

March 10-12, 1942.

Y10 x Y45.

A) $T(B_1)$ plates.

Strata on lac S agar.

+	-	/ 20
18	2	

7 tested were all T_1^S as expected.B). Lac S B_1 plates.

Add. A 11.

+	-	/ 149.
$\begin{array}{r} 41 \\ 72 \\ 32 \end{array}$ \hline	$\begin{array}{r} 1 \\ 2 \\ 1 \end{array}$ \hline	
145	4	

Recount A 12.

LB,

+	-	/ 412.
$\begin{array}{r} 100 \\ 121 \\ 71 \\ 117 \end{array}$ \hline	$\begin{array}{r} 8 \\ 13 \\ 3 \\ 9 \end{array}$ \hline	
409	33	

Lac - = 7.5%

Compare with 8.6%
of p. 42.

Lac (o)

+	-	/ 70
$\begin{array}{r} 31 \\ 35 \end{array}$ \hline	$\begin{array}{r} 2 \\ 2 \end{array}$ \hline	
66	4	

5.7%.

March 8, 1948.

Cross on Lac(0) Agar, W~~2232~~ 337 with the following:
3 plates each (.1 ml scoop.)

w45.

No colonies.



w35 8 Lac - colonies all told.

w72 3 Lac - colonies all told.

Y87 9 Lac - colonies.

Crosses should be repeated.

Glucose - 1-phosphate

153.

Mix up T(m)-BHTLB, + equivalent of .05% glucose in 5cc volumes.

Granulate lightly with: P10. [Filter - sterilized].

	Y10	W-108	W-327
1. K. glucose-1-phosphate (barley)	P11 -	-	-
	A12 -	-	-
	A12 -	-	-
	A13. -	-	-
2. Glucose.	++ - ✓	- -	- -

March 9, 1948.

T(B₁) Y10 + Y87. Measured digest dilute suspension.

A) + 4 ml H₂O/plate B) 4 ml H₂O + 700 μg B₂ + 35 mg Glutamate
100 ml medium.

A. P10 (ca 36 h.) 33 / 7, 12, 9, 5 m = 8

B. 34 / 4, 11 (2 days), 6, 13 m = 8 1/2.

No pronounced effect of B₂ + glutamate.

(More colonies may appear later). 12 appeared altogether.

See 155.

March 8, 1948.

1. Y-87 X Y-10

2. Y-53 X Y-40

3. W-183 X Y-46

IA On EMS (-B₁) plates.

a) Readings from plates.

XWY5
"reading label!"

+ 13 13 10	- 5 5 4	b. S.O. T-Lacs'
22	22	
36	14	
63	44	77.
36	18	

a' Repeat A12. :

64	78	82	52.	14	3	27	8
36	18						

IB. On EMS(B₁) lac plates

a. direct counts.

a' repeat A12

17	4
16	8
16	3
30	15
30	5
16	3
22	11
28	11
19	3

Total : 6 sectors.

194	63	257.
227	101	338

IC. From T(B₁) plates.

See page following for raw data. Totals of all experiments this page are:

S. 445 (.294)	-R 131	-S 6	+R 207	+S 101
		.013	.465	.227

Cf published results:

13	211	13
----	-----	----

1a. Scored originally as Lac+.

-R	-S	+R	+S.
0	0	14 13	3 <u>5</u>

As Lac-

14	3		
<u>.14</u>	<u>.3</u>	<u>.27</u>	<u>.8</u>
.269	.058	.519	.154

] 52:

1B. As Lac+

+?	1	1	29	13
0	0	33	21	
0	0	15	8	
0	0	12	9	
0	0	16	6	
0	0	13	7	
1	0	13	5	
1	0	15	5	

As Lac-

49	1	0	0	
39	1	0	0	
1 91	3	146	74	314,
<u>.290</u>	<u>.095</u>	<u>.465</u>	<u>.1236</u>]

1C. (plates = B₂, glut). 16 0 21 12

10	0	13	7	
26	0	34	19	79.
<u>.329</u>	<u>0</u>	<u>.430</u>	<u>.240</u>]

131	6	207	101	445✓
<u>.294</u>	<u>.013</u>	<u>.465</u>	<u>.227</u> ✓]

A total of 6 sectored colonies were noted. These were purified and tested with T1. All 12 cultures were V_1^R .

	BM	Lac	V_1	TL
O	-	X	R	+
O	-	-	R	+
O	+	X ₂	S	+
	+	+	S	-

No X_1 +R. X_2
 -R. +S.

In calculating p , the chances of X_2 being in Lac- V_1 , $\therefore V_1 \div TL$ only should be considered. X_1 is almost completely fixed in region D as -R. An expectation of 4:2 is not sign. different from the experimental value of 6:0.

Test on B, for requirement.

A Lac - B Lac +

~~A~~ B

1. B,
2. + B,
3. B, B,
4. B, B,
5. B, B,
- 6.

Also, test Y10 on pyrimidine + thymole:

1. TL -
2. TLB, +++
3. TL P_y +
4. TL-Th ++
5. TL-Th P_y. ++
- 6.

specific Reasons.

#64

April 8.

Streaks out W-108 on MB glucose, mannose, fuctose. EM 13

Apr 17. No papillae seen on these plates.

Lactose Analogues

167.

March 31, 1948.

Test strains on lactose, epi-lactose, neolactose + galactosan received from N.K. Richtmyer. 1% - EMB (small plates).

	<u>Str.</u>	<u>lac</u>	<u>Neolac</u>	<u>Epilac.</u>	<u>Galactosan</u>	[M. gal].
1	+ K-12	+ P	- *	+	-	+
2	+ Y10	+	-	+	-	+
3	lac ₁ W-53	-	-	-	-	+
4	lac ₂ W-45	-	-	-	-	-
5	lac ₃ W-108	-	- P	-	-	+
6	lac ₄ W-126	-	-	-	-	-
7	lac ₅ W-145	(+)	-	-	-	+
8	lac ₆ W-125	(+)!	-	±	-	-
9	lac ₇ W-133	- P	-	-	-	-
10	sl. W-117	- P	- P.	-	-	-
11	sl. W-252	+	±	+	-	+
12	sl. W-328	+	-	+	-	+
13	gal-W-254	+	-	+	-	-
14		* Papillae to form & showing v. considerable utilization				

Galactosan - all.

Lactose. All -

Neolactose all -

epilactose follows lactose.

Strains out papillae of K-12 / Neolactose in lactose. Test colonies on neolactose. 8+ 3-. Isolate + as W-341. Still lac + See over.

Inoculate 58-161^K into 25 ml T(m) + Neolactose 25%.
+ galactosan

Delayed growth on neolactose.

Streak out and test on neolactose EMB. 11- 0+.

Repeat streaking.

per 10 liter bottle.

Use technical grade chemicals.

NaCl	50	g.
K ₂ HPO ₄	30	
K ₂ HPO ₄	10	
(NH ₄) ₂ SO ₄	50	

Sugars 150 g. Sterilize separately.

Grow K-12 74h. aer., wanton, undil., with lactose.

Collect 44 g. cells Divide & incubate each portion for 3/4 hour in 100 ml 1% peptone + 5% lactose or glucose. for adaptation. Sediment after 3/4 h. & resuspend each in 50 ml 1/100 Na citrate under toluene & autolyse. P8 - P10.

Autolyzate volume after bartering are removed as 50 ml each. The autolyzates give very high blanks on Baiford's method, so they cannot be directly assayed.

∴ ~~soy~~ To 10 ml samples add 3.5 g AS + sediment. Assay ppt redissolved in 1/100 saline citrate. ~~use~~ 1/100 H₂SO₄.

G alone	< 1 drop.
.1 ml G + 10mg lac	.90
1.0 " " "	.41

Neither preparation hydrolyzed lactose beyond the blank (ca 6%).

L alone	< 1 drop
.1 ml L + 10mg lac	.90
1.0 " " "	.33

Lactose 10 mg. 1.14 [B (blank)].

Glucose + galactose 10 mg. 19.06
" " 1 mg. 1.97

163 B2 + lac. 5.42
" (blank) < 1 drop

$$\frac{5.42 - 1.14}{19.06} = \text{ca } 22\% \text{ hydrolyzed}$$

in 20 mins.

W-125, W-145

April 9, 1948.

In neolactose tests it was noted that W-125 and W-145 were positive or slow positive on lactose. When streaked out again as controls on outcrosses, this was noted again, and suggests the need for reexamination.

Streak out on lactose EMB and compare:

W-145 stock slant < 1% Lac - colonies. - colonies quite small.

W-145, lyophil tube All Gma -, Mal -, Lac -. Recover to slant.

W-125. Numerous fairly good sized colonies that might be considered slow. Streak out must be good +.

[It seems that ^{slow.} 145 colonies more + are more likely to be lac + than those further removed. This suggests a pH or redox effect.]

Apr. 9, 1948.

Inoculate 58-161 or Y10 heavily into T(m) TLB₁BM with 0.1% sugar.

25 ml.

24h. 48h.

1. Lactositol	Y10	±	++
2. "	Y10	±	++
3. "	58-161	±	+++
4. "	58-161	±	+++

Apparently lactitol mutants
can be selected for.

100 ml.

5 Galactosan 58-161

— — A28

6 Galactose 58-161

+++ — ~~+++~~

Throw out

A28. Strains out 1 and 3.
on lactitol which was +.

I was sterile. (3) gave 1 colony

A29. S.O., side by side W-349 and 58-161.

W-349 is pure tol+, but relatively weak; 58-161 is definitely -.

LACTITOL

EMB - 1% (from Wolfram, dihydrate)

K-12	-
Y10	-
Y53	-
W45	-
W-108	-
W-145	-
W-125	-
W-126	-
W-133	-
K-12 Neot+	- ± slow
5816t Neot+	-

see p. 170 for selection of Neot+ mutants.

Antibiotic Lactose variants.

173

W125, W145. Predominantly lac+ or streaking.

W126 x ~~58-161~~ 58-161. + -
lac- v. small colonies EMS

W133 x 58-161 + -
not so small 33 53
16 45
—
49 128

W45 x Y10 >10+: 1 -

W108 x 58-161.
3 types noted.
original streak shows not
but some variations.

++ $\frac{±}{25}$ 31. 76.

See W-342 ff.

April. 9/9/48.

410 5 mmis 4V Hanover.

L-Arabnose EMB. Ca 2000/plate unevenly spread + difficult to score.
36 plates = ca. 70,000 colonies.
11-30. 20 "mutants"

d-Xylose EMB. 50 plates. ca 1000 scoreable colonies per plate
1-10 ca. 50,000 colonies

	Xyl	Ara	Lac	10 "mutants"	Mal	Bru	Dna	Sal	T1
W -	-	-	+	+	+	+	+	+	
351	1	-	-	-	-	-	-	-	
352	2	-	-	-	-	-	-	-	
353	3	-	-	-	-	-	-	-	
354	4	-	-	-	-	-	-	-	
355	5	-	-	-	-	-	-	-	
	6	-	-	-	-	-	-	-	
	7	-	-	-	-	-	-	-	
	8	-	-	-	-	-	-	-	
	9	-	-	-	-	-	-	-	
L 360	10	-	-	-	-	-	-	-	
361	11	-	-	-	-	-	-	-	
	12	-	-	-	-	-	-	-	
	13	-	-	-	-	-	-	-	
	14	-	-	-	-	-	-	-	
	15	-	-	-	-	-	-	-	
	16	-	-	-	-	-	-	-	
	17	-	-	-	-	-	-	-	
	18	-	-	-	-	-	-	-	
	19	-	-	-	-	-	-	-	
370	20	-	-	-	-	-	-	-	
	21	-	-	-	-	-	-	-	
	22	-	-	-	-	-	-	-	
	23	-	-	-	-	-	-	-	
	24	-	-	-	-	-	-	-	
	25	-	-	-	-	-	-	-	
	26	-	-	-	-	-	-	-	
	27	-	-	-	-	-	-	-	
	28	-	-	-	-	-	-	-	
	29	-	-	-	-	-	-	-	S S S
380	30	-	-	-	-	-	-	-	R R

29 + 30 are probably contaminants, but nutrition should be checked.

EMB ± 1% glucose +. Read at 24 h.

1. 2% F. no growth.
2. 2% + G no growth.
3. 1% F. distributed growth; some papillae?
4. 1% F G small translucent colonies.
5. .5% F Moderate colonies translucent.
6. .5% FG large colonies. Milky or blue. ← good selection level.
7. .1% F Moderate colonies translucent.
8. .1% F G large, purple colonies.

9. 1% oxalate + .4% glucose
10. ~~1% oxalate~~
.4% glucose.

For formic "decarboxylase" selection medium, use
.4% Naformate, 1% glucose EMB.