

11 SEP 1946

S12	L, I, H, C. ±	L, I, C. ±	L, H, C. ±	H, I, C. -	L, I, H, M. -	Leucine, cystine ...
S13	L, C ± ++	C -	L ±±	L, M ±±		leucine, methionine cystine
S14	L, A, M -	L, M, C ++	L, A, C -	L, A, M, C ++	BAK	Leucine, methionine, cystine
S15	L -	O -	LC +	C -	LM. +	leucine, S
S16	L -	C +	LC ±			leucine S
S17	L -	C -	LC -			
S1 (9) S1	M -	T ₁ -	MT ₁ +			methionine, trypt
450.	O	N, C				

1 SEP 1946

1 ml ino. S1, S17 (S15?) in 4B \bar{e} single controls. Sh. 30°

1230A II

Plate N13. into T(0) dupl.

no colonies on any plates.

12 SEP 1946

	ENV	EN	NV	EV	O	thioglyc.	
S18	+	+	+	+	-	+	} these must be a substitution to acct. for these results; probably cystine + methionine. Check with thioglycollate also
19	+	+	+	+	-	+	
20	+	+	+	+	-	+	
21	+	+	-	+	+	+	
22	+	+	±	+	+	+	
23	+	+	+	+	-	+	} (also: arginine - prot; glut ad. - tyrosine
24	+	+	+	+	-	+	
25	+	+	±	+	-	+	} φ-alanine - tyrosine tryptophane - leucine
26	+	+	+	+	+	+	
27	+	+	±	+	-	+	
28	+	+	+	+	-	+	
29	+	+	±	+	+	+	} brought?
30	+	+	+	+	-	+	
31	+	+	+	+	-	+	
32	+	+	+	+	+	-	} parathiotroph.
33	+	+	+	+	-	+	
34	+	+	-	±	-	+	} parathiotroph
35	+	+	-	+	-	+	
36	+	+	±	+	-	-	
37	+	+	±	+	-	±	} parathiotroph
38	+	+	±	+	-	+	

Ev. no good mutants here!

S19 } parathiotrophs; others are prototrophs
S32 }
S34 }

S35-8 may be mutants.

These tests were not too careful.

In interest, only S36, S37 held up as mutant types.

Streaks for linkage study.

3/2

18 SEP 1946

IP18 inoc NSB to prepare inocula.

A19 - plate \bar{c} T1, T3 to obtain resistant mutants.

A22: Titer of T3 is very low + continuous lysis is not obtained!

Y41, TL, BL, BT, no sens.

Y10/1 completely lysed!
(~~without~~ rechecks).

Y24, BB, LB, *

Y10/3 OK for resistance.

TL/1 and TL/3 plates have \bar{c} mucous dense white colonies.

do. LB.

Y24/1 on T3 showed no lysis. (cross-resistance \Rightarrow - confirm!)
3x trials before used.

BM/3 OK. R.

Pick to YB tubes. Test for
resistance to 1; 3.

BB/1 - mucoid.

Y10/3 OK R.

BT/1 Mucoid.

LB/1 v.s. colonies v. dry - probably cont.

LB/3 } looks like coli, but green!
TT/3 }

Lineage: BM R x PT; BM x PTR.

313

18 SEP 1946

1 P18. Inoc 4B to prepare inocula, 679-183 from tyophil.

3 P24. Inoc ~~10~~ 4B.

Plate P26

1. PT x Y24/1 5 tubes. -

2. PT x Y40 5 tubes

3. Y41 x 58-161 5 tubes.

R to T1.

1: 27/30
20/20
27/30.

}

$$74/80 = .92$$

PT x ~~BM~~ BOC/1

2: 18/20
26/30
5/7

}

$$49/57 = .86$$

PT x BM/1

3. 0/3
0/1
4/10
1/10

}

~~5/10~~
 $5/24$

$$.21$$

PT/1 x BM.

Ref.

257 BM x PT/1 2/10 R .02
 BM/1 x PT 7/10 R .71 compare 313

259 BFC x PT/1 2/9 R .2	313: $\frac{30}{137} = .22$.78	BFC/1 x PT. 27/30
278 " 16/60 R .27		20/20
284 " 5/20 R .25		27/30
284 " 5/21 R .24		74/80 = .92

287- BFC x PT/1 2/27 R .1
 on B: 5/23 .22 .92

293- BFC x TLB, /1 10/35 R .29
 290A " 10/29 R .34
 290B BFC/1 x TLB, 19/26 R .73

301-1. BFC x TLB, /1 20/49 R .41
 BFC/1 x TLB, 27/37 .73

SUM: A. BFC x TLB, /1 49/183 .35
 B. BFC/1 x TLB, 46/63 .73

$\chi^2 = 0.8$ for .35 vs. .27 (1-.73)
 $\chi^2 = 9$ for .35 vs. .50
 $\chi^2 = 5.5$.73 vs. .50

1.08

315- BFC x TLB, /1 13/73 R .18
 318 BFC/1 x TLB, 34/40 .85

~~388~~ R: 440 x 493 464 x 58-161

SUM:	A. 424 x 410/1 53/186 .28	
	B. 424/1 x 410. 80/103 .77	

353 47/57
 358 16/18
 364 49/70
 359 32/50

313 BM x PT/1 0/3
 0/1
 4/10
 1/10
 5/24 .21

313	BM/1 x PT	18/20 .86
		26/30
		5/7
		49/57

19 SEP 1946.

A. BOC x TLB | 1

B', A' is mixture in old medium

B. BOC/1 x TLB.

Use 1ml inocula for flasks; .1 for 10ml medium in tubes

a) 5 of A; 5 of B in 50ml YB 30° sh. (A_L, B_L)

b) 10 each in 10ml YB in tubes, 30° sh.

Wash + plate N20.

A series: same large col.
many small
heavily seeded microcol.

A

Prototypes.	T-1 R.
a:1	2/8
2	0/3
3	0/4
4	0/13
5	2/7

B

B series: all heavily turbid; maybe contam, or
ecom. rate may be very high.

A' numerous col. - not coli etc. 1/10 3.
B' like other B plates.

A

b. 1	0/3
2	0/6
3	2/7
4	2/5
5	1/3
6	0/5
7	5/10
8	1/6
9	
10	
Sum total:	13/73 = .18

B.

- 1
- 2
- 3
- 4
- 5
- 6
- 7
- 8
- 9
- 10.

Y10/1 x Y24/1.

3/6

20 SEP 46.

10 ml. YB at 30° also controls

n.g. - Y24/1 cont
start new tubes

Staph. flavovirens

317

20 SEPT 1946

ENV	NS.	NV	EV	EN
++	+	+	+	-

try on vials - n.g. ~~Repeat!~~

24 Sept 1946

In 10 ml YB, 1 P24.

do. - spread .1 ml mixture on
CC plates.

(A) + (B)

1. Y24 x Y10/1 (A)

2. Y10 x Y24/1 (B)

Wash + plate p. 26.
with (0).

controls: Y24 - no colonies
Y10 - no colonies
Y24/1
Y10/1
in broth

(A) (B) p 28 - colonies as in both tests.
No evidence of marked increase.

B: $\begin{array}{r} \Gamma - 1 R. \\ 28 / 33 \\ 2 / 3 \\ \hline \text{||} \\ \text{||} \\ \text{||} \\ \text{||} \\ \hline 34 / 40 = .85 \end{array}$

A: $\begin{array}{r} 1 / 9 \\ 4 / 10 \end{array}$

September 29, 1946.

P29. B4/3 x P7/1 in YB Broth 10ml.

P1. Plate into T(0). only a few colonies.

30 SEP 1946

N30.

A. To 10ml YB, add 1 drop 24hr. Y2Y/1 and

10² 1. 1 drop 679-183 (PT) * compare \bar{c} final PT population. 1 drop = 0.1 ml.
 10³ 2. 10⁻² drops. " use.

Dilution: 10² 3. 10⁻⁴ drops " (plate .1ml.). original = ca 10⁹/ml.
 15. 4. 10⁻⁷ drops. " ~~plate~~ OK.
~~5. 10⁻² drops. "~~

B. To 10ml YB, add 1 drop 24hr. Y2Y/1 and 1ml of the following.

b see A1.

- o 7. 1ml u-v irradiated 5 mins. plate .1ml. 10⁴
- o 8. 1ml u-v ir. 10 mins. 20
- o 9. 1ml u-v. ir. 15 mins. 20

Plate P into T(o).

∴ sublethal doses of ultra-violet prevent recombination!

30 SEP 1946

11P30. Inoc 10ml Y10 Y10/1; Y24.

P1. 1. Mix cultures, centrifuge and wash. Plate into T(0);

A. Centrifuge together 3-4 hours to pack cells together. 0, 1, colony.

B. Wash ^{separately} ~~together~~ + plate together. 4, 2, 0 colonies. Ten days to pick.

This, therefore, is the better procedure.

Salmonella typhi - murium

322

Mutants - (u.v.).

30 SEP 1946

P1. Inoc 520 and 527 into YB.

11P2. Incubate each culture 1, 2, 5 mins. + inoc 1 ml samples into 10 ml YB. Detection plates SP3. 25°.

A 1 Y20-1m
2 2m
3 5m use.

2000 cells sampled. 7 small colonies.

Pick A8.

B 1 Y21 1
2 2
3 5 use.

1200 sampled

2 small colonies.

	YB.	O.	E	N	V	ENV
A 1	+	+	→			
2	+	+	→			
3	+	+	→			
4	+++		→			
5	+		→			
6	+++	-	+++	+++	-	+++
7	+++		→			
B 8	+		→			
B 9	+		→			

green cont.
green
pink cont.
OK. ni color
yellow
OK. 570.
OK. ni color.
green cont.
yellow

parathiosoph

some app. do not grow well even on YB

570:

30 SEP 1946

A1:

T(10) ENV EN EV NV

524 abeg.	-	-	++	++	+	++	±	++
36 gall.	-	-	++	±	+	++	±	±
37 dubl.	-	+	++	±	++	+	±	++
42 t. n.	-	++	+	++	±	++	±	-
45 ent.	-	-	±	++	±	++	±	-
50 para A	-	-	++	-	-	(±)	-	-
51 B	-	-	+	++	+	++	+	++
52 B	-	-	-	-	-	-	-	-
55 chol.	-	-	±	++	-	++	+	-
56 chol.	-	+	±	++	±	++	-	±

? (Ethin) grows well as T(10). [Fisch for] V, E. Thiamin
 (E) Tryp. Thiamin.
 E. Arginine ✓
 E, N, V. BIOTIN, TRY.
 (N) ? - clasp?
 E(N) TRYPTOPHANE; (methionine); ~~serine~~
 () SERINE.

The cleanest ones are:

- ✓ 542 - typhi mucrosus - (E)
- ✓ 545 - dublides (E)
- ✓ 550 - para A (ENV)
- ✓ 555 - cholerae suis (E(N))

others are:

- 524 - prototypus
- gallinarum V, E
- dublides } strain? wosp. req.
- 556 chol. suis }
- 550 para B
- 552 - did not grow

42 no. gr. Trypt.

45 Arginine (citr +) (over)

50 Biotin; TRYPT; grows S.N.E.A. (meth) BT CM n.g.

55 TRYPTOPHANE; SERINE T+S n.g.

36 ENV+++ V++ E++ N- 0-

37 ENV+++ V# E# N- 0- Try Valine

56 ENV+++ V-# E-# NE-# 0- F-#

V or E + - B.
 V or E + - B.
 E or NE

|| 52 ENV+4 EV+ EN- NV- 0- they all.
 || 51 ENV+++ V+ E+ NE- 0-

no!!

Penicillin

1 OCT 1946

add ind. amts. of Penicillin (oxford units) to 10 ml YB broth.
add 2 drops of Y24 + Y10/11. 10 P1. Sh. 30°.

Penicillin (per 10 ml).

10A2.

- | | | | | |
|---|----|------|------------------------|-----------------------------------|
| 3 | 1. | 0 | | |
| 1 | 2. | 100. | | some very motile |
| 0 | 3. | 200 | v. "stringy" turbidity | v. filamentous |
| 0 | 4. | 500. | partial inhibition. | v. filamentous; many "zygospores" |

Wash + plate ..

10A2 also inoc. 1 ml each into 10 ml YB.

- | | |
|---|----|
| 1 | 1' |
| 7 | 2' |
| 2 | 3' |
| 8 | 4' |

penicillin is sufficient.

Edwards 10/9/46.

S	O	ENV	EN	EV	NV
57 typhi suis	-	++	-	-	-
58 abortus ovis	-	+++	++	+++	-
59 sendai	-	+++	+++	+++	+ ++
60 sendai 571	-				
61 Taube I	-				
62 Taube II	-				
63 P-1	-	+++		+++	+++
64 P-2	-	+++	++	+++	+++
65 P-11	-				
66 1181-1	-	+++			
67 1181-2	-	+++			
68 1166	-				
69 Pigeon 1366.	-				
70 327/65 20.	-				

ENV
E
Try E. (uses any 9 EAH)
E or N? methisamine. 56 etc. do.
caripal

E; N. +³ methisamine. (SH.).

Perth Israel 10/21/46

- 71 typhi suis 2943
- 72 sendai 3007
- 73 para A 3280
- 74 para A 3089
- 75 typhi 3166

Wageningen

- 76 S. para A
- 77 " deuringo
- 78 typhi 2
- 79 " Wageningen
- 80 S. sendai
- 81 negdam -
- 82 " +

2 OCT 1946

1. P2 Y24 x Y10/1.

[2. Y24 x 183]

[3. 183]

4. Y24 x Y41

5. Y41.

Plate .5 ml eq. (1) .2 ml eq. 4, 5 into T(0) and suppl. to detect linkages of R.

Plate .5 ml eq. Y41 into O, T, P.

20 colonies on T(0)!
crowded on plate.

Exp 1. 4:

12 OCT 1946

Moze 1 deep of 12 hr. cultures into 10 ml 1/13. Plate 10/14/46.

S1 x 13
1 x 45
1 x 50
1 x 55
1 x TL

cdi)

1 colony found. [pair A + pair A!!!!]. Needs rechecking.
~~later found that S1 sweets.~~

13 x 45
13 x 50
13 x 55
13 x BT

cdi)

45 x 50
45 x 55
45 x TL

cdi)

50 x 55
50 x TL
55 x TL

cdi)

1
13
45
55
TL
BT

repeat. S1 x 550. 10/16/46. Plate into T(0), T(trypt).

very many minute colonies in presence of tryptophane. Should be repeated in a more diluted inoculum. v. much smaller (a few cells / colony) on trypt.

Infections

326

20 Oct 1946

Mix 2. growth cultures. N2. (4B) 30° sl.

424 x 410/1

Plate 3P3

~~Infection plates - 7(0) - 10⁻⁷ - P2~~

2 colonies. (1 inf eq.)

Tritilateral combinations
test for transformation.

12 OCT 1946

Inoc 1 Loop 12 hour cultures into 4B Plate 10/14/46.

	O	T	L	B.
TL	-	-	ca 10 ⁴ .	
BL	-	B	L	
BT	-	B	T.	
		+	+	

BT+TL } 1/4 plates (turbid). turbid. ca 10³
+ BL } not so numbers?

This expt. is inconclusive, since there were a large number of L⁻ cells present. Quantitatively, it seems to support the recombination hypothesis.

Reisolate TL.

Salmonella - N.R.

545, 550, 555.

545. ✓

1	2	3	4	5	6	7	8	9
0	PROL	GLUT	ORN	CITR	ARG.	Aminothione		
-	-	-	+++	+++	+++			

550. ✓

NV-TR. ✓	NV-TRM. ✓	V-TAL ✓	V-TAM ✓	N-B-E ✓	BIOTIN+E ✓	E ✓	NE+E ✓	NE.V. ✓
-	±	-	±	±	-	-	++	+++

555 ✓

SERINE-E ✓	NE-TR ✓	Ser-TA ✓	NE-E ✓	E ✓	NE ✓?	try Trypt + meth.		
+++	+++	- +	+++	++	+++			

556. ✓ NE-

Tyrosine (??)

0.

536 ✓ V

(B₂) also 0.

537 ✓

B₁ ✓
B₂ ✓
B₆ ✓

B ₂	3	4	5	6	7	8	9	10
B ₁ only OK. Thiamine ✓								

542 E-

Tryptophane.

558 E-

559 E-

Knows on any combination of 9 EAA.

561 E-

Knows on any combini. of 9 EAA.

(Heterozygotes)

327.

4 OCT 1946

brox 24 hr. cultures of Y24 and Y41 into T(0) varying amounts.
(washed separately. In water together ca. 3 mins. Mix with agar before
pouring. A4.

most plates sterile. A few have many non-coli bugs.

15 OCT 1946.

Add 440+679-183. & varying chloral hydrate

	Concn.	Growth.	Plates:
1	1%	±	
2	.1%	+++	ca 100.
3	.01%	+++	ca 500
4	.001%	+++	
5	.0001%	+++	ca 500
6	.00001%	+++	
7	-	+++	ca 10 ³

sl. (?) inhibition, but not feasible for exp. use.

plate. A18.

Compare, however, effect of 5% salt.

But swimming in motility-gelatin-agar is only partially inhibited by .1% chloral hydrate (1-2 cm/24h.) Try .2%, - 5%.

Vit. Req. - nei. Fris.

no growth - pal

delayed growth - nei.

Salmoneella NR

21 Oct 1945

536. ✓ 0 - B. Vits. ~~+++~~ ~~+++~~ OK. Thiamin. ✓

556 ✓ 0 - tyr tyr+glyc Vits Tyt+Vits @A+Vits which Vits? ✓
 +++ +++ ++ +++ ++

550: 0 ENV EN NV EV Vhy try leuc^{NV} try leuc try meth try leuc^{NV}
 +++ - +++ - -

555. ✓ 0 - Su + E E Serriv Tr. NE Tryp. NE-Ser. + tryp. NE
 + ++ ± ++ - ± + ++ ± +++ ∴ Tryp. + Ess.

542. ✓ 0 - Tryp. n.g. cella ~~very slow!~~ Tryptophane.
 ± +++

558 0 - Tryp.

559 ✓ 0 Tryp. single amino ac. E: any a.a. +++ @ ++. adapted??

560 0 ~~Tryp.~~ single amino ac. E: meth +++ others -

507
567
564
etc

570. 0 Tryp. single amino ac. E: meth +++ others -

Sauv. Luid.

grows on B vits. etc
 - pet, folii OK on - folii. ∴ - pet, - mic!
 - mic.

23 Oct 1946

	O	EN	ENV	ENB	ENB.	NV	BN	
S50.	-	-	+++	+++	-	(+)	(+)	∴ Biotin. can disp. z.e.a.a.
S61	±	-	Cyst	Meth.	+++			Trypt. histidine Pantoea
S70.	-	±	+++	+++	++	+++		two parathiosph!
S42	-				±±	✓ ± ±	✓ - -	autolys → inf → trypt. ✓ E. F. Ideo.
S62								
63								
64								
65								
66								
67								
68								
69								
S9.	±	±						heavy moi. ±± (and adapts!)

S56. Try all amino acids + vitamins separately. V +++ E +.
 Protein ++ no E. alanine ±. lys. ±
 trypt ± tyros. ± || 48 hrs: Biotin +++
 alanine ±
 tyrosine +++

S55. Trypt + NE (all +; letatine) and ~~trypt~~ NE (all):
 T + N +++
 N ++
 individually: - - exc. cysteine ++
 O - heavy moi. in O +++
 OK. Try cyst. alone.

later O: -
 any NE. ++
 all NE do.

Test growths in peptone, for transf. growth
 in minimal, peptone; compare original culture.

E. coli mutants.

24 OCT 1946

Selective media (lactose)

1. EMB Difco.

OK.

2. Linds: lactose 1%
Ker SO₃ 2.5
Fuchsin .4g
K₂HPO₄ 3.5g
Nutr. agar

u.g. arranged up.

3. Purple lactose:

Nutr. agar
lactose
BCP .025g/l.

u.g.

Streaks K-12, S1 and K-12+S1 on each of these plates, for decision as to most appropriate medium.

S50 Biotin +: (NV)- (EAA) BV + none grow.

S56. O BIOTIN TYROSINE dl-tyrosine 'Biotin'??
 - ± ± ±
 - ± ± ±

S55. ✓ O TRYPT. CYST. TR+CYST. TR+PROL.
 S59. ✓ - - - - -
needs Trypt, cyst adapts readily.

S57. EN V- : none grow! -Leucine found later (tubes allowed to stand)
 EV N- :
 NV E- :

S58 E- : none grow!
 O ENV NV EV.

S60
 S51 NE+ proline +++. others - NE+++ Serine ±

S52. NV E-

S61. ✓ O Meth. ~~NV~~ ~~NV~~-cyst Cyst.
 +++ +++ +++ +++ +++ all adapt!

	Meth.	Tryp.	
62	++	+++	++ ✓
63	+	+++	± ✓
64	+	+++	± ✓
65	±	+++	- ✓
66	±	+++	- ✓
67	±	+++	- ✓
68	±	+++	- ✓
69	-	+++	- ✓

Yeast

Arado-lectare	K-12	S-49
a. Mcd School	+++	- ±
b. mids up	+++	-
c. Arado-glucose	+++	+++
d. Arado-sucrose.	-	±
e. maltose	+++	slow +

EMB.	N.A. + 1% sugar + 2g K ₂ HPO ₄ , .4g Eosin Y; .065g MB/L.	
a. lactose	- +++	-
b. glucose	+++	shrunken +++ kosher!
c. sucrose	±	-
d. maltose	+++	-

Try: e oil suppl.

E. coli (T10) 1/2% glue n.g.

Fries (N10) 2% suc. requires pab.

* Burkholder's 5% gluc. only on vits.

E. coli T10 5% glue n.g.

- * / liter
- glucose 20
- aspar. 2
- KH₂PO₄ 1.5
- MgSO₄ .5
- CaCl₂ .3
- (NH₄)₂SO₄ 2.0

no K1.7