " lys
 " meth
 " cyst
 AA - arg.
 - lys
 - meth
 - cyst
 AA-A4 + dal
 + tyr
 + hyp
 AA - dal
 - tyr
 - hyp
 O
 H.C.

INHIBITORY!

dal inhib? hyp stimulatory.

Activation of Lactase.

Sept. 30, 1948.

EDC

A. Phosphate vs. citrate. System is, as usual, 10 ml and 1/2000 in ONPG.
.001 ml of Lactase 319A used for test.

- | | |
|----------------------|-----|
| 1. 1ml 1/5 Phosphate | 222 |
| 2. 1ml 1/5 Citrate | 021 |
| 3. 1ml each. | 022 |

All contain 1ml Phosphate Buffer

- B.
- | | |
|------------------------|------|
| 1. Add — | 189 |
| 2. 1ml EDC | 012 |
| 3. 1ml Na citrate 1/5. | 190. |

The inhibition is clearly due to the ethylene diamine component of the EDC buffer!

Oct. 1. Test .002 ml of 319A in the following buffers, each at 1/50 pH 7.5

- | | |
|----------------------|--------------|
| 1. Phosphate | D420.
310 |
| 2. Glycero-phosphate | 488 |
| 3. " + Phosph. | 477 |
| 4. Barbitol | 513 |
| 5. " + " | 494 |

Deficiency in phosphate was visibly apparent. A NaCl effect?

Phosphate is not required for the reaction.

ONPM/5000 in: 1/50

- | | |
|----------------------|------|
| 1. Phosphate | 694 |
| 2. Barbitol | 645. |
| 3. Glycero-phosphate | 725 |

Activation of lactase + other assays 32/9.

To test influence of NaCl add 1ml of M/5 NaCl, HCl, and Na_2SO_4 respectively to a phosphate buffer system as above. 319A .002ml
 Phosphate M/50+:

1. — 275
2. NaCl 395
3. HCl 259
4. Na_2SO_4 514.

M/50. Repeat

1. ~~NaCl~~ 317
2. NaCl 512
3. Na_2SO_4 592
4. HCl 298
5. LiCl 218
6. NH_4Cl 230
7. $(\text{NH}_4)_2\text{SO}_4$ 252
8. MgSO_4 257

Inhibitory.

NaCl concentration series:

1. — 318 410
- M/50x 2. .1 405
3. .5 388
4. 1.0
5. 5.0

↓
 inhibitory

Sept 30, 1948.

17g. wet paste K-12 harvested from 20 ^{liters} ~~gallons~~ (low yield!)
S(Lac)

Add ca 50cc cold acetone to dehydrate, filter, and desiccate
the residue. Assay sample of cells for activity.

Dyno. A. B. Also, other assays:

325

314B.	1mg	134	1150
	.1mg	022	379
	.01mg	012	046

ca. 35 u/mg.

319B.	1mg	68	1070
	.1mg	51	960
	.01mg	17	193

ca. 190 u/mg.

→ 3.2 grams dry powder obtained: Lactase 325A.