

28

A. Y40 x Y53.

T(0).	-R	-S	+R	+S.	
1.	7	7	6	0	
2.	3	1	4	0	
Σ	10	8	10	0	28

T(B.)					
1.	2	3	4	0	
2.	4	3	4	0	
3.	11	2	6	0	
4.	9	4	7	0	
5.	10	5	2	1	
7.	15	6	9	1	
8.	7	5	7	1	
9.	5	7	7	0	
10.	2	4	1	1	
11.	8	7	3	0	
Σ	73	46	50	4	173.

Expressed as percentages.

	-R	-S	+R	+S.	
A(0).	35	30	35	0	28
A(B.)	42.3	26.6	28.8	2.3	173
B(0)	31.9	47.4	4.3	16.4	116
B(B.)	34.9	39.0	2.5	22.4	312

B. Y64 x 58-161.

T(0).					
1.	4	4	0	0	
2.	2	3	1	1	
3.	2	5	0	2	
4.	1	4	1	0	
5.	6	9	2	2	
6.	5	9	0	4	
7.	3	5	0	2	
8.	2	4	0	2	
9.	5	8	1	3	
10.	7	4	0	3	
Σ	37	55	5	19	116.

T(B.)					
Σ	109	125	8	70	312.
	see next page.				

See summaries of data

Y₀Y x 58 - 161 B

484a

Y40 x Y53 A

April 20, 1947

A :

-R	-S	+R	+S.	
7	9	3	0	
4	8	3	1	
7	5	4	2	
20	22	10	3	55

In previous expts,
+R >> -S occasionally.

A(B₁)

9	4	4	1	18
8	4	4	0	16
8	8	4	0	20
11	4	3 4	1	19
7	4	8	1	20
16	10	8	0	34
59	34	31	3	127

B :

8	9	1	2	20
---	---	---	---	----

B(B₁)

6	7	0	6	
4	9	1	2	
10	16	1	8	
10	5	0	1	
20	21	1	9	71
28		2	11	51 B
				A
8	1	1	2	A(B ₁)

1192
22
1214

484 ctd. Y64 x 58-161. T(B₁)

	-R	-S	+R	+S.
1.	7	4	1	7
2a	5	7	1	7
b.	8	10	1	4
c.	9	5	0	4
3a	4	14	1	1
b.	6	7	1	5
4a.	5	12	0	3
b.	6	8	1	4
5a	8	10	0	2
b.				
6.	11	6	6	17
7b	10	6	0	0
7a	5	8	0	6
8a	9	5	1	5
8b.	2	10	0	7
9a	7	10	0	2
b.	10	3	0	5
10a	10 9	12	0	3
b.	7	4	1	7
Σ	109	125	8	70. 312

numerals designate separate recombinant plates. letters are testing plates.

} Retest - phase n.g.?? 13-R: 6+R.
 } appearance very poor!

N. tetrasperma

A. Pr 16 / sub. & B. Sub

Spread spores (or pinthecia) on agar surface.

2/26/47. Irradiate spores \bar{c} ca 20,000 r (courtesy of Pollard)

Isolated 2/27 Transfer spores to small corn meal agar slants & heat-activate.

Do \bar{c} unirradiated controls.

Fruit scoring -

Substrate	# plates	# spores	# plates	# spores	Score	# plates	%
Control	100	80	37	16	53	26	85%
X-ray	100	69	9	7	16	51	= 76%

Substrate	J	E	J	E	J	E	J	E
Sub 16	47	21	34	7	69	25	23	4
X-ray	50	14	14	1	14	1	13	6
	77	12	35	2	41	3	13	6

D	d
10	4

+ 3 scumbo

Sanitation: transfer sterile to 100 - C 44 C → 28 X → 51

C → 18 X → 13

31/8
Sub / 2000
12/6/47
103

April 15, 1947.

FH (2% glucose +).
HC 1/2%

noc 5 P15

	N2 case 1/2%	+++	+++
		++	++
1 - Glutamic ac.	growth 12h.	++	++
2 - Glycine		++	++
3 - Serine		+ +	++
4 - Aspartic ac.		+	+
5 - Asparagine		+	++
6 - Glutamine		++	++
7 - Proline		+	+
8 - Hydroxyproline		++	++
9 - Cysteine		++ 1/2 S. + g.	++
10 - Alanine		+	+
11 - Tyrosine		++	++
12 - O.		+	++

1 mg / 10 ea.

The production of gas in the minimal medium is at odds with previous results, and perhaps speaks for some error.

Repeat: OK. - gas produced on FH (O) by K-12 in 36-48 h.

abandon voluntarily. One could seek an ~~ant~~ antagonist of formate, however, which seems to be present in N2 case.

Stutshorn?

Phycomonas - Recombination.

485

April 1947

Series: B. A6/Pro/MG. A. A6/Str/Sun. moc separately and together
 after growth on slants, into caout medium. - 4 wks. Sediment + wash pellets.
 1: caout 2: slant. moc from caout into YB liq: 3.

on 1AB/Noch. Agar plate, "rough" colony noted.
 1. cl to recover

- A = Prolacme 100r/ml.
- B = Maleshute Sun 100r/ml.
- C = Streptomycin 50u/ml.
- D = Streptomycin 100u/ml.

Read at 48 hrs.

		1A [ⓐ]	1B [ⓑ]	1AB [ⓒ]	2A [ⓓ]	2B [ⓔ]	2AB [ⓖ]	3A	3B	3AB
11	A	-R	+	++	-	-	++			+
12	B	-	±R	±	-	-	-			++
13	C	+	++	✓	±	++	✓			++
14	D	+	++	-	±	++	-			++
5	AB	-	-	but prof.	-	-	-			++
7	AC	-	R	✓	-	-	-			many R
8	AD	-	R	-	-	-	-			±R of 1B
9	BC	-	-	IR	-	-	-			-
10	BD	-	-	-	-	-	-			-
6	CD	+	++	prof.	±	++	✓			+
1	ABC	-	-	✓	-	-	-			-
2	ABD	-	-	✓	-	-	-			-
3	ACD	-	IR	✓	-	-	-			- IR
4	BCD	-	IR	✓	-	-	-			-
15	ABCD	-	-	✓	-	-	-			-
	NA.	+	+	+						

See series ~~A+B~~ P 21; also moc on NA plates for series 3.

Streak on plates in order:

Read at 1) 12h.

2) 48h

3) 60h.

Sun and Str OK. (see CD). But 3 and 4 do not suggest recombinations

(see 5).

No evidence of recombinations. Prolac + H.T. apparently interact.

R = individual resistant colonies (1-10)

noc 3AB N24.

Use ACD < B in combinations.

Digenization of Cl^R .

486.

28/11/11

Y40 x Y88.

Compare c 181A.

A. T(O) B. T(B).

Streakout on Cl^A -minimal or Cl^A -biotin agar.

A. 1R/6S. = 1/7

sp R!

B. 1R/6S = 1/7.

2/16 = R.

middle data on digenization c B⁻
Use selection of Cl^R on B plates, c reverse cross to establish
complementation.

#14/16 Res.

23 APR 1957

(A) Plate 440 x 488 on B₁-Acetate medium. This should check for B₁⁻ ~~la⁵~~ segregants. By plating these colonies into BMTL-like medium it should be possible to eliminate the parental types (B-H-~~repla~~ sensitivity; T-L-B₁-~~key~~ B₁ requirement) and find any complementary appearance.

Growth is meth-eaten, suggesting phage! very strikingly.] See 489. 4-9. probably not.

(B) Plate 440 x 488 on B₁ medium, spreading very lightly, (to avoid contamination.) Pick 100 colonies carefully to minimal (ca 20 colonies/plate). and test: streak out original isolates of those with a B₁⁺ component to find any possible B₁⁻ types

Scoring as B₁⁻ or B₁⁺ not very clear cut. Group A. more definitely B₁⁺ Group B doubtful.

A. - 15. B. - 17. streak out on Lac plates.

- 1 all -
- 2 all +
- 3 +
- 4 -
- 5 -
- 6 + ; (-)
- 7 -
- 8 -
- 9 + ((-))
- 10 -
- 11 -
- 12 -
- 13 -
- 14 -
- 15 no cols.

10/4
all phage homogeneous unless Lac reaction different. Test these individually

- 1 100+ : 1 -
- 2 100+ : 1 -
- 3 all -
- 4 "
- 5 "
- 6 "
- 7 all +
- 8 ~~no cols~~
- 9 all -
- 10 all -
- 11 all +
- 12 all -
- 13 "
- 14 "
- 15 no colonies
- 16 all -
- 17 all -

11- / 5+

T do.

See over.

finds virus, plates 4 cols. each plate.

A) mostly +R ~~ca~~ ca^R
~~R.~~ ca^R
 1-S. TLB_i^-
 parental.

B) mostly -R $ca^R (B_i^-)$
 1+R $ca^S (B_i)$

no new combinations aside
 from main component in these colonies!

25/11/2011

A. Y40 x Y86. (to compare V^R loci)

B₁. (to reduce recombination required). B-M-Lac + $V_{IA}^R Muc^-$ x T-L-B₁-Lac - $V_{IB}^R Muc^+$

-S Sum. $V^R M^+ Lac -$ 16

$\therefore V_{IA}^R \neq V_{IB}^R$

$V^R M^+ Lac +$ 3

$V^R M^- Lac -$ 1

488-1 $-V^S M^- Lac -$ 1

This Recombination suggests that

$V_{IA}^R \neq V_{IB}^R$

488-2 $V^R M^+ - Lac +$ 1

27

$V_{IB}^R = T5 S.$

B. ~~58-161~~ x Y86. (to test complexity of $Muc^+ - V^R$). Lac + $V_{IA}^R M^-$ x Lac - $V_{IB}^R M^+$

(to reduce recombination required.)

-R Sum. $V^R M^+ Lac -$ 13

$\therefore Muc^+ \neq V_{IB}^R$

$V^R M^+ Lac +$ 3

$V^S M^- Lac -$ 1

488-3 $V^R M^- Lac -$ 1

This recombination suggests that $Muc^+ \neq V_{IB}^R$

[resistant to all phages. Contam?]
"diplococci"

Compare the resistance patterns of 488-1; 2; 3; Y40; Y86

Compare recombination values of $Mucoid / Lac$ & Standard V_{IA}^R / Lac .

From accumulated data:

45.79

Y40 x Y53. $V_{IA}^R Lac -$ 846 $V_{IA}^R Lac +$ 546. / 1392 = 60.6% . 39%.

Y40 x Y88 56 15 / 71 = 79% . 21%

$\therefore V_{Muc}^R$ is ~~not~~

Reversion of Cl^{R} .

488

24 11 1947

Plate 487 into Acetate - minimal.
.1%

Indefinite background growth. nodewite colonies.

probably
Ac too low.

490.
Allelism of V_1^R

April 23, 1947

allelism of V_1^R . 440 x 464.

all resistant.

T(0). lac - lact
4 1 (Mucoid)

159/159

T(0₁)
11 4
15 4
11 7
13 4
13 5
18 0 (!)
15 3
9 0
13 5

122 32 / 154. = 79% Lac - This distribution fits 464 x 58-161 better than 453 x 454

Compare fit to accumulated data.

a).
106 48 ✓
122 32 154
562 1784
1222 1206 578 1784
1528 610 1938

$\chi^2 = \frac{162}{9}$
= .25
2.42
5.32
1.45
8.44

for 440 x 453.

$\chi^2 = < 1$.
119 122 55 32 154
540 537 156 159 696
659 191 850

Linkage Relations of Cl_a .

491.

24 APR 1957

Plate 440 x 488 into Biotin - Acetate-agar ^{B_{1-}} [select for $Ac^+ = Cl_a^s$ and compare segregation of B^- and B_1^- in the Cl_a^s class I.

n.g. see 489

24 APR SAT

1. Cf. 490. 1 drop of Y40 x Y64 mixture in B, suppl medium, to compare \bar{c} Y90 for rate, (grow in Y10; plate in MW) 0.

2. Y40 x Y53. grow in M-W:∞ Plate do:

3. Y40 x Y53 grow in M-W Plate in T(0)

~~4. Y40 x Y53 grow in YB~~

4. Y40 x Y53 grow in YB Plate T(0).

5. do. do. Plate M-W 0.

} adjusted to ca. same inoculum.

> 200.

39; 35. - ca 40

The medium is n.g. for plating, but may be OK, or better than YB for growing cells personally. Try 5 buffer., 5 KNO₃.
 Difference is within range of normal variation 1 expt. to another.

Medium: per liter

- KNO₃ 1
- glucose 10
- NaCl 5
- K₂HPO₄ 3
- Mg₂PO₄ 1
- H₂SO₄ .1
- trace + CaCO₂

I (N2 case 5 Yeast ext 2.5) for 10.

April 26, 1947.

A. Y87 (β -M-V,^RLac-) x Y10 (T-L-B,^S-V,^SLac+). for segregation of Lac-. (prediction: +R > +S = -R >>> -S.)

B. Y87 x ~~Y87~~ Y64 (β -M-V,^RLac- x T-L-B,^R-V,^RLac-). for test of allelism.

B). 134 tests of prototrophs all lac- ∴ loci are allelic.

A) Segregation:

Plated mT(0)	-R	-S	+R	+S.	
	7	1	7	6	
	5	3	7	3	
	3	0	9	9	
	7	0	6	6	
	2	1	11	7	
	4		6	6	
Σ	28	6	46	37	117

mT(B ₁)					
	9	1	16	9	
	5	0	16	10	
	7	0	8	4	
	5	1	13	3	
	4	0	10	3	
	4	0	10	4	
	5	0	9	4	
	4	0	12	1	
	14	1	10	5	
	4	0	6	6	
	2	0	8	4	
	6	1	14	6	
	6	0	12	3	
	4	1	6	5	
	4	0	7	4	
	9	1	13	3	
	6	0	5	3	
	4	0	7	4	
	4	0	10	3	
	102	7	201	91	401.

Y91 x Y53 (B-M-Cl^R V₁^R x B₁-T-L-lac - V₁^S Cl^S)

Minimal plates too crowded.

B₁ - streak on B₁-Cl^R or ~~B₁~~ Cl^R NA. to clarify resistant/susc.

4/56 ~~resistant~~ resistant

Use plating on B₁-Cl^R Hagar for
longer times.

April 26, 1947

Struck on indicated plates.

	Cl _a	Bac ₁	Papillae
Y64	Cl _a 1	+	+++
58-161	Cl _a 1	+++	-
<u>Y90</u>		+	+++

4/27.

	Cl _a	Bac ₁	Pap.		Bac ₁	Pap.
Y64	Cl _a 2 ± ±	+ ±	± ±	Y94	Salin. 20 ±	++
58-161	Cl _a 2 ± ±	+ ±	++	Y96	Salin. 21 ±	++
Y90	Cl _a 2 ± ±	+ ±	++	Y95	Sour man.	+++
Y92	A2 50	+++	-		Phytomeres	++
Y92	A2 100.	++	-		Staph	++
Y53	A2 50	++	-			
Y53	A2 100	-	++			
Y40	Ia 25	++	-			
Y93	Ia 25	+++	-			
Y40	Ia 50	++ -	++	Y97		
Y93	Ia 50	+++	++ -			

need a buffered test medium probably.

Y95. Cl_a ++, I_a +++, Cl_a+I_a -
Interaction??

Struckout all mutants on N.A.
Test on inhibitor and transfer to plants.

Note - terminology: unless otherwise stated, figures are v/ml. Undeclined figures are mg/ml.

Virus Resistance Pattern.

498

	T1	T3	T4	T5	T6	T7.	
Y40	"R"	R	S	S	S	S	note!
K-12	"R"	R	S	S	S	S	
"V ₁ " ⁴ -lac-	488-1	"R"	R	S	S	S	
"V ₁ " ⁴ -lac+	488-2	"R"	R	S	S	S	
Z ₂	488-3	"R"	R	R	R	R	contaminated?
	Y86	"R"	R	R	R	S	
	Y65	"R"	R	-?	S	R	too light nice,
	Y68.	"R"	R	S	S	S	

This phage ok on this plate. Repeat! phage?

Y40 V₁^R V₃^S V₅^S. compare original description
 K12 = S...
 488-1 = S...
 488-2 = V₁^R, S...
 Y86 = V₁^S! (unstable: reverted??)
 Y65 = R....
 Y68 = S....

Repeat T3, T1.

	T1	T3
Y40	R	S
K12	S	S
488-1	S	S
488-2	R	S
! Y86	S	S
Y65	R	R
Y68.	S.	S

Unstable resistant?? Less mucoid on this plate

streak out {

~~streak~~ Y86 is predominantly mucoid; a few smooth colonies
 this ~~streak~~ is predominantly smooth; a few mucoid colonies.

Campthor & Polyploidy

499

April 25, 1947.

Add varying amounts of 30% Campthor in 95% Alcohol to plates, to give following "concentrations" of campthor. Incubate 3 days. ^{of M. l. agar} streak 153:

- A. 0
- 2. 100
- 3. 1
- 4. 2
- 5. 5
- 6. 10

Very little growth inhibition was noted except in # 6 (10% campthor!) where there was considerable retardation. Comparison of cells from 6 and 1 reveals the presence of many beaded, ~~slightly~~ slightly elongate bacteria.

Streak out 6 on EM13 to isolate clones and test for diploidization by the suppression of recessive mutations (e.g. cl_1^+). Many smooth-mucoid colonies noted.

Papillae on LA?

1	±	16	+
2	±	17	++
3	+++	18	+++
4	-	19	++
5	-	20	+
6	++	21	+
7	+	22	+
8	-	23	++
9	-		
10	++		
11	+++		
12	±		
13	+++		
14	±		
15	±		

Recover 4, 5, 8, 9 to test for polyploidy.

Utilization of Acetyl Glycine

~~447~~
500

April 27, 1947

Sec 480. ~~Glucose Glycine~~
Acetate ~~Glycine~~ Glucose

48h. - 60h.

~~48h~~ 489 Y K-12 K
489. K-12

Group	Concn	1	2	3	4	5	489 Y	K-12 K	489.	K-12	
A.	.1%	1	✓				-	-	±	±	± ±
		2					-	-	-	±	± ±
		3			✓		+	-	±	±	± ±
		4	✓				-	-	±	±	± ±
		5	✓		✓		±	-	-	-	-
B.	.2%	1	✓				±	-	±	±	± ±
		2					±	±	±	±	± ±
		3			✓		±	±	±	±	± ±
		4	✓				-	±	±	±	± ±
		5	✓		✓		±	±	±	±	± ±
C.	.5%	1	✓				-	-	±	±	± ±
		2			ACETATE		±	-	-	±	±
		3			GLYCINE	✓	±	-	-	-	±
		4	✓			✓	±	-	-	±	±
		5	✓			✓	-	-	-	±	±
D.	1%	1	✓				-	-	-	±	± ±
		2					-	-	-	±	± ±
		3			✓		±	±	±	±	± ±
		4	✓				-	-	±	±	± ±
		5	✓		✓		-	-	±	±	± ±
E.	1%	1.		✓			+	±	+	± ±	
F.	.5%	1.		✓			+	+			
		2.			+		± ±	± ±			

Glucose 1% throughout
Readings at 24h;

Concl. Glycine is not utilized; not inhibitory
Acetyl glycine is utilized by both
Acetate is not utilized by mutant comp/wild

Staining in zone of lysis.

501

April 27, 1947.

Compass 453 lysed by T'ca:

EMB - lactose :

- sucrose :

- blank. :

all show coloration in ^{margin.} lysis zone, suggesting that it is mostly staining of debris.

Segregation of A2 and Ia

SD 2

April 28, 1947

A. Y90 x Y53
(Y40/Ia)

B. Y92 x Y53
(Y40/A2)

A.

T(0).

readings on complete are unreliable.
Technique for tests on synthetics should be developed.

B: Lac Vi.	R	S
T(B ₁). -S.	29	2
-R	8	1
+R	12	1
	<hr/>	
	49	4

Use 100v/ml NaN₃ in T(0)+B₁. Some heat too essential:

B ₁	B _M	Lac	V	TL	A ₂
-	++	-	S	--	S
+	--	+	R	++	R

Mostly R. ∴ A₂ is near TL.

ca 8% recombination.

either beyond or between T-L Use selection to locate

April 28, 1947.

A) 486 x 58-161

B) ~~486 x 440.~~

not useful. interesting types could be merely mutants. [Accelerate mutation?]

T(10) - no colonies (excluded??)

T(B₁) -

Mucoid character too poorly expressed, although many of the colonies picked looked as if there should be Muc. Is there progressive "attenuation" of this character??

A 1. 1. Streak out 486 stocks on EMB-lactose; ~~etc.~~

34 Muc: 31 Smooth.

P 2 2. Streak out: A. Muc from 1.

B. Mix pop. from 1.

A: "all" mucoid ~~B.~~

B: 19 Muc: 70 Smooth.

P 4 3 A - mucoid from 2A.

B. Mix pop. from 2B.

all mucoid.

ca 100:1 smooth: mucoid

A 6 4. A - mucoid from 3A.

B Mix from 3B.

all mucoid.

> 200:1 smooth: mucoid

P 7 5. B. (mix) from streak of 4.

all mucoid

P 10 6 streak from mass streak of 5: ca 10 Mucoid: 1 smooth.

Selection and mutation of V_{mi}^R

503a

May 15, 1947.

A15. 7. Stalk from mass-stalk of 6.
ca 45:20 M:Sm.

A17 8. Stalk from mass-stalk of 7.
ca 23:43 M:Sm.

P18 9. do. 9:21 M:Sm.

P20. 10. do. 19:48 M:Sm.

P22 11. do.

Acetyl utilization

April 29, 1947

Ac.	Glucose	Glycine	12-24h.		36-48h.		60h.		
			K-12	Y89	K-12	Y89			
1.	1/2%		-	-	-	-	+	±	
2.	1/4%	1/4%	-	+	±	+	++	±	
3.	1/4%	1/4%	-	-	-	-	±	++	±
4.		1/2%	±	+	±	+	++	+++	+++
5.		1/2%	±	+++	+	++	++	+++	+++
6.		1/2%	-	-	-	-	-	-	-
7.		1/4%	-	-	-	-	-	-	-
8.	Acetyl-Glycine	1/2%	±	±	-	±	+	+	+
9.	o	o	o	o	o	o	o	o	o

Autoclave separately from medium. Adjust acetate to pH 6.8 ± AcOH before using.

The differential between K-12 and Y89 on acetate is not complete; there is a definite residual growth. Stimulation by glycine (not used by itself) accentuates the difference.

(Use either aerobic or anaerobic conditions)

84h: K-12 Y89.

1. Ac.	++	+
3.	++	±
8	+	+

! } eventually the bug does better on acetate than on Ac Gly!

diacetyl-diketopiperazine "2%"
 neither K-12 nor Y89 showed any response!

Tests of Camphor treatments

May 1, 1947.

See 499.

Recover presumptive strains.

a) streak again on CA agar. b) Cross \bar{c} 440.
 4, 5, 8 finally threw off many variants
 9 only \bar{c} .

c) streak out on EHB lactose
 all isolates are $\text{lac}^- \text{V}_1^s$
 but semi-mucoid character
 interferes with determination
 of serotype.

b. P2. ~~4 ml mixtures~~ 0.1 ml mixtures into B₁, plain agar respectively.

1. 499-4 2. 499-5 3. 499-8 4. 499-9. x 440.

Discrepancies between O and B₁ plates are only ca 3-fold rather than 10-fold.

1-(C0).	Smooth				Mucoid			
	+R	-R	+S	-S	+R	-R	+S	-S
① T(O)	1				2	9		1
B ₁	11	5			9	22		3
② T(O) —								
T(B ₁)	14	32	1	11	typical segregation.			
③ T(O)	1					2	1	
T(B ₁)	13	1			1	21		
④ T(O)	2	8			2	10		
T(B ₁)	16	21			2	16		2

The 453 2x doubled is not a good test; better would be 440 which carries more dominant alleles.

Resistance Patterns.

	①	②	③	④	⑤	⑥
	T-1	T ₃ A	T ₃ B	T ₃ Batch 2	T ₅	T ₅ Batch 2
K-12	S	R	S	S	S	S
Y40 (from 140 phif)	R	R	S	S	S	R
Y40 phage	R	R	S	S	S	R
Y53	S	S	S	S	S	S
Y64	R	S	S	S	S	R
Y87	R	R	S	S	S	R

probably contain T₁. Start new stocks.

large + small plaques

large + small

This phage stocks which have varied, not original cultures since Y40 = Y40 in all respects. T₃A must be fallacious. T₅ Batch 2 behaves like T₁ and is similar to previous responses. Could it be contaminated??

Phages and purification! Recheck T₃A. Present indications favor the interpretation that the results of last fall were due to gross contamination of T₃ and T₅ & T₁.

Program: Purify T₅ and isolate components.

⑤ was streaked out and exhibited both large and small plaques. Pick from a large and a small plaque and streak each with K-12 and Y40.

⑤ T-5 from original culture (Demerec) was plated with K-12 but gave uniform lysis. several mutants appeared; first these with T₁, etc. Use this to reinitiate T₅ stocks.

Phage stocks

507.

1. Start new T5 stocks from 1) lysate using original T5 on K-12
2) small colony picked from existing T5.

2. Other stocks OK. Renew T1 on K-12.

3. Test a large-plaque component of old T5 on K-12, Y40, K/5.

4. Test T1 on K-12, Y40, K/5. (from 506 R5).

	T1	"T5" large.	"T5" small	"T5"
K-12	S	S	S	S
Y40	R	R	R	R
K/5.	R	R	R	R
"	R	R	R	R
"	R	R	R	R

These are isolates

from 506 - (5) which, previously, lysed Y40.

T5 from original bottle

May 2, 1947.

~~Plate~~ Pour ca 10^9 / .1 ml on middle of NA plates + irradiate at ca 2500 r/min 20 min \approx 50,000 r. Weight 60 to 25 mg.

1. After shaking agar strip in H_2O ca 3h, streak out on EMB.

~~2. Streak out original sample on EMB. (Y40 only.~~

3. Killing very great. Probably only ca $10^2 - 10^3$ survivors.

Streak out proliferated cultures on EMB.

Isolate 14 colonies each from Y40X and Y53X and streak across each other. Plate mixed growth on T(0) agar. (28 tests).

of protts.

1	20
2	30
3	20
4	10
5	30
6	20
7	20
8	20
9	200
10	10
11	20
12	20
13	100
14	10

No crossover suppression here!