

New heterozygotes

376

December 4, 1948.

A. w65 x w595

No yield

B. w48 x w595

C. w182 x w595

12/17/48.

~~By~~ W45 x W595 on Lac EMS.

12/8. No yield! (3 + colonies in 15 plates!)
2 not col. 3d +.

Note. AT6.A12 streaks out W-1 to W595 series to establish
mutability.

on lac

Y53	lac -	M (regularly; many colo. stable).
W1	"Gal -	M consistently.
W566	"Gal -	S
W582	"Gal -	SS
W583	"Gal -	S
W595	"Gal -	S

The mutation to Gal - seems to have been accompanied by stability of
lac - , possibly fortuitous.

12/28/48. Test other Gal - mutants of this series on Lac EMS for
mutability:

W 565	Stable, thin colonies	575 mostly small stable colonies; some large unstable.
W 566	" heaped up centres.	
567	" (very occasional papillae).	
568.	Stable.	
569	r. on colonies; some revert to typical unstable.	576 small colonies uniformly.
570.		
571	like 565	
572	like 567	
573	stable, large colonies	
574	typical unstable	

12/6/48

A. W126B x Y87B Dec 373.

B. W495 x W45

C. " W48

D. " W65

E. " W182

Yields low:

A. 5 plates 100/plate. 3+ colonies. S.O. on LacEMB + EMS. ++

B. 10 plates > 2/plate 2+ colonies. ++

C. " ca 1/plate No +

D. " ca 1/plate No +

E. 9 " " No +

12/5/58.

Rich 1 - colony from each of four mosaics of H119 - H122 & test as indicated.

	Lac	U ₁	Budal	Nut.	Summary:	Bug	V
119	A - S B - M C - S D - M?	S R S S	- + - ±	TM TM TL T	* ✓ -	3 1 2 5	+ R + S - R - S
120	A - M B - M C - M D - M	R R R R	+	M MT M T			
121	A - M B - S C - M D - S	R S R S	+	MT (MTL) Y HM * ✓ - MTL	* ✓ -		
122	A - S B - S C - M D - S	R S R R	- - +	MT ✓ TL ✓ MT ✓ MT ✓			
temp. data	Y87 W126	- M - S	R S	+	BM TLB,		
		6S:10R					

Note preponderance of T- and M- streaks over indefinite Budal streaks. *

There is a general correlation between mutability and Budal - but it is not perfect here.

Maintenance of heterozygotes.

380.

12/17/48. H1 ^o ✓✓

Lac 22 All+ All+ Return to previous EMS plates.

Lac 52 ✓ ✓✓

Lac 62 ✓ ✓

Lac 72 ✓✓ ✓?

Xyl 85 ✓ ✓

Xyl 93 ✓✓ ✓✓

Lac 118 - -

+ colonies from previous EMS plates restreaked as EMS. These streaked, 2 / type, on EMB and streaked as EMS; also on Nutrient agar slant (subculture 1).

A6. Stake out NA slant from H1 and H118 to determine feasibility of recovery at this stage.

4 tests each.

H1. 1-3 Var. 4+ or Var?

H118. All 4 are Var.

This may be a suitable method

Dec. 13-14, 1948.

- A. W45 x W595
 B. W45 x W583
 C. 58-161 x W595.

} EMS Lac

①. 15 plates each P13. A16: all but blank.

A:	7 colonies altogether.	5 possibly +.	All --	No Var.
B:	17 "	10? "	+. All ++ or --	
C.	-	+	4 tested: 3++ 1-	

Pick all possibly + colonies and streak out on EMIBloc, ~~or agar~~ sugar + EMS.

②. 15 plates A + B A14. low yields, but pick up apparent +'s. (28)

Mostly - mostly scored as +. No vacuolated.

Some weak(?) + noted. Specify on Lac. EM B

19, 2, 6, 14, 16. keep as W-460.

16 is Gal++ Lac++ others are Gal- Lac very weak

Papillae seen on lac + Gal plates. Streak out on both media.

Gal + pop:	Lac	Gal
6	++	++
14	++	++
Lac + from 6		
o	++	++
1	++	++

Note: on lactose, residual Gal - colonies show near + reaction when they are situated in vicinity of + colonies.

Conclusion: The Gal- in these stocks is also an inhibitor of lactose fermentation, in distinction to W-255. H93, therefore, may now be lac+ and Gal- . It is not proven that lac- can be homozygous!

12/23/48.

Cf. W460 on 1% and 3% LacEMB.

At 48 hrs. W460 is nearly ++ on 3% lactose
still slow on 1% ".

Break out W595 on EMB galactose for reversion.

Test revert on lactose for mutability.
(W₆₆₀) #4. All are lac mutable like Y53.

heterozygotes

391

Dec. 18, 1948.

Cross W45 x W595 on Lac synthetic media.

A) "EMA" .5% asparagine as C source.

(B) EMS, fresh batch. No succe "

(C) EMA+B. Asparagine + Succinate .5% each.

(D) Like B. But standard.

Very heavy
(4x conc.)
mucula.

1-8. 8+ / 11 plates. A few lac-. Pick + test +

9-15 7+? / 13 plates. Survival -.

c) 12 plates. Poorly scored, but yields much higher.

16-46 25+.

D) 4 plates. 1? +.

42

Very few scored + on EMB. Some were lac unstable. (W45?)

6++ altogether. Numerous slow + à la 389

Test media processes.

December 22, 1948.

Cross W478 x W595 on
various media using constant
moisture. (1 dyes 1/2 del. parents)

Also conc. moisture as Lac Eqs.

+ B₁: → 1-6.

5 plates each.

					100cc
Mesum	.59				
Hypoxor	.59	red			
Null	.19				
4 mg salt	0.019				
Agar	1.59	metabolic			
K ₂ HPO ₄	.2				
Lac	19				
8,					
Sucrose	0.04				
met. base	0.0065				

12/24:

T(B₁). 51, 43 49 46 43 m = 46.4

T(0) 4 3 5 10 1 m = 6.6

(1) EMB.	1	0	0	0	0	.2
(2) MB	0	0	0	0	0	0
(3) E	0	0	4	1	0	1.0
(4) No dye.	10	8	14	4	10	9.2

The dyes are certainly inhibitory, but the minimal medium
base is certainly not very satisfactory, possibly due to use of lactose
as main carbon source.

Finn 20 states mainly sucrose, about 3% + lactose
& 1% agar using sterilizing 10% formalin. 100cc mixture
of a suitable Lac +.

3978 x 3979

3934.

12/29 - 1/48.

1. 26 ✓
2. 46 ✓
3. 171 ✓
4. 188 ✓

not heterozygous.

: 139, 140, 141.

	bac	xyl	Mannitol	Gal	Neb	17al
139:	±	±	-	++		
140	±	±	±	++		
141	±	+	-	++		

12/23/48.

Recover H93 from nutrient agar slant and from ~~this~~^{Xyl.} plates from NA to Xyl EMB. Prod. Xyl-. Ca 2% mosaic colonies.

Nutrient agar probably remains a preferred means of maintaining heterozygotes.

Similarly on EMS Xyl. Pick a few to Xyl EMB to test recovery of H-93.

From EMS plate 7½ are still mosaic. Recover bacteria from ~~to~~ EMB; EMS Xyl.

Where a heterozygous colony is streaked out on

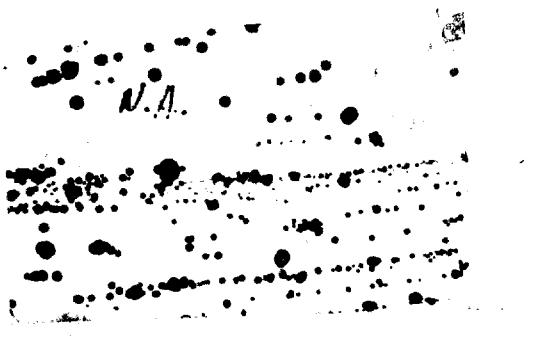
EMB:

Gal negative

Xyl almost all mosaic colonies

Lac slow + (or two colonies finally ++).

Lac 3% full +; no signs of variegation.



H93 is therefore probably Xyl+/- Lac++/+ Gal-/-

December 24, 1948.

S(lac) 7 carbays eq 30 hours

1/2 ml .251 .340.

∴ Each ml of culture will
provide an equivalent of 1.01 ml.
of 31% H.

104 g. collected from two carbays (20 liters). i.e. equivalent to
20 ml 31% H.

58 g. suspended in a very heavy cream in the P 7/80 for grinding
but ~~the~~ pump did not draw properly. Retain cream & running
paste.

12/25/48. Recardetion mill + grind remainder of cells. An acetamin
basis. (ca 40-50 g. paste probable).

Ca. 10 ml of extract. Assays 2970 u/ml.

Galactose mutation run
X-glycose.

396

12/24/48.

Y87 7secs. etc.
Galactose

80 plates ca 200/plate
16,000

W870 7secs. etc

33 plates. ca 300/plate

10,000

→ W641.
642 :
643 very thin.

Xylose.

W Galactose:

644	1	-
45	2	- thin
56	3	slow +
47	4	slow ++
48	5	- small col.
49	6	slow ++
50	7	slow +
51	8	-

lac

M
S
M
S
M
M
S

use 644 for

further studies.

52	9	slow +
53	10	slow +
54	11	slow ++
55	12	slow ++
56	13	slow +
57	14	- thin,
58	15	- thin,
59	16.	- thin,

M
M
S
M
M

pop. to have viscosity, plate. salt +
S.

Assimilation by various batches of
MB; Eosin. - Crosses.

398.

12 | 19 | 5

Weighs out 40 mg Eosin Y and 6.5 mg MB of batches indicated. (Identification numbers etc.)

	Batch	Time	2	3	4	5	6	7	Average
1.	23	27	59	78					
2.	23	28 ✓	113	38					
3.	23.	29	93	25					
4	24 23	11	56	8					
5	24 23	28 ✓	103	23					
6.	24 23	29.	49	49	8.				
7.	22±	28	2368	13					
8.	24 23	28 ✓	473	866					
8.	7/0).		146						

all batches gave results comparable to 7/0).