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PST Test for γ growth in presence of UV inact. γ .

- 150 .05 cc 24^h irradiated B $\rightarrow$ 20 cc broth, aerate at 37° = B_I
- 10 γ assay (small tube black stopper, 1/7) : 2×10^6 $\frac{51}{52} = \frac{386}{305}$ [Y] = 1.4×10^{10}
- 4 B_I assay : 2.5×10^4 $\frac{1}{2} = \frac{152}{169}$ [B] = 8×10^7
- 2 .85 cc B_I + .15 cc [UV γ 10 min] = B_Y, at 37°
- 0 .02 cc [γ : 10] $\rightarrow$ B_Y
- 5 .04 cc B_Y $\rightarrow$ 2 cc broth of tube, prewarmed.

cf. 4^m, supernatant : 20 B $\frac{3-413}{4-407}$

	I 1 cc $\rightarrow$ 9 cc broth :10 B	II 1 cc $\rightarrow$ 9.9 cc broth :10 B	rel. assay
8	5-127 6-122		1
19	7-126 8-121		1
22	9-82 10-69 (:20)		1.2
30		11-3 12-9	5
50		13-41 14-46	34

In B_Y : [Y] input = 2.8×10^6 [B] = 6.8×10^7
 [Y] free = 8.2×10^6 = ~29%
 [Y] adsorbed = 19.8×10^6 = ~71%

[Y] inf. cont. = 1.25×10^7 (= Y free + $\frac{Y \text{ adsorbed}}{4.5}$)

Step size = 34 • [Y] in tube I : 8^m : 2.5×10^4
 50^m : 8.5×10^5

$\frac{Y_{50m}}{Y_{inf. cont.} - Y_{free}} = \frac{8.5 \times 10^5}{8.5 \times 10^3} = 100$

Comment :

γ growth is small and slow - Uncertainty due to unusually high γ titer (confirmed, however, by regular adsorption). Apparently only a small number of ^{infected} bacteria can show ^{up} as a plaque and give a burst (~1/5). The ratio $\frac{Y_{infect.}}{B}$ is about 10; a bacterium should have in average 8-8 inact. particles. Possibility of some particles of active γ reaching bacteria not yet infected by inact. γ .

Test for α adsorption on B, $[B + \gamma]$, $[B + UV_{inact.} \gamma]$

PST

-120 .15 cc 24^h aerated B $\rightarrow$ 20 cc br., aerate at 37° = B_I

-10 α assay: 2×10^6 $\frac{1}{2} = \frac{219}{180}$ $[\alpha] = 8 \times 10^9$

-3 B_I assay: 2.5×10^4 $\frac{3}{4} = \frac{215}{210}$ $[B] = 1.05 \times 10^8$

0 .85 cc B_I + .15 cc $[\gamma \text{ "}]$ + .02 cc $[\alpha:10] = A$
 1 .85 cc B_I + .15 cc $[\gamma UV_{10^m}]$ + .02 cc $[\alpha:10] = B$
 2 .85 cc B_I + .15 cc $[\text{froth}]$ + .02 cc $[\alpha:10] = C$ } at 37°

4 .02 cc A $\rightarrow$ 2 cc broth not prewarmed = B_A

5 .02 cc B $\rightarrow$ 2 cc broth " " = B_B
 6 .02 cc C $\rightarrow$ 2 cc broth " " = B_C

7 B_A: 50 [col. A] $\frac{5}{8} = \frac{95}{113}$

8 B_B: 50 " $\frac{7}{8} = \frac{118}{113}$

9 B_C: 50 " $\frac{9}{10} = \frac{184}{164}$

11 In C_A: 20 [col. A] $\frac{11}{12} = \frac{185}{200}$

12 " C_B: 20 " $\frac{13}{14} = \frac{243}{245}$

13.5 " C_C: 20 " $\frac{15}{16} = \frac{174}{203}$

α input = 1.6×10^7

α free = (A) $.77 \times 10^7$ (48%) (B) $.98 \times 10^7$ (60%) (C) $.76 \times 10^7$ (48%)

α inf. centers = 1×10^7 (62%) 1.15×10^7 (72%) 1.7×10^6 (100%)

Conclusions:

The adsorption of α on B is probably not significantly changed by the presence of γ .

The infective centers are as many as the α input in absence of γ . In presence of γ only about 20% or less of α adsorbed show as plaques.

PST Test for α growth in presence of [60^m UV γ]-

150 .05 cc 24^h aerated B $\rightarrow$ 20 cc broth, aerate at 37° = B_I

0 .85 cc B_I + .15 cc [γ UV 60^m] + .02 cc [α :10] = B _{$\alpha\gamma$} , at 37°

5 B _{$\alpha\gamma$} .05 cc $\rightarrow$

	I $\rightarrow$ 20 cc broth	TE 1 cc $\rightarrow$ 19 cc broth	rel. assay
9	1-173 2-175 110 B	10 B	1
14	$\frac{3}{4}$ = confluent (<1000)		5
20		5- ~400 6- ~400	45
32		7- ~400 8- ~400	45

Comment : α grows as usual -

all α input (see other experiments) shows up
as infective centers.

Necessity to repeat the experiment using A as plating strain.

Test for changes in bacteria after action of UV_γ and of [UV_γ + α]

PST

-150 .05 cc 24^h aerated B → 20 cc broth, aerate at 37° = B_I
-3 B_I assay : 2.5 × 10⁴ $\frac{1}{2} = \frac{83}{63}$ [B] = 3.75 × 10⁷

0 .85 cc B_I + .15 cc [γ irr. UV 10^m] = B_γ

18 B_γ : 10³ $\frac{3}{4} = \frac{25}{19}$ (normal colonies).

2 .85 cc B_I + .15 cc [γ irr. UV 10^m] = B_{αγ}

5 .02 cc [α:10] → B_{αγ}

14 B_{αγ} : 10³ $\frac{5}{6} = \frac{41}{24}$ (irregular colonies, in partial lysis).

After 24 h. the tube B_{αγ} shows lysis (almost complete clearing).

Comments.

$$[B]_{\text{input}} \approx 3.2 \times 10^7$$

$$[B]_{\text{colonies in } B_{\gamma}} = 4.4 \times 10^5$$

$$[B]_{\text{colonies in } B_{\alpha\gamma}} = 6.5 \times 10^5$$

Bacteria in presence of UV_γ inact. γ do not reproduce any active φ,
but are, either lysed ~~are~~ inhibited from reproduction.

Moreover, their descendent seem to be still sensitive to α.*

(possibility that even the firstly infected bacteria undergo lysis by α
without reproducing any active phage).

Different possibilities:

- 1) B are lysed by UV_γ. (with or without reproducing active φ)
- 2) B " inhibited " UV_γ
- 3) B " " " and lysed by α (fid.).
- 4) B " " and become carriers of φ UV.

Other possibilities.

* Repeat experiment of α growth in presence of UV_γ, sampling until 300 min.

See 1/21 -

Colonies from plates 3 and 4 ($B + UV\gamma$) passed on agar slant grow normally - Slant kept in ice-box - [$B\gamma$] labeled.

Colonies from plates 5 and 6 ($B + UV\gamma + \alpha$) passed on agar slant give after 24 h only a few colonies of secondary growth, the main area of the slant has undergone [α] lysis. Slant kept in icebox - [$B\gamma$] labeled.

Comment -

Bacteria grown from $B + \gamma$ give descendance which is sensitive to α [even if α has acted upon the first generation (not sure, as only a few bacteria of the first gen. had been attacked by α , see 1/21)]. The resistance to α is probably only of those bacteria which have been attacked by $UV\gamma$. The lysis in the tube $B\gamma$ is probably due to proliferation of the [$UV\gamma$ infected bacteria], elimination of γ and recovery of sensitivity to α .

Test whether [$UV\gamma$ treated B] are still sensitive to γ after division.

Tube $B\alpha\gamma$ 1/21, containing $B + UV\gamma + \alpha$ (clear after 24 h, cloudy after 30 h) carried through ~~the~~ three passages in broth (dilution more than 10^{-4} each passage) labeled as tube I.

Tube $B\gamma$ 1/21, containing $B + UV\gamma$, cloudy after 24 h, carried through three passages in broth (id.), labeled as tube II.

1/26 - Tube I from ~~the~~ ^{third} passage in broth ~~plated~~ inoculated on slant (I), then filtered, and filtrate stored (I filtrate 1/26).

Tube II from 3d passage in broth inoculated on slant (II) then filtered, and filtrate stored (II filtrate 1/26).

1/27 • Test for acquired resistance after the action of $UV\gamma$ and of $UV\gamma + \alpha$.

	plated with $\frac{\alpha}{0}$	$\frac{\gamma}{400}$
$B\alpha\gamma$		
$B\gamma$	250	400
I	0	400
II	250	400

Comment - ~~B~~) Bacteria B, acted upon by $UV\gamma$ for 18 min, then plated, giving a few colonies of bacteria normally sensitive to α and γ (explains why there is lysis in broth with $B + UV\gamma + \alpha$) -

$B\alpha\gamma$) Bacteria B, acted upon by $UV\gamma$ and α ^{10 min}, give then plated, give colonies of bacteria resistant to α and sensit. to γ (why are the first colonies "nibbel"? 1/21). [Because they had not been acted upon by α (not in excess 1/21).]

I) and II) the same characters have the bacteria developed from the same treatment, but passed 3 times in broth.

$UV\gamma$ doesn't produce resistance in bacteria. [Question: ~~have~~ the bacteria which still develop after action of $UV\gamma$ been infected with $UV\gamma$ at all?] The resistance seems to be produced by growth in presence of active phage only.

The few colonies which develop from $B + UV\gamma$, ^{being sensitive to γ ,} seem not to be originated by the ~~few~~ ^{same} bacteria originally present, which were unsensitible to γ and which therefore didn't adsorb it. One must think that the resistance is induced upon a few bacteria by active γ .

Microscopic observation of bacteria B + UVγ

- 150 .05 cc 24^h aer. B → 20 cc broth, aer. at 37° = B_I
- 0 .85 cc B_I + .15 cc [UVγ 10^m], at 37° = B_γ
- 11 .025 cc B_γ plated, covered with coverslide, and observed -
see ~~page~~^{map} on the opposite side.

Comment. Most bacteria, in presence of UVγ in excess, seem to be stopped in their dividing capacity - Some of them (a few percent) undergo a process of clearing and swelling (lysis from without?), some become longer and longer - Exceptionally succeed some bacteria in dividing.